

Package ‘reportRmd’

January 24, 2025

Title Tidy Presentation of Clinical Reporting

Version 0.1.1

Date 2024-12-30

Description Streamlined statistical reporting in 'Rmarkdown' environments. Facilitates the automated reporting of descriptive statistics, multiple univariate models, multivariable models and tables combining these outputs. Plotting functions include customisable survival curves, forest plots from logistic and ordinal regression and bivariate comparison plots.

License MIT + file LICENSE

Suggests nlme, tidycmprsk, rmarkdown, testthat (>= 3.0.0)

Config/testthat/edition 3

Encoding UTF-8

RoxygenNote 7.3.2

Imports aod, boot, cmprsk, cowplot, dplyr, geepack, ggplot2, ggpubr, gridExtra, kableExtra, knitr, lifecycle, MASS, pander, plyr, rlang, rstatix, scales, stats, survival, tidyselect

Collate 'helper.R' 'main.R' 'globals.R' 'data.R' 'lblCode.R' 'rm_compactsum.R' 'rm_uvsum2.R' 'autoreg.R' 'autosum.R' 'getVarLevels.R' 'rm_mvsum2.R'

Depends R (>= 4.2)

LazyData no

VignetteBuilder knitr, rmarkdown

NeedsCompilation no

Author Lisa Avery [cre, aut] (<<https://orcid.org/0000-0002-8431-5143>>),
Ryan Del Bel [aut],
Osvaldo Espin-Garcia [aut],
Katherine Lajkosz [aut] (<<https://orcid.org/0000-0003-3760-5401>>),
Clarina Ong [aut],
Tyler Pittman [aut] (<<https://orcid.org/0000-0002-5013-6980>>),
Anna Santiago [aut] (<<https://orcid.org/0000-0002-0932-2386>>),
Yanning Wang [ctr],
Jessica Weiss [aut],
Wei Xu [aut]

Maintainer Lisa Avery <lisa.avery@uhn.ca>

Repository CRAN

Date/Publication 2025-01-24 18:40:02 UTC

Contents

addspace	3
boxcoxfitRx	3
cap	4
clear_labels	4
covsum	5
crrRx	7
ctDNA	8
excelCol	8
excelColLetters	9
extract_labels	10
forestplot2	10
forestplotMV	11
forestplotUV	13
forestplotUVMV	14
formatp	16
geoR_boxcoxfit	16
ggkmcif	17
ggkmcif2	21
ggkmcif_paste	23
hbld	24
lbld	24
lpvalue	24
mvsum	25
nestTable	26
nicename	27
niceNum	28
outTable	28
pembrolizumab	30
plotuv	31
psthr	33
pstprn	33
pvalue	34
replace_plot_labels	34
rmds	35
rm_cifsum	35
rm_compactsum	37
rm_covsum	40
rm_mvsum	43
rm_survdiff	47
rm_survsum	48
rm_survtime	50

`addspace` 3

<code>rm_uvsum</code>	52
<code>rm_uv_mv</code>	57
<code>sanitizestr</code>	59
<code>scrolling_table</code>	59
<code>set_labels</code>	60
<code>set_var_labels</code>	61
<code>uvmodels</code>	61
<code>uvsum</code>	62

Index 65

<code>addspace</code>	<i>Add spaces to strings in LaTeX</i>
-----------------------	---------------------------------------

Description

Add spaces to strings in LaTeX. Returns appends ~~~ before the string

Usage

```
addspace(x)
```

Arguments

<code>x</code>	string
----------------	--------

<code>boxcoxfitRx</code>	<i>fit box cox transformed linear model</i>
--------------------------	---

Description

Wrapper function to fit fine and gray competing risk model using function `crr` from package `cmprsk`

Usage

```
boxcoxfitRx(f, data, lambda = FALSE)
```

Arguments

<code>f</code>	formula for the model. Currently the formula only works by using the name of the column in a dataframe. It does not work by using \$ or [] notation.
<code>data</code>	dataframe containing data
<code>lambda</code>	boolean indicating if you want to output the lamda used in the boxcox transformation. If so the function will return a list of length 2 with the model as the first element and a vector of length 2 as the second.

