

# Package ‘IDSL.IPA’

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**Type** Package

**Title** Intrinsic Peak Analysis (IPA) for HRMS Data

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**Imports** IDSL.MXP, readxl

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**Description** A multi-layered untargeted pipeline for high-throughput LC/HRMS data processing to extract signals of organic small molecules. The package performs ion pairing, peak detection, peak table alignment, retention time correction, aligned peak table gap filling, peak annotation and visualization of extracted ion chromatograms (EICs) and total ion chromatograms (TICs). The 'IDSL.IPA' package was introduced in <doi:10.1021/acs.jproteome.2c00120> .

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---

*alignedPeakPropertyTableCorrelationListCalculator*  
*Aligned Peak Property Table Correlation List Calculator*

---

### **Description**

Aligned Peak Property Table Correlation List Calculator

### **Usage**

```
alignedPeakPropertyTableCorrelationListCalculator(peakPropertyTable,  
RTtolerance = 0.05, minFreqDetection = 3, minRatioDetection = 0.01,  
method = "pearson", minThresholdCorrelation = 0, number_processing_threads = 1)
```

### **Arguments**

peakPropertyTable  
peak property table such as 'peak\_height', 'peak\_area' and 'peak\_R13C'

RTtolerance retention time tolerance (min)

minFreqDetection  
minimum frequency of detection for a (m/z-RT) peak across the peak property table

minRatioDetection  
minimum ratio of detection for a (m/z-RT) peak across the peak property table. This value should be between (0 - 1).

method  
a character string indicating which correlation coefficient (or covariance) is to be computed. One of "pearson" (default), or "spearman": can be abbreviated. (from 'cor' function of the 'stats' package)

minThresholdCorrelation  
minimum threshold for the correlation method

number\_processing\_threads  
number of processing threads

### **Value**

A list of related peak IDs for each individual (m/z-RT) pair on the peak property table

---

analyteRetentionTimeCorrector  
*analyte retention time corrector*

---

### Description

This function calculates corrected retention times for the peaklists.

### Usage

```
analyteRetentionTimeCorrector(referenceMZRTpeaks, inputPathPeaklist, peaklistFileName,
massAccuracy, RTcorrectionMethod, refPeakTolerance = 1, degreePolynomial = 3)
```

### Arguments

referenceMZRTpeaks  
a matrix of reference peaks for retention time correction.

inputPathPeaklist  
input path to peaklist

peaklistFileName  
file name peaklist

massAccuracy  
mass error to detect common reference peaks.

RTcorrectionMethod  
c('RetentionIndex','Polynomial')

refPeakTolerance  
number of reference peaks for retention time correction using the 'RetentionIndex' method.

degreePolynomial  
polynomial degree for retention time correction using the 'Polynomial' method.

### Value

a list of corrected retention times for each peaklist.

---

chromatogramMatrix     *chromatogram builder for m/z = 263.1678 in 003.d from cord blood sample*

---

### Description

This data illustrates a chromatogram and baseline vectors to indicate chromatogram development.

### Usage

```
data("chromatogramMatrix")
```

**Format**

A data frame with 219 observations on the following 6 variables.

scanNumber a numeric vector  
 retentionTime a numeric vector  
 smoothChromatogram a numeric vector  
 rawChromatogram a numeric vector  
 ‘12C/13C Isotopologue Pairs’ a numeric vector  
 Baseline a numeric vector

**Examples**

```
data(chromatogramMatrix)
```

---

```
chromatographicPeakAnalysis  

Chromatography analysis
```

---

**Description**

This function detects individual chromatographic peaks and measures their peak qualification metrics.

**Usage**

```
chromatographicPeakAnalysis(spectraScanXIC, aggregatedSpectralList, retentionTime,  

  LretentionTime, massAccuracy, mzTarget, rtTarget = NULL, scanNumberStart,  

  scanNumberEnd, smoothingWindow, peakResolvingPower, minNIonPair, minPeakHeight,  

  minRatioIonPair, maxRPW, minSNRbaseline, maxR13CcumulatedIntensity,  

  maxPercentageMissingScans, nSpline, exportEICparameters = NULL)
```

**Arguments**

spectraScanXIC a matrix consists of 5 columns. The column contents are the m/z of 12C isotopologues, intensity of 12C isotopologues, scan number (t), m/z of 13C isotopologues, and intensity of 13C isotopologues, respectively. Redundant scan numbers are not allowed for this module.

aggregatedSpectralList aggregated spectralList and spectra matrix from the ‘IPA\_spectralListAggregator’ module

retentionTime a vector of retention times vs. corresponding scan numbers

LretentionTime length of the retention time vector

massAccuracy mass error to perform chromatography analysis

mzTarget m/z value to perform chromatography analysis

rtTarget	retention time value for a targeted peak to calculate the ancillary chromatography parameters. When this parameter set at 0, the ancillary chromatography parameters are calculated for the entire detected peaks.
scanNumberStart	the first scan number.
scanNumberEnd	the last scan number.
smoothingWindow	number of scans for peak smoothing
peakResolvingPower	a value to represent peak resolving power
minNIonPair	minimum number of nIsoPair for an individual peak
minPeakHeight	minimum peak height for an individual peak
minRatioIonPair	minimum ratio of nIsoPair per number of available scans within an individual peak
maxRPW	maximum allowed value of ratio of peak width at half-height to baseline (RPW) for an individual peak
minSNRbaseline	minimum S/N baseline for an individual peak
maxR13CcumulatedIntensity	maximum allowed value of average R13C for an individual peak
maxPercentageMissingScans	maximum allowed value of percentage missing scans on the raw chromatogram for an individual peak.
nSpline	number of points for further smoothing using a cubic spline smoothing method to calculate ancillary chromatographic parameters
exportEICparameters	When 'NULL', EICs are not plotted. 'exportEICparameters' should contain three variables of 1) an address to save IPA EICs figures, 2) 'HRMS' file name, and 3) a valid string of characters.