Value

a list containing the linear model (`lm`) object and, if requested, `lambda`

cap	<i>Capitalize a string</i>
-----	----------------------------

Description

Capitalize a string

Usage

```
cap(x)
```

Arguments

x	string
---	--------

clear_labels	<i>Clear variable labels from a data frame</i>
--------------	--

Description

This function will remove all label attributes from variables in the data.

Usage

```
clear_labels(data)
```

Arguments

data	the data frame to remove labels from
------	--------------------------------------

Details

To change or remove individual labels use `set_labels` or `set_var_labels`

Examples

```
# Set a few variable labels for ctDNA
data("ctDNA")
ctDNA <- ctDNA |> set_var_labels(
  ctdna_status="detectable ctDNA",
  cohort="A cohort label")
# Clear all variable data frames and check
clear_labels(ctDNA)
```

covsum	<i>Get covariate summary dataframe</i>
--------	--

Description

Returns a dataframe corresponding to a descriptive table.

Usage

```
covsum(
  data,
  covs,
  maincov = NULL,
  digits = 1,
  numobs = NULL,
  markup = FALSE,
  sanitize = FALSE,
  nicenames = TRUE,
  IQR = FALSE,
  all.stats = FALSE,
  pvalue = TRUE,
  effSize = FALSE,
  show.tests = FALSE,
  dropLevels = TRUE,
  excludeLevels = NULL,
  full = TRUE,
  digits.cat = 0,
  testcont = c("rank-sum test", "ANOVA"),
  testcat = c("Chi-squared", "Fisher"),
  include_missing = FALSE,
  percentage = c("column", "row")
)
```

Arguments

data	dataframe containing data
covs	character vector with the names of columns to include in table
maincov	covariate to stratify table by
digits	number of digits for summarizing mean data, does not affect p-values
numobs	named list overriding the number of people you expect to have the covariate
markup	boolean indicating if you want latex markup
sanitize	boolean indicating if you want to sanitize all strings to not break LaTeX
nicenames	boolean indicating if you want to replace . and _ in strings with a space
IQR	boolean indicating if you want to display the inter quantile range (Q1,Q3) as opposed to (min,max) in the summary for continuous variables

<code>all.stats</code>	boolean indicating if all summary statistics (Q1,Q3 + min,max on a separate line) should be displayed. Overrides IQR.
<code>pvalue</code>	boolean indicating if you want p-values included in the table
<code>effSize</code>	boolean indicating if you want effect sizes included in the table. Can only be obtained if <code>pvalue</code> is also requested. Effect sizes calculated include Cramer's V for categorical variables, Cohen's d, Wilcoxon r, or Eta-squared for numeric/continuous variables.
<code>show.tests</code>	boolean indicating if the type of statistical test and effect size used should be shown in a column beside the pvalues. Ignored if <code>pvalue=FALSE</code> .
<code>dropLevels</code>	logical, indicating if empty factor levels be dropped from the output, default is TRUE.
<code>excludeLevels</code>	a named list of covariate levels to exclude from statistical tests in the form <code>list(varname =c('level1','level2'))</code> . These levels will be excluded from association tests, but not the table. This can be useful for levels where there is a logical skip (ie not missing, but not presented). Ignored if <code>pvalue=FALSE</code> .
<code>full</code>	boolean indicating if you want the full sample included in the table, ignored if <code>maincov</code> is NULL
<code>digits.cat</code>	number of digits for the proportions when summarizing categorical data (default: 0)
<code>testcont</code>	test of choice for continuous variables,one of <i>rank-sum</i> (default) or <i>ANOVA</i>
<code>testcat</code>	test of choice for categorical variables,one of <i>Chi-squared</i> (default) or <i>Fisher</i>
<code>include_missing</code>	Option to include NA values of <code>maincov</code> . NAs will not be included in statistical tests
<code>percentage</code>	choice of how percentages are presented ,one of <i>column</i> (default) or <i>row</i>

Details

Comparisons for categorical variables default to chi-square tests, but if there are counts of <5 then the Fisher Exact test will be used and if this is unsuccessful then a second attempt will be made computing p-values using MC simulation. If `testcont='ANOVA'` then the t-test with unequal variance will be used for two groups and an ANOVA will be used for three or more. The statistical test used can be displayed by specifying `show.tests=TRUE`.

The number of decimals places to display the statistics can be changed with `digits`, but this will not change the display of p-values. If more significant digits are required for p-values then use `tableOnly=TRUE` and format as desired.

References

- Ellis, P.D. (2010) The essential guide to effect sizes: statistical power, meta-analysis, and the interpretation of research results. Cambridge: Cambridge University Press.[doi:10.1017/CBO9780511761676](https://doi.org/10.1017/CBO9780511761676)
- Lakens, D. (2013) Calculating and reporting effect sizes to facilitate cumulative science: a practical primer for t-tests and ANOVAs. *Frontiers in Psychology*, 4; 863:1-12. [doi:10.3389/fpsyg.2013.00863](https://doi.org/10.3389/fpsyg.2013.00863)

See Also

[fisher.test](#), [chisq.test](#), [wilcox.test](#), [kruskal.test](#), and [anova](#)

crrRx

fit crr model

Description

Wrapper function to fit fine and gray competing risk model using function `crr` from package `cmprsk`

Usage

```
crrRx(f, data)
```

Arguments

<code>f</code>	formula for the model. Currently the formula only works by using the name of the column in a dataframe. It does not work by using <code>\$</code> or <code>[]</code> notation.
<code>data</code>	dataframe containing data

Value

a competing risk model with the call appended to the list

See Also

[cmprsk::crr](#)

Examples

```
# From the crr help file:
set.seed(10)
ftime <- rexp(200)
fstatus <- sample(0:2,200,replace=TRUE)
cov <- matrix(runif(600),nrow=200)
dimnames(cov)[[2]] <- c('x1','x2','x3')
df <- data.frame(ftime,fstatus,cov)
m1 <- crrRx(as.formula('ftime+fstatus~x1+x2+x3'),df)
# Nicely output to report:
rm_mvsum(m1,data=df,showN = TRUE,vif=TRUE)
```

ctDNA	<i>Tumour size change over time Longitudinal changes in tumour size since baseline for patients by changes in ctDNA status (clearance, decrease or increase) since baseline.</i>
-------	--

Description

Tumour size change over time

Longitudinal changes in tumour size since baseline for patients by changes in ctDNA status (clearance, decrease or increase) since baseline.

Usage

```
data('ctDNA')
```

Format

A data frame with 270 rows and 5 variables:

id Patient ID

cohort Study Cohort

ctdna_status Change in ctDNA since baseline

time Number of weeks on treatment

size_change Percentage change in tumour measurement

Source

<https://www.nature.com/articles/s43018-020-0096-5>

excelCol	<i>Retrieve columns number from spreadsheet columns specified as unquoted letters</i>
----------	---

Description

Retrieve columns number from spreadsheet columns specified as unquoted letters

Usage

```
excelCol(...)
```

Arguments

... unquoted excel column headers (i.e. excelCol(A,CG,AA)) separated by commas

Value

a numeric vector corresponding to columns in a spreadsheet

Examples

```
## Find the column numbers for excel columns AB, CE and BB
excelCol(AB,CE,bb)
## Get the columns between A and K and Z
excelCol(A-K,Z)
```

excelColLetters	<i>Retrieve spreadsheet column letter-names from columns indices</i>
-----------------	--

Description

Creates a vector of spreadsheet-style letter-names corresponding to column numbers

Usage

```
excelColLetters(columnIndices)
```

Arguments

columnIndices vector of integer column indices

Details

This is the inverse function of excelCol

Value

a character vector corresponding to the spreadsheet column headings

Examples

```
## Find the column numbers for excel columns AB, CE and BB
colIndices <- excelCol(AB,CE,bb)
## Go back to the column names
excelColLetters(colIndices)
```

extract_labels *Extract variable labels from labelled data frame*

Description

Extract variable labels from data and return a data frame with labels

Usage

```
extract_labels(data, sep = "_")
```

Arguments

data the data frame to extract labels from
sep character used to separate multiple labels, defaults to "_"