**Value**

a data frame consisting of 24 columns representing chromatography and mass spectrometry parameters. Each row represents an individual separated chromatographic peak.

---

chromatographicPeakDetector

*peak detection*

---

**Description**

This function detects separated chromatographic peaks on the chromatogram.

**Usage**

```
chromatographicPeakDetector(int)
```

**Arguments**

int                    a vector of intensities of the chromatogram.

**Value**

A matrix of 2 columns. Each row indicates peak boundary indices on the 'int' vector.

**Examples**

```
data(chromatogramMatrix)
int <- chromatogramMatrix$smoothChromatogram
chromatographicPeakDetector(int)
```

---

derivative5pointsStencil

*Numerical differentiation by five-point stencil method*

---

**Description**

This module performs numerical differentiation using the five-point stencil method.

**Usage**

```
derivative5pointsStencil(x, y, n)
```

**Arguments**

x                    a vector of values for x.  
y                    a vector of values for y.  
n                    order of numerical differentiation (n=1-4).

**Value**

A matrix of 2 columns. The first column represents x and the second column represents numerical differentiation values. This matrix has four rows (two rows from the beginning and 2 rows from the end) less than length of x or y.

**Examples**

```
data(peak_spline)
rt <- peak_spline[, 1]
int <- peak_spline[, 2]
n <- 2 # second order derivative
derivative5pointsStencil(rt, int, n)
```

---

gapFillingCore	<i>Gap-Filling Core Function</i>
----------------	----------------------------------

---

### Description

Gap-Filling Core Function

### Usage

```
gapFillingCore(input_path_hrms, peakXcol, massAccuracy, RTtolerance, scanTolerance,  
retentionTimeCorrectionCheck = FALSE, listCorrectedRTpeaklists = NULL,  
inputPathPeaklist = NULL, ionMassDifference = 1.003354835336,  
number_processing_threads = 1)
```

### Arguments

input_path_hrms	input_path_hrms
peakXcol	peakXcol
massAccuracy	massAccuracy
RTtolerance	RTtolerance
scanTolerance	a scan tolerance to extend the chromatogram for better calculations.
retentionTimeCorrectionCheck	retentionTimeCorrectionCheck
listCorrectedRTpeaklists	listCorrectedRTpeaklists
inputPathPeaklist	inputPathPeaklist
ionMassDifference	ionMassDifference
number_processing_threads	number of processing threads

### Value

A list of gap-filled data



---

IPA_aggregate	<i>aggregation method for the IDSL.IPA modules</i>
---------------	--

---

**Description**

This module is to optimize the 'indexVec' variable by removing elements that have redundant 'idVec' numbers.

**Usage**

```
IPA_aggregate(idVec, variableVec, indexVec, targetVar)
```

**Arguments**

idVec	a vector of id numbers. Repeated id numbers are allowed.
variableVec	a vector of variable of the interest such as RT, m/z, etc.
indexVec	a vector of indices
targetVar	the targeted value in 'variableVec'

**Value**

a clean indexVec after removing repeated 'idVec'.

---

IPA_baselineDeveloper	<i>Develop a baseline for the chromatogram using local minima</i>
-----------------------	---

---

**Description**

This function generates a vector of baselines for the chromatogram using local minima. It also is capable of excluding outlier local minima to generate a realistic baseline including true baseline regions. This baseline may represent the local noise levels for the chromatogram.

**Usage**

```
IPA_baselineDeveloper(segment, int)
```

**Arguments**

segment	a matrix or a vector of adjusted scan number of local minima w/ or w/o redundant local minima. Adjusted scan numbers are the scan numbers but adjusted to start at 1.
int	a vector of intensities of the chromatogram.

**Value**

A vector of baselines in the same size of the "int" vector.

**Examples**

```
data(segment)
data(chromatogramMatrix)
int <- chromatogramMatrix$smoothChromatogram
IPA_baselineDeveloper(segment, int)
```

---

IPA\_CompoundsAnnotation

*Compound-centric peak annotation*

---

**Description**

This function performs compound-centric peak annotation.

**Usage**

```
IPA_CompoundsAnnotation(PARAM)
```

**Arguments**

PARAM            a data frame from IPA\_xlsxAnalyzer function containing the IPA parameters.

**Value**

This function saves individual .csv files for each compound in the "compound\_centric\_annotation" folder.

---

IPA\_GapFiller

*IPA GapFiller*

---

**Description**

This function fills the gaps on the peak table.

**Usage**

```
IPA_GapFiller(PARAM)
```

**Arguments**

PARAM            a data frame from the 'IPA\_xlsxAnalyzer' function containing the IPA parameters.

**Value**

This function saves individual .csv and .Rdata files for the gap-filled peak tables for peak height, area, and R13C properties in the "peak\_alignment" folder.

---

IPA_IonPairing	<i>IPA Ion Pairing</i>
----------------	------------------------

---

**Description**

This function pairs two ions with a fixed distance in high-resolution mass spectral datasets

**Usage**

```
IPA_IonPairing(spectraList, minSpectraNoiseLevel, massAccuracyIonPair = 0.015,  
ionMassDifference = 1.003354835336, number_processing_threads = 1)
```

**Arguments**

`spectraList` list of mass spectra in each chromatogram scan

`minSpectraNoiseLevel` intensity threshold at each chromatogram scan

`massAccuracyIonPair` mass error to detect pair ions

`ionMassDifference` mass difference to pair ions. (Default =  $\Delta C = 13C - 12C = 1.003354835336$ , or  $\Delta S = 34S - 32S = 1.9957958356$ , or any numerical value.)