Details

All variable names will be returned, even those with no labels. If the label attribute has length greater than one the values will be concatenated and returned as a single string separated by sep

Examples

```
# Set a few variable labels for ctDNA  
data("ctDNA")  
ctDNA <- ctDNA |> set_var_labels(  
  ctdna_status="detectable ctDNA",  
  cohort="A cohort label")  
# Extract labels  
extract_labels(ctDNA)
```

forestplot2 *Create a forest plot using ggplot2*

Description

This function will accept a log or logistic regression fit from glm or geeglm, and display the OR or RR for each variable on the appropriate log scale.

Usage

```
forestplot2(
  model,
  conf.level = 0.95,
  orderByRisk = TRUE,
  colours = "default",
  showEst = TRUE,
  rmRef = FALSE,
  logScale = getOption("reportRmd.logScale", TRUE),
  nxTicks = 5
)
```

Arguments

model	an object output from the glm or geeglm function, must be from a logistic regression
conf.level	controls the width of the confidence interval
orderByRisk	logical, should the plot be ordered by risk
colours	can specify colours for risks less than, 1 and greater than 1.0. Default is red, black, green
showEst	logical, should the risks be displayed on the plot in text
rmRef	logical, should the reference levels be removed for the plot?
logScale	logical, should OR/RR be shown on log scale, defaults to TRUE, or reportRmd.logScale if set. See https://doi.org/10.1093/aje/kwr156 for why you may prefer a linear scale.
nxTicks	Number of tick marks supplied to the log_breaks function to produce

Value

a plot object

Examples

```
data("pembrolizumab")
glm_fit = glm(orr~change_ctdna_group+sex+age+l_size,
  data=pembrolizumab,family = 'binomial')
forestplot2(glm_fit)
```

forestplotMV

Create a multivariable forest plot using ggplot2

Description

This function will send and take log or logistic regression fit from glm or geeglm from mvsum function, and display the OR or RR for each variable on the appropriate log scale.

Usage

```
forestplotMV(
  model,
  data,
  conf.level = 0.95,
  orderByRisk = TRUE,
  colours = "default",
  showEst = TRUE,
  rmRef = FALSE,
  digits = getOption("reportRmd.digits", 2),
  logScale = getOption("reportRmd.logScale", TRUE),
  nxTicks = 5,
  showN = TRUE,
  showEvent = TRUE
)
```

Arguments

model	an object output from the glm or geeglm function, must be from a logistic regression
data	dataframe containing your data
conf.level	controls the width of the confidence interval
orderByRisk	logical, should the plot be ordered by risk
colours	can specify colours for risks less than, 1 and greater than 1.0. Default is red, black, green
showEst	logical, should the risks be displayed on the plot in text
rmRef	logical, should the reference levels be removed for the plot?
digits	number of digits to use displaying estimates
logScale	logical, should OR/RR be shown on log scale, defaults to TRUE, or reportRmd.logScale if set. See https://doi.org/10.1093/aje/kwr156 for why you may prefer a linear scale.
nxTicks	Number of tick marks supplied to the log_breaks function to produce
showN	Show number of observations per variable and category
showEvent	Show number of events per variable and category

Value

a plot object

Examples

```
data("pembrolizumab")
glm_fit = glm(orr~change_ctdna_group+sex+age+l_size,
  data=pembrolizumab,family = 'binomial')
forestplotMV(glm_fit)
```

forestplotUV

Create an univariable forest plot using ggplot2

Description

This function will send and take log or logistic regression fit from glm or geeglm from uvsum function, and display the OR or RR for each variable on the appropriate log scale.

Usage

```
forestplotUV(
  response,
  covs,
  data,
  id = NULL,
  corstr = NULL,
  model = "glm",
  family = NULL,
  digits = getOption("reportRmd.digits", 2),
  conf.level = 0.95,
  orderByRisk = TRUE,
  colours = "default",
  showEst = TRUE,
  rmRef = FALSE,
  logScale = getOption("reportRmd.logScale", TRUE),
  nxTicks = 5,
  showN = TRUE,
  showEvent = TRUE
)
```

Arguments

response	character vector with names of columns to use for response
covs	character vector with names of columns to use for covariates
data	dataframe containing your data
id	character vector which identifies clusters. Only used for geeglm
corstr	character string specifying the correlation structure. Only used for geeglm. The following are permitted: "independence", "exchangeable", "ar1", "unstructured" and "userdefined"
model	fitted model object
family	description of the error distribution and link function to be used in the model. Only used for geeglm
digits	number of digits to round to
conf.level	controls the width of the confidence interval

orderByRisk	logical, should the plot be ordered by risk
colours	can specify colours for risks less than, 1 and greater than 1.0. Default is red, black, green
showEst	logical, should the risks be displayed on the plot in text
rmRef	logical, should the reference levels be removed for the plot?
logScale	logical, should OR/RR be shown on log scale, defaults to TRUE, or reportRmd.logScale if set. See https://doi.org/10.1093/aje/kwr156 for why you may prefer a linear scale.
nxTicks	Number of tick marks supplied to the log_breaks function to produce
showN	Show number of observations per variable and category
showEvent	Show number of events per variable and category

Value

a plot object

Examples

```
data("pembrolizumab")
forestplotUV(response="orr", covs=c("change_ctdna_group", "sex", "age", "l_size"),
data=pembrolizumab, family='binomial')
```

forestplotUVMV

Combine an univariable and multivariable forest plot using ggplot2

Description

This function will take log or logistic regression fit forest plot output from forestplotUV and forestplotMV functions and display the combined adjusted and unadjusted OR or RR for each variable on the appropriate log scale. Please note that total N and reference-level N is taken from unadjusted model.

Usage

```
forestplotUVMV(
  UVmodel,
  MVmodel,
  model = "glm",
  family = NULL,
  digits = getOption("reportRmd.digits", 2),
  orderByRisk = TRUE,
  colours = "default",
  showEst = TRUE,
  rmRef = FALSE,
  logScale = FALSE,
```

```

    nxTicks = 5,
    showN = TRUE,
    showEvent = TRUE
  )

```

Arguments

UVmodel	an UV model object output from the forestplotUV function
MVmodel	a MV model object output from the forestplotMV function
model	fitted model object
family	description of the error distribution and link function to be used in the model. Only used for geeglm
digits	number of digits to round to
orderByRisk	logical, should the plot be ordered by risk
colours	can specify colours for risks less than, 1 and greater than 1.0. Default is red, black, green
showEst	logical, should the risks be displayed on the plot in text
rmRef	logical, should the reference levels be removed for the plot?
logScale	logical, should OR/RR be shown on log scale, defaults to TRUE. See https://doi.org/10.1093/aje/kwr156 for why you may prefer a linear scale.
nxTicks	Number of tick marks supplied to the log_breaks function to produce
showN	Show number of observations per variable and category
showEvent	Show number of events per variable and category

Value

a plot object

Examples

```

data("pembrolizumab")
UVp = forestplotUV(response="orr", covs=c("change_ctdna_group", "sex", "age",
    "l_size"), data=pembrolizumab, family='binomial')
MVp = forestplotMV(glm(orr~change_ctdna_group+sex+age+l_size,
    data=pembrolizumab,family = 'binomial'))
forestplotUVMV(UVp, MVp)

```

formatp	<i>Specific p-value formatting</i>
---------	------------------------------------

Description

If $p < 0.001$ returns "<0.001", if $p < 0.01$ returns p to 3 decimal places otherwise returns p to 2 decimal places

Usage

```
formatp(pvalues)
```

Arguments

pvalues	a vector of p values
---------	----------------------

geoR_boxcoffit	<i>Parameter Estimation for the Box-Cox Transformation</i>
----------------	--

Description

This function is copied from the geoR package which has been removed from the CRAN repository.