`number_processing_threads` number of processing threads

**Value**

A matrix consists of 5 columns. The column contents are the m/z of 12C isotopologues, intensity of 12C isotopologues, scan number (t), m/z of 13C isotopologues, and intensity of 13C isotopologues, respectively.

---

IPA_logRecorder	<i>IPA_logRecorder</i>
-----------------	------------------------

---

**Description**

IPA\_logRecorder

**Usage**

```
IPA_logRecorder(messageQuote, allowedPrinting = TRUE)
```

**Arguments**

messageQuote	messageQuote
allowedPrinting	allowedPrinting

**Value**

a line of communication messages is exported to the console and the log .txt file.

---

IPA_message	<i>IPA message</i>
-------------	--------------------

---

**Description**

IPA\_message

**Usage**

```
IPA_message(messageQuote, failedMessage= TRUE)
```

**Arguments**

messageQuote	messageQuote
failedMessage	failedMessage

**Value**

a line of communication messages is exported to the console.

---

IPA_MSdeconvoluter	<i>MS deconvoluter</i>
--------------------	------------------------

---

**Description**

This function deconvolutes mass spectrometry files into a list of mass spectra and a vector of retention times.

**Usage**

```
IPA_MSdeconvoluter(inputHRMSfolderPath, MSfileName, MSlevel = 1)
```

**Arguments**

inputHRMSfolderPath	address of the mass spectrometry file
MSfileName	mass spectrometry file.
MSlevel	MS level to extract information.

**Value**

spectralList	a list of mass spectra.
retentionTime	a vector of retention times for scan numbers.
MS_polarity	mass spectrometry ionization mode (+/-)

---

IPA_PeakAlignment	<i>IPA peak alignment</i>
-------------------	---------------------------

---

**Description**

This function produces an aligned peak table from individual peaklists.

**Usage**

```
IPA_PeakAlignment(PARAM)
```

**Arguments**

PARAM	a data frame from the 'IPA_xlsxAnalyzer' function.
-------	--

**Value**

This function saves individual .csv and .Rdata files for the aligned peak tables for peak height, area, and R13C properties in the "peak\_alignment" folder.

IPA\_PeakAnalyzer      *IPA Peak Analyzer*

---

**Description**

This function performs the IPA peak detection module.

**Usage**

```
IPA_PeakAnalyzer(PARAM)
```

**Arguments**

PARAM                  is a data frame from IPA\_xlsxAnalyzer function.

**Value**

This function saves individual peaklist files in ‘.csv’ and ‘.Rdata’ formats for HRMS files in the ‘peaklists’ folder.

---

IPA\_PeaklistAnnotation  
*IPA Peaklist Annotation*

---

**Description**

This function performs sample-centric peak annotation.

**Usage**

```
IPA_PeaklistAnnotation(PARAM)
```

**Arguments**

PARAM                  a data frame from IPA\_xlsxAnalyzer function.

**Value**

This function saves individual .csv files for peak height, area, and R13C properties in the "sample\_centric\_annotation" folder.

---

IPA\_peak\_alignment\_folder\_xlsxAnalyzer  
*IPA peak alignment folder xlsxAnalyzer*

---

**Description**

IPA peak alignment folder xlsxAnalyzer

**Usage**

IPA\_peak\_alignment\_folder\_xlsxAnalyzer(PARAM, PARAM\_ID, checkpoint\_parameter, correctedRTcheck = FALSE, CSAcheck = FALSE, allowedVerbose = TRUE)

**Arguments**

PARAM	PARAM
PARAM_ID	PARAM_ID
checkpoint_parameter	checkpoint_parameter
correctedRTcheck	correctedRTcheck
CSAcheck	CSAcheck
allowedVerbose	c(TRUE, FALSE). A 'TRUE' allowedVerbose provides messages about the flow of the function.

**Value**

PARAM	PARAM
checkpoint_parameter	checkpoint_parameter

---

IPA\_spectralListAggregator  
*spectralList filtering*

---

**Description**

This module stacks the spectralList object and creates a list of ions for a rapid spectra query.

**Usage**

IPA\_spectralListAggregator(spectralList)

**Arguments**

spectraList      a list of mass spectra in each chromatogram scan.

**Value**

aggregatedSpectraList  
                                  aggregated spectraList  
 spectralListMatrix  
                                  matrix of row bounded spectraList

---

IPA\_targeted                    *IPA Targeted Analysis*

---

**Description**

This function plots extracted ion chromatogram (EIC) figures in the targeted mode.

**Usage**

```
IPA_targeted(PARAM_targeted, allowedVerbose = TRUE)
```

**Arguments**

PARAM\_targeted    IPA parameters to feed the 'IPA\_targeted' module. This variable can be produced using the 'IPA\_targeted\_xlsxAnalyzer' module.

allowedVerbose    c(TRUE, FALSE). A 'TRUE' allowedVerbose provides messages about the flow of the function.

**Value**

This module saves extracted ion chromatograms (EICs) in .png format in the "Targeted\_EICs" folder and saves a table of peak properties.

---

IPA\_targeted\_xlsxAnalyzer  
                                  *IPA Targeted xlsxAnalyzer*

---

**Description**

This function processes the spreadsheet of the IPA parameters to ensure the parameter inputs are in agreement with the 'IPA\_targeted' function.

**Usage**

```
IPA_targeted_xlsxAnalyzer(spreadsheet)
```



**Arguments**

spreadsheet      contains the IPA parameters.