Usage

```
geoR_boxcoffit(object, xmat, lambda, lambda2 = NULL, add.to.data = 0)
```

Arguments

object	a vector with the data
xmat	a matrix with covariates values. Defaults to rep(1, length(y)).
lambda	numerical value(s) for the transformation parameter lambda. Used as the initial value in the function for parameter estimation. If not provided default values are assumed. If multiple values are passed the one with highest likelihood is used as initial value.
lambda2	logical or numerical value(s) of the additional transformation (see DETAILS below). Defaults to NULL. If TRUE this parameter is also estimated and the initial value is set to the absolute value of the minimum data. A numerical value is provided it is used as the initial value. Multiple values are allowed as for lambda.
add.to.data	a constant value to be added to the data.

Details

For more information see: <https://cran.r-project.org/web/packages/geoR/index.html>

`ggkmcif`*Plot KM and CIF curves with ggplot*

Description

This function will plot a KM or CIF curve with option to add the number at risk. You can specify if you want confidence bands, the hazard ratio, and pvalues, as well as the units of time used.

Usage

```
ggkmcif(  
  response,  
  cov = NULL,  
  data,  
  type = NULL,  
  pval = TRUE,  
  HR = FALSE,  
  HR_pval = FALSE,  
  conf.curves = FALSE,  
  conf.type = "log",  
  table = TRUE,  
  times = NULL,  
  xlab = "Time",  
  ylab = NULL,  
  main = NULL,  
  stratalabs = NULL,  
  strataname = nice(cov),  
  stratalabs.table = NULL,  
  strataname.table = strataname,  
  median.text = FALSE,  
  median.lines = FALSE,  
  median.CI = FALSE,  
  set.time.text = NULL,  
  set.time.line = FALSE,  
  set.time = 5,  
  set.time.CI = FALSE,  
  censor.marks = TRUE,  
  censor.size = 3,  
  censor.stroke = 1.5,  
  fsize = 10,  
  nsize = 3,  
  lsize = 1,  
  psize = 3.5,  
  median.size = 3,  
  median.pos = NULL,  
  median.lsize = 1,  
  set.size = 3,  
)
```

```

set.pos = NULL,
set.lsize = 1,
ylim = c(0, 1),
col = NULL,
linetype = NULL,
xlim = NULL,
legend.pos = NULL,
pval.pos = NULL,
plot.event = 1,
event = c("col", "linetype"),
flip.CIF = FALSE,
cut = NULL,
eventlabs = NULL,
event.name = NULL,
Numbers_at_risk_text = "Numbers at risk",
HR.digits = 2,
HR.pval.digits = 3,
pval.digits = 3,
median.digits = 3,
set.time.digits = 3,
returns = FALSE,
print.n.missing = TRUE
)

```

Arguments

response	character vector with names of columns to use for response
cov	String specifying the column name of stratification variable
data	dataframe containing your data
type	string indicating the type of univariate model to fit. The function will try and guess what type you want based on your response. If you want to override this you can manually specify the type. Options include "KM", and "CIF"
pval	boolean to specify if you want p-values in the plot (Log Rank test for KM and Gray's test for CIF)
HR	boolean to specify if you want hazard ratios included in the plot
HR_pval	boolean to specify if you want HR p-values in the plot
conf.curves	boolean to specify if you want confidence interval bands
conf.type	One of "none"(the default), "plain", "log", "log-log" or "logit". Only enough of the string to uniquely identify it is necessary. The first option causes confidence intervals not to be generated. The second causes the standard intervals curve $\pm k * se(\text{curve})$, where k is determined from conf.int. The log option calculates intervals based on the cumulative hazard or log(survival). The log-log option bases the intervals on the log hazard or log(-log(survival)), and the logit option on $\log(\text{survival}/(1-\text{survival}))$.
table	Logical value. If TRUE, includes the number at risk table
times	Numeric vector of times for the x-axis

xlab	String corresponding to xlabel. By default is "Time"
ylab	String corresponding to ylabel. When NULL uses "Survival probability" for KM curves, and "Probability of an event" for CIF
main	String corresponding to main title. When NULL uses Kaplan-Meier Plots, and "Cumulative Incidence Plot for CIF"
stratalabs	string corresponding to the labels of the covariate, when NULL will use the levels of the covariate
strataname	String of the covariate name default is nicename(cov)
stratalabs.table	String corresponding to the levels of the covariate for the number at risk table, when NULL will use the levels of the covariate. Can use a string of "-" when the labels are long
strataname.table	String of the covariate name for the number at risk table default is nicename(cov)
median.text	boolean to specify if you want the median values added to the legend (or as added text if there are no covariates), for KM only
median.lines	boolean to specify if you want the median values added as lines to the plot, for KM only
median.CI	boolean to specify if you want the 95% with the median text (Only for KM)
set.time.text	string for the text to add survival at a specified time (eg. year OS)
set.time.line	boolean to specify if you want the survival added as lines to the plot at a specified point
set.time	Numeric values of the specific time of interest, default is 5 (Multiple values can be entered)
set.time.CI	boolean to specify if you want the 95% interval with the set time text
censor.marks	logical value. If TRUE, includes censor marks (only for KM curves)
censor.size	size of censor marks, default is 3
censor.stroke	stroke of censor marks, default is 1.5
fsize	font size
nsize	font size for numbers in the numbers at risk table
lsize	line size
psize	size of the pvalue
median.size	size of the median text (Only when there are no covariates)
median.pos	vector of length 2 corresponding to the median position (Only when there are no covariates)
median.lsize	line size of the median lines
set.size	size of the survival at a set time text (Only when there are no covariates)
set.pos	vector of length 2 corresponding to the survival at a set point position (Only when there are no covariates)
set.lsize	line size of the survival at set points

<code>ylim</code>	vector of length 2 corresponding to limits of y-axis. Default to NULL
<code>col</code>	vector of colours
<code>linetype</code>	vector of line types
<code>xlim</code>	vector of length 2 corresponding to limits of x-axis. Default to NULL
<code>legend.pos</code>	Can be either a string corresponding to the legend position ("left", "top", "right", "bottom", "none") or a vector of length 2 corresponding to the legend position (uses normalized units (ie the c(0.5,0.5) is the middle of the plot))
<code>pval.pos</code>	vector of length 2 corresponding to the p-value position
<code>plot.event</code>	Which event(s) to plot (1,2, or c(1,2))
<code>event</code>	String specifying if the event should be mapped to the colour, or linetype when plotting both events to colour = "col", line type
<code>flip.CIF</code>	boolean to flip the CIF curve to start at 1
<code>cut</code>	numeric value indicating where to divide a continuous covariate (default is the median)
<code>eventlabs</code>	String corresponding to the event type names
<code>event.name</code>	String corresponding to the label of the event types
<code>Numbers_at_risk_text</code>	String for the label of the number at risk
<code>HR.digits</code>	Number of digits printed of the hazard ratio
<code>HR.pval.digits</code>	Number of digits printed of the hazard ratio pvalue
<code>pval.digits</code>	Number of digits printed of the Gray's/log rank pvalue
<code>median.digits</code>	Number of digits printed of the median pvalue
<code>set.time.digits</code>	Number of digits printed of the probability at a specified time
<code>returns</code>	Logical value returns a list with all ggplot objects in a list
<code>print.n.missing</code>	Logical, should the number of missing be shown !Needs to be checked

Details

Note that for proper pdf output of special characters the following code needs to be included in the first chunk of the rmd `knitr::opts_chunk$set(dev="cairo_pdf")`

Value

Nothing is returned unless `returns = TRUE` is used. With `returns = TRUE`, if `table=TRUE` (the default) a table style graphic with survival plot and number at risk table is returned. Otherwise a plot with the survival curves is returned.