**Value**

‘PARAM\_targeted‘ which is the IPA parameters to feed the ‘IPA\_targeted‘ function.

**Examples**

```
## To generate the IPA spreadsheet parameters
s_path <- system.file("extdata", package = "IDSL.IPA")
SSh1 <- paste0(s_path, "/IPA_parameters.xlsx")
spreadsheet <- readxl::read_xlsx(SSh1, sheet = 'IPA_targeted')
PARAM_targeted = cbind(spreadsheet[, 2], spreadsheet[, 4])
temp_wd <- tempdir()
temp_wd_zip <- paste0(temp_wd, "/idsl_ipa_test_files.zip")
tryCatch({download.file(paste0("https://github.com/idslme/IDSL.IPA/blob/main/",
                             "IPA_educational_files/idsl_ipa_test_files.zip?raw=true"),
                    destfile = temp_wd_zip)
  unzip(temp_wd_zip, exdir = temp_wd)
  pass_download <- TRUE},
  error = function(e) {pass_download <- FALSE},
  warning = function(w) {pass_download <- FALSE})
if (pass_download) {
  PARAM_targeted[3, 2] <- temp_wd
  PARAM_targeted[7, 2] <- temp_wd
  PARAM_targeted[8, 2] <- "53.01853, 61.00759"
  PARAM_targeted[9, 2] <- "0.951, 0.961"
  ##
  PARAM_targeted <- IPA_targeted_xlsxAnalyzer(PARAM_targeted)
}
```

---

 IPA\_workflow

*IPA Workflow*


---

**Description**

This function executes the IPA workflow in order.

**Usage**

```
IPA_workflow(spreadsheet)
IPA_Workflow(spreadsheet)
```

**Arguments**

spreadsheet      IPA spreadsheet

**Value**

This function organizes the IPA file processing for a better performance using the template spreadsheet.

**Examples**

```
s_path <- system.file("extdata", package = "IDSL.IPA")
SSh1 <- paste0(s_path, "/IPA_parameters.xlsx")
## To see the results, use a known folder instead of the `tempdir()` command
temp_wd <- tempdir()
temp_wd_zip <- paste0(temp_wd, "/idsl_ipa_test_files.zip")
spreadsheet <- readxl::read_xlsx(SSh1)
PARAM = cbind(spreadsheet[, 2], spreadsheet[, 4])
tryCatch({download.file(paste0("https://github.com/idslme/IDSL.IPA/blob/main/",
                             "IPA_educational_files/idsl_ipa_test_files.zip?raw=true"),
                    destfile = temp_wd_zip)
  unzip(temp_wd_zip, exdir = temp_wd)
  pass_download <- TRUE},
  error = function(e) {pass_download <- FALSE},
  warning = function(w) {pass_download <- FALSE})
if (pass_download) {
  PARAM[7, 2] <- temp_wd
  PARAM[44, 2] <- s_path
  PARAM[10, 2] <- temp_wd
  IPA_workflow(PARAM)
}
```

---

IPA\_xlsxAnalyzer

*IPA xlsx Analyzer*

---

**Description**

This function processes the spreadsheet of the IPA parameters to ensure the parameter inputs are in agreement with the IPA requirements.

**Usage**

```
IPA_xlsxAnalyzer(spreadsheet)
```

**Arguments**

spreadsheet      IPA spreadsheet

**Value**

This function returns the IPA parameters to feed the `IPA_Workflow`, `IPA_CompoundsAnnotation`, `IPA_GapFiller`, `IPA_PeakAlignment`, `IPA_PeakAnalyzer`, and `IPA_PeaklistAnnotation` functions.

**Examples**

```

s_path <- system.file("extdata", package = "IDSL.IPA")
SSh1 <- paste0(s_path, "/IPA_parameters.xlsx")
temp_wd <- tempdir()
temp_wd_zip <- paste0(temp_wd, "/idsl_ipa_test_files.zip")
spreadsheet <- readxl::read_xlsx(SSh1)
PARAM <- cbind(spreadsheet[, 2], spreadsheet[, 4])
tryCatch({download.file(paste0("https://github.com/idslme/IDSL.IPA/blob/main/",
                             "IPA_educational_files/idsl_ipa_test_files.zip?raw=true"),
                    destfile = temp_wd_zip)
unzip(temp_wd_zip, exdir = temp_wd)
pass_download <- TRUE},
error = function(e) {pass_download <- FALSE},
warning = function(w) {pass_download <- FALSE})
if (pass_download) {
  PARAM[7, 2] <- temp_wd
  PARAM[10, 2] <- temp_wd # output data location
  PARAM[44, 2] <- s_path # reference file location
  PARAM <- IDSL.IPA::IPA_xlsxAnalyzer(PARAM)
}

```

---

islocalminimum

*islocalminimum*


---

**Description**

This function returns indices of local minimum points on a curve.

**Usage**

```
islocalminimum(y)
```

**Arguments**

*y* is a vector of *y* values.

**Value**

A vector in the same size of the vector 'y'. Local minimum arrays represented by -1.

**Examples**

```

data(chromatogramMatrix)
int <- chromatogramMatrix$smoothChromatogram
islocalminimum(int)

```

---

islocaloptimum	<i>islocaloptimum</i>
----------------	-----------------------

---

**Description**

This function returns indices of local minimum and maximum points on a curve.

**Usage**

```
islocaloptimum(y)
```

**Arguments**

`y` is a vector of y values.

**Value**

A vector in the same size of the vector 'y'. Local minimum and maximum arrays represented by -1 and +1, respectively.

**Examples**

```
data(chromatogramMatrix)
int <- chromatogramMatrix$smoothChromatogram
islocaloptimum(int)
```

---

loadRdata	<i>loadRdata</i>
-----------	------------------

---

**Description**

This function loads .Rdata files into a variable.

**Usage**

```
loadRdata(fileName)
```

**Arguments**

`fileName` is an '.Rdata' file.

**Value**

The called variable into the new assigned variable name.

---

mzClusteringRawXIC	<i>m/z clustering raw XIC</i>
--------------------	-------------------------------

---

**Description**

This function clusters related 12C m/z values.

**Usage**

```
mzClusteringRawXIC(spectraScan123, massAccuracy, minNIonPair, minPeakHeightXIC)
```

**Arguments**

spectraScan123	a matrix consists of 3 columns. The column contents are the m/z of 12C isotopologues, intensity of 12C isotopologues, and scan number (t).
massAccuracy	mass accuracy to detect related 12C m/z values.
minNIonPair	minimum number of nIsoPair for an individual peak.
minPeakHeightXIC	minimum peak height for an individual raw EIC

**Value**

This function returns an list on index numbers of EICs for the "spectraScan" variable.

---

mzRTindexer	<i>m/z - RT Indexer</i>
-------------	-------------------------

---

**Description**

This function locate the closest pair of a reference (m/z - RT) pair in a 2-D grid of 'm/z' and 'RT' vectors.

**Usage**

```
mzRTindexer(MZvec, RTvec, MZref, RTref, massAccuracy, RTtolerance)
```

**Arguments**

MZvec	m/z vector
RTvec	RT vector
MZref	a reference m/z
RTref	a reference RT
massAccuracy	m/z tolerance
RTtolerance	RT tolerance

**Value**

index of closest pair to the reference (m/z - RT) pair

**Note**

This function returns NULL in case no match is detected.

---

peakAlignmentCore      *Peak Alignment Core*

---

**Description**

This function aligns peaks from multiple peaklists and produces an aligned table of common peaks among multiple samples.

**Usage**

```
peakAlignmentCore(peaklistInputFolderPath, peaklistFileNames, listCorrectedRTpeaklists,  
massAccuracy, RTtolerance, number_processing_threads = 1)
```

**Arguments**

peaklistInputFolderPath      path to directory of peaklists.  
peaklistFileNames            name of peaklists for peak table production.  
listCorrectedRTpeaklists      a list of corrected or uncorrected retention times for each peaklist.  
massAccuracy                  mass error to detect common peaks.  
RTtolerance                   retention time tolerance to detect common peaks.  
number\_processing\_threads      number of processing threads

**Value**

This function returns an aligned peak table with index numbers from individual peaklists for each peak.

---

peakAreaCalculator     *peak area*

---

**Description**

This function calculates area under the curve using a trapezoid method.

**Usage**

```
peakAreaCalculator(x, y)
```

**Arguments**

x                    is a vector of x values.  
y                    is a vector of y values.

**Value**

A number for the integrated peak area.

**Examples**

```
data("peak_spline")  
rt <- peak_spline[, 1]  
int <- peak_spline[, 2]  
peakAreaCalculator(rt, int)
```

---

peakAsymmetryFactorCalculator

*Asymmetry factor for a chromatographic peak*

---

**Description**

This function calculates an asymmetry factor for a chromatographic peak.

**Usage**

```
peakAsymmetryFactorCalculator(rt, int)
```

**Arguments**

rt                    a vector of retention times for the chromatographic peak.  
int                   a vector of intensities corresponding to the vector of retention times for the chromatographic peak.

**Value**

asymmetry of the chromatographic peak. 1 is for very symmetric peak.

**Examples**

```
data(peak_spline)
rt <- peak_spline[, 1]
int <- peak_spline[, 2] - peak_spline[, 3]
peakAsymmetryFactorCalculator(rt, int)
```

---

peakDerivativeSkewnessCalculator

*Peak Derivative Skewness Calculator*

---

**Description**

This function calculates skewness of a chromatographic peak using first order degree of numerical differentiation.

**Usage**

```
peakDerivativeSkewnessCalculator(rt, int)
```

**Arguments**

**rt** a vector representing retention times of the chromatographic peak.  
**int** a vector representing intensities of the chromatographic peak.

**Value**

Skewness of a chromatographic peak. 1 is for very symmetric peak. Minimum is 0 from this function.

**Examples**

```
data(peak_spline)
rt <- peak_spline[, 1]
int <- peak_spline[, 2]
peakDerivativeSkewnessCalculator(rt, int)
```



---

`peakFrontingTailingResolver`*Fronting and tailing peaks resolver*

---

**Description**

This function attempts to resolve peak tailings or frontings into the main peak in case they were detected as separate peaks.

**Usage**

```
peakFrontingTailingResolver(segment, int, maxScanDifference, peakResolvingPower = 0.025)
```

**Arguments**

<code>segment</code>	a matrix or a vector of peak boundaries.
<code>int</code>	a vector of intensities of the entire chromatogram.
<code>maxScanDifference</code>	maximum scan number difference between peak tailing or fronting and the main peak.
<code>peakResolvingPower</code>	power of peak resolving tool.

**Value**

A matrix of 2 columns. Each row indicates peak boundary indices on the 'int' vector after resolving fronting and tailing peaks.

**Examples**

```
data(segment)
data(chromatogramMatrix)
int <- chromatogramMatrix$smoothChromatogram
maxScanDifference <- 7
peakResolvingPower <- 0.2
peakFrontingTailingResolver(segment, int, maxScanDifference, peakResolvingPower)
```

peakGaussianityCalculator

*Peak Gaussianity Calculator*

---

### Description

This module measures gaussianity of chromatographic peak using Pearson correlation coefficients ( $\rho$ ) at top 80 percent of peak.

### Usage

```
peakGaussianityCalculator(RT, Int, BL, gauge = 0.8)
```

### Arguments

RT	a vector of retention times of the chromatographic peak.
Int	a vector of intensities of the chromatographic peak.
BL	a vector of baseline of the chromatographic peak.
gauge	represents the gauge height of peak for Gaussianity measurement.

### Value

Gaussianity of the chromatographic peak.

### Examples

```
data("peak_spline")
RT <- peak_spline[, 1]
Int <- peak_spline[, 2]
BL <- peak_spline[, 3]
peakGaussianityCalculator(RT, Int, BL, gauge = 0.8)
```

---

peakPropertyTableFreqCalculator

*Peak Property Table Frequency Calculator*

---

### Description

Peak Property Table Frequency Calculator

### Usage

```
peakPropertyTableFreqCalculator(peakPropertyTable, startColumnIndex = 3,
number_processing_threads = 1, allowedVerbose = TRUE)
```

**Arguments**

peakPropertyTable  
                                  peakPropertyTable  
startColumnIndex  
                                  startColumnIndex  
number\_processing\_threads  
                                  number\_processing\_threads  
allowedVerbose c(TRUE, FALSE). A 'TRUE' allowedVerbose provides messages about the flow of the function.

**Value**

a vector of frequency of detection.

---

peakPropertyTableMedianCalculator  
*Peak Property Table Median Calculator*

---

**Description**

Peak Property Table Median Calculator

**Usage**

```
peakPropertyTableMedianCalculator(peakPropertyTable, falggingVector = NULL,  
number_processing_threads = 1, allowedVerbose = TRUE)
```

**Arguments**

peakPropertyTable  
                                  peakPropertyTable  
falggingVector falggingVector  
number\_processing\_threads  
                                  number\_processing\_threads  
allowedVerbose c(TRUE, FALSE). A 'TRUE' allowedVerbose provides messages about the flow of the function.

**Value**

updated peak property table

---

`peakPseudomomentsSymmetryCalculator`*Peak Pseudomoments Symmetry Calculator*

---

**Description**

This function measures peak symmetry and skewness using the inflection points of the peak on both sides.

**Usage**

```
peakPseudomomentsSymmetryCalculator(rt, int)
```

**Arguments**

<code>rt</code>	a vector of retention times for the chromatographic peak.
<code>int</code>	a vector of intensities corresponding to the vector of retention times for the chromatographic peak.

**Value**

PeakSymmetry	peak symmetry for the chromatographic peak.
Skewness	skewness for the chromatographic peak.

**Examples**

```
data("peak_spline")
rt <- peak_spline[, 1]
int <- peak_spline[, 2] - peak_spline[, 3]
peakPseudomomentsSymmetryCalculator(rt, int)
```

---

`peakSharpnessCalculator`*Peak Sharpness Calculator*

---

**Description**

This function measures sharpness of a chromatographic peak

**Usage**

```
peakSharpnessCalculator(int)
```

**Arguments**

<code>int</code>	a vector of intensities of the chromatographic peak.
------------------	--

**Value**

A number representing peak sharpness. The higher values indicate higher sharpness.

**Examples**

```
data("peak_spline")
int <- peak_spline[, 2]
peakSharpnessCalculator(int)
```

---

peakUSPtailingFactorCalculator  
*Peak USP Tailing Factor Calculator*

---

**Description**

This function calculates USP tailing factor at above 10 percent of the height.

**Usage**

```
peakUSPtailingFactorCalculator(rt, int)
```

**Arguments**

**rt** a vector of retention times for the chromatographic peak.  
**int** a vector of intensities corresponding to the vector of retention times for the chromatographic peak.

**Value**

USP tailing factor for the chromatographic peak.

**Examples**

```
data(peak_spline)
rt <- peak_spline[, 1]
int <- peak_spline[, 2] - peak_spline[, 3]
peakUSPtailingFactorCalculator(rt, int)
```

---

peakWidthCalculator     *peak width measuement*

---

**Description**

This function measures peak width at different peak heights.

**Usage**

```
peakWidthCalculator(rt, int, gauge)
```

**Arguments**

**rt**                    a vector of retention times of the chromatographic peak.  
**int**                    a vector of intensities of the chromatographic peak.  
**gauge**                 a height gauge to measure the peak width. This parameter should be between 0-1.

**Value**

A peak width at the guaged height.

**Examples**

```
data("peak_spline")  
rt <- peak_spline[, 1]  
int <- peak_spline[, 2] - peak_spline[, 3]  
gauge <- 0.5  
peakWidthCalculator(rt, int, gauge)
```

---

peakXcolFiller             *Peak table producer*

---

**Description**

This function fills the peak table from individual peaklists.

**Usage**

```
peakXcolFiller(peakXcol, inputPathPeaklist)
```

**Arguments**

**peakXcol**                a matrix of index numbers in individual peaklists for each peak (m/z-RT).  
**inputPathPeaklist**     address of the peaklists.

**Value**

peak_height	peak table for height values
peak_area	peak table for area values
peak_R13C	peak table for R13C values

---

peakXcolFlagger	<i>PeakXcol Flagger</i>
-----------------	-------------------------

---

**Description**

PeakXcol Flagger

**Usage**

```
peakXcolFlagger(mzPeakXcol, rtPeakXcol, freqPeakXcol, massAccuracy, RTtolerance,
maxRedundantPeakFlagging)
```

**Arguments**

mzPeakXcol	mzPeakXcol
rtPeakXcol	rtPeakXcol
freqPeakXcol	freqPeakXcol
massAccuracy	massAccuracy
RTtolerance	RTtolerance
maxRedundantPeakFlagging	maxRedundantPeakFlagging

**Value**

a vector with flagged numbers

---

peak_spline	<i>peak spline</i>
-------------	--------------------

---

**Description**

illustrates a smoothe peak using cubic spline smoothing method

**Usage**

```
data("peak_spline")
```

**Format**

A data frame with 100 observations on the following 3 variables.

rt\_spline a numeric vector

int\_spline a numeric vector

bl\_approx a numeric vector

**Examples**

```
data(peak_spline)
```

---

plot\_mz\_eic

*plot\_mz\_eic*

---

**Description**

plot\_mz\_eic

**Usage**

```
plot_mz_eic(filelist, filelocation, mzTarget, massAccuracy,  
number_processing_threads = 1, rtstart = 0, rtend = 0, plotTitle = "")
```

**Arguments**

filelist	filelist
filelocation	filelocation
mzTarget	mzTarget
massAccuracy	massAccuracy
number_processing_threads	number of processing threads
rtstart	rtstart
rtend	rtend
plotTitle	plotTitle

**Value**

plot\_mz\_eic



---

plot_simple_tic	<i>plot_simple_tic</i>
-----------------	------------------------

---

**Description**

plot\_simple\_tic

**Usage**

```
plot_simple_tic(filelist, filelocation, number_processing_threads = 1,
plotTitle = "Total Ion Chromatogram")
```

**Arguments**

filelist	filelist
filelocation	filelocation
number_processing_threads	number of processing threads
plotTitle	plotTitle

**Value**

plot\_simple\_tic

---

primaryXICdeconvoluter	<i>Primary peak analyzer</i>
------------------------	------------------------------

---

**Description**

This function performs the first round of the chromatography analysis.

**Usage**

```
primaryXICdeconvoluter(spectraScan, scanTolerance, indexXIC, aggregatedSpectralList,
retentionTime, massAccuracy, smoothingWindow, peakResolvingPower, minNIonPair,
minPeakHeight, minRatioIonPair, maxRPW, minSNRbaseline, maxR13CcumulatedIntensity,
maxPercentageMissingScans, nSpline, exportEICparameters = NULL,
number_processing_threads = 1)
```

**Arguments**

spectraScan	a matrix consists of 5 columns. The column contents are the m/z of 12C isotopologues, intensity of 12C isotopologues, scan number (t), m/z of 13C isotopologues, and intensity of 13C isotopologues.
scanTolerance	a scan tolerance to extend the chromatogram for better calculations.
indexXIC	a list of indices of candidate 12C m/z values from spectraScan matrix.
aggregatedSpectralList	aggregated spectraList and spectra matrix from the 'IPA_spectraListAggregator' module
retentionTime	a vector of retention times vs. corresponding scan numbers.
massAccuracy	a m/z value to perform chromatography analysis.
smoothingWindow	number of scans for peak smoothing.
peakResolvingPower	a value to represent peak resolving power.
minNIonPair	minimum number of nIsoPair for an individual peak.
minPeakHeight	minimum peak height for an individual peak.
minRatioIonPair	minimum ratio of nIsoPair per number of available scans within an individual peak.
maxRPW	maximum allowed value of ratio of peak width at half-height to baseline (RPW) for an individual peak.
minSNRbaseline	minimum S/N baseline for an individual peak.
maxR13CcumulatedIntensity	maximum allowed value of average R13C for an individual peak.
maxPercentageMissingScans	maximum allowed value of percentage missing scans on the raw chromatogram for an individual peak.
nSpline	number of points for further smoothing using a cubic spline smoothing method.
exportEICparameters	When 'NULL', EICs are not plotted. 'exportEICparameters' should contain three variables of 1) an address to save IPA EICs figures, 2) 'HRMS' file name, and 3) a valid string of characters.
number_processing_threads	number of processing threads

**Value**

a data frame consisting of 24 columns representing chromatography and mass spectrometry parameters. Each row represents an individual separated chromatographic peak.

---

```
recursiveMZpeaklistCorrector
    recursive mass correction
```

---

## Description

This function performs recursive mass correction.

## Usage

```
recursiveMZpeaklistCorrector(peaklist, spectraScan, scanTolerance,
    aggregatedSpectralList, retentionTime, massAccuracy, smoothingWindow,
    peakResolvingPower, minNIonPair, minPeakHeight, minRatioIonPair, maxRPW,
    minSNRbaseline, maxR13CcumulatedIntensity, maxPercentageMissingScans, nSpline,
    exportEICparameters = NULL, number_processing_threads = 1)
```

## Arguments

peaklist	an IPA peaklist from 'primaryXICdeconvoluter' function.
spectraScan	a matrix consists of 5 columns. The column contents are the m/z of 12C isotopologues, intensity of 12C isotopologues, scan number (t), m/z of 13C isotopologues, and intensity of 13C isotopologues.
scanTolerance	a scan tolerance to extend the chromatogram for better calculations.
aggregatedSpectralList	aggregated spectralList and spectra matrix from the 'IPA_spectralListAggregator' module
retentionTime	a vector of retention times for corresponding scan numbers.
massAccuracy	an m/z value to perform chromatography analysis.
smoothingWindow	a number of scans for peak smoothing.
peakResolvingPower	a value to represent peak resolving power.
minNIonPair	minimum number of nIsoPair for an individual peak.
minPeakHeight	minimum peak height for an individual peak.
minRatioIonPair	minimum ratio of nIsoPair per number of available scans within an individual peak.
maxRPW	maximum allowed value of ratio of peak width at half-height to baseline (RPW) for an individual peak.
minSNRbaseline	minimum S/N baseline for an individual peak.
maxR13CcumulatedIntensity	maximum allowed value of average R13C for an individual peak.

maxPercentageMissingScans	maximum allowed value of percentage missing scans on the raw chromatogram for an individual peak.
nSpline	number of points for further smoothing using a cubic spline smoothing method to calculate ancillary chromatographic parameters.
exportEICparameters	When 'NULL', EICs are not plotted. 'exportEICparameters' should contain three variables of 1) an address to save IPA EICs figures, 2) 'HRMS' file name, and 3) a valid string of characters.
number_processing_threads	number of processing threads

**Value**

a dataframe consisting of 24 columns representing chromatography and mass spectrometry parameters. Each row represents an individual separated chromatographic peak.

---

referenceRetentionTimeDetector  
*Reference retention time detector*

---

**Description**

This module detects recurring reference peaks (m/z-RT) for retention time correction.

**Usage**

```
referenceRetentionTimeDetector(inputPathPeaklist, refPeaklistFileNames,
minFrequencyRefPeaks, massAccuracy, RTtolerance, number_processing_threads = 1)
```

**Arguments**

inputPathPeaklist	path to directory of peaklists.
refPeaklistFileNames	name of peaklists files to detect recurring reference peaks (m/z-RT).
minFrequencyRefPeaks	minimum frequency of the recurring reference peaks (m/z-RT) in the reference files.
massAccuracy	mass error to detect common peaks.
RTtolerance	retention time tolerance to detect common peaks.
number_processing_threads	number of processing threads

**Value**

referenceMZRTpeaks  
a matrix of two columns of m/z and RT of common peaks in the reference samples.

listRefRT  
a list of corrected or uncorrected retention times for each peaklist.

---

segment	<i>segment</i>
---------	----------------

---

**Description**

This data illustrates an output matrix of chromatogram peak detection module from the "chromatogramMatrix.rda" object.

**Usage**

```
data("segment")
```

**Format**

The format is: num [1:16, 1:2] 7 15 23 33 38 46 67 86 102 118 ...

**Examples**

```
data(segment)
```

---

SNRbaseline	<i>SNR baseline</i>
-------------	---------------------

---

**Description**

This function calculates S/N using local noise levels from baseline,

**Usage**

```
SNRbaseline(int, baseline)
```

**Arguments**

int  
a vector of intensities corresponding to the vector of retention times for the chromatographic peak.

baseline  
a vector of baseline of the chromatographic peak.

**Value**

S/N value

## Examples

```
data("peak_spline")
int <- peak_spline[, 2]
baseline <- peak_spline[, 3]
SNRbaseline(int, baseline)
```

---

SNRrms

*SNR RMS*

---

## Description

This function calculates signal-to-noise ratio using root mean square.

## Usage

```
SNRrms(int, baseline, gauge = 0.80)
```

## Arguments

<code>int</code>	is the vector of intensities corresponding to the vector of retention times for the chromatographic peak.
<code>baseline</code>	is a vector of baseline of the chromatographic peak.
<code>gauge</code>	represents the gauge height of peak for gaussianity measurement.

## Value

S/N value

## Examples

```
data("peak_spline")
int <- peak_spline[, 2]
baseline <- peak_spline[, 3]
SNRrms(int, baseline)
```

---

 SNRxcms

*SNR xcms*


---

**Description**

This function calculates S/N values using a method suggested in the xcms paper (Tautenhahn, 2008).

**Usage**

```
SNRxcms(int)
```

**Arguments**

`int` a vector of intensities corresponding to the vector of retention times for the chromatographic peak.

**Value**

S/N value

**References**

Tautenhahn, R., Böttcher, C. and Neumann, S. (2008). Highly sensitive feature detection for high resolution LC/MS. *BMC bioinformatics*, 9(1), 1-16, [doi:10.1186/147121059504](https://doi.org/10.1186/147121059504).

**Examples**

```
data(peak_spline)
int <- peak_spline[, 2]
SNRxcms(int)
```

---

 targetedIonPairing

*Targeted Ion Pairing*


---

**Description**

This module only pairs ‘mzTarget’ values across ‘scanNumberStart’ through ‘scanNumberEnd’ scan numbers.

**Usage**

```
targetedIonPairing(spectralList, scanNumberStart, scanNumberEnd, mzTarget,
  massAccuracy, ionMassDifference = 1.003354835336, massAccuracyIonPair = massAccuracy*1.5)
```

**Arguments**

spectraList	spectraList which is a list of mass spectra
scanNumberStart	the first scan number.
scanNumberEnd	the last scan number.
mzTarget	m/z value to perform chromatography analysis
massAccuracy	mass accuracy to select the dominant ion
ionMassDifference	mass difference to pair ions. (Default = $\Delta C = 13C - 12C = 1.003354835336$ ), or $\Delta S = 34S - 32S = 1.9957958356$ , or any numerical value.
massAccuracyIonPair	mass accuracy to select the second ion

**Value**

A targeted ion paired spectra and their scan numbers

---

XIC	<i>XIC</i>
-----	------------

---

**Description**

XIC

**Usage**

XIC(aggregatedSpectraList, scanNumberStart, scanNumberEnd, mzTarget, massAccuracy)

**Arguments**

aggregatedSpectraList	aggregated spectraList and spectra matrix from the 'IPA_spectraListAggregator' module
scanNumberStart	the first scan number.
scanNumberEnd	the last scan number.
mzTarget	an m/z value to perform XIC analysis.
massAccuracy	a mass error to perform XIC analysis.

**Value**

A matrix of three columns representing scan number, m/z, and intensity.



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