Examples

```

data("pembrolizumab")
# Simple plot without confidence intervals
ggkmcif(response = c('os_time','os_status'),
cov='cohort',
data=pembrolizumab)

# Plot with median survival time
ggkmcif(response = c('os_time','os_status'),
cov='sex',
data=pembrolizumab,
median.text = TRUE,median.lines=TRUE,conf.curves=TRUE)

# Plot with specified survival times and log-log CI
ggkmcif(response = c('os_time','os_status'),
cov='sex',
data=pembrolizumab,
median.text = FALSE,set.time.text = 'mo OS',
set.time = c(12,24),conf.type = 'log-log',conf.curves=TRUE)

# KM plot with 95% CI and censor marks
ggkmcif(c('os_time','os_status'),'sex',data = pembrolizumab, type = 'KM',
HR=TRUE, HR_pval = TRUE, conf.curves = TRUE,conf.type='log-log',
set.time.CI = TRUE, censor.marks=TRUE)

```

ggkmcif2

*Plot KM and CIF curves with ggplot***Description**

This function will plot a KM or CIF curve with option to add the number at risk. You can specify if you want confidence bands, the hazard ratio, and pvalues, as well as the units of time used.

Usage

```

ggkmcif2(
  response,
  cov = NULL,
  data,
  pval = TRUE,
  conf.curves = FALSE,
  table = TRUE,
  xlab = "Time",
  ylab = NULL,
  col = NULL,
  times = NULL,
  type = NULL,
  plot.event = 1,

```

```

    returns = FALSE,
    ...
  )

```

Arguments

response	character vector with names of columns to use for response
cov	String specifying the column name of stratification variable
data	dataframe containing your data
pval	boolean to specify if you want p-values in the plot (Log Rank test for KM and Gray's test for CIF)
conf.curves	boolean to specify if you want confidence interval bands
table	Logical value. If TRUE, includes the number at risk table
xlab	String corresponding to xlabel. By default is "Time"
ylab	String corresponding to ylabel. When NULL uses "Survival"
col	vector of colours
times	Numeric vector of times for the x-axis probability" for KM cuves, and "Probability of an event" for CIF
type	string indicating he type of univariate model to fit. The function will try and guess what type you want based on your response. If you want to override this you can manually specify the type. Options include "KM", and ,"CIF"
plot.event	Which event(s) to plot (1,2, or c(1,2))
returns	boolean indicating if a list with the objects should be returned. Default is FALSE and plot will be printed
...	for additional plotting arguments see ggkmcif2Parameters

Details

Note that for proper pdf output of special characters the following code needs to be included in the first chunk of the rmd knitr::opts_chunk\$set(dev="cairo_pdf")

Value

ggplot object; if table = F then only curves are output; if table = T then curves and risk table are output together

Examples

```

# Simple plot without confidence intervals
data("pembrolizumab")
ggkmcif2(response = c('os_time', 'os_status'),
cov='cohort',
data=pembrolizumab)

# Plot with median survival time
ggkmcif2(response = c('os_time', 'os_status'),

```

```

cov='sex',
data=pembrolizumab,
median.text = TRUE,median.lines=TRUE,conf.curves=TRUE)

# Plot with specified survival times and log-log CI
ggkmcif2(response = c('os_time','os_status'),
cov='sex',
data=pembrolizumab,
median.text = FALSE,set.time.text = 'mo OS',
set.time = c(12,24),conf.type = 'log-log',conf.curves=TRUE)

# KM plot with 95% CI and censor marks
ggkmcif2(c('os_time','os_status'),'sex',data = pembrolizumab, type = 'KM',
HR=TRUE, HR_pval = TRUE, conf.curves = TRUE,conf.type='log-log',
set.time.CI = TRUE, censor.marks=TRUE)

```

ggkmcif_paste

Plot KM and CIF curves with ggplot

Description

This function puts together a survival curve, and a number at risk table

Usage

```
ggkmcif_paste(list_gg)
```

Arguments

`list_gg` list containing the results of ggkmcif

Value

a gtable with three elements, the survival curve, a spacer and the number at risk table

Examples

```

data("pembrolizumab")
plot <- ggkmcif(response=c('pfs_time','pfs_status'),
data=pembrolizumab,returns = TRUE)

# Highlighting a section of the curve
plot[[1]] <- plot[[1]] +
ggplot2::geom_rect(xmin=4,xmax=8,ymin=0.15,ymax=0.4,alpha=0.01,fill='yellow')

# Putting the curve back together
ggkmcif_paste(plot)

```

hbold	<i>Bold strings in HTML</i>
-------	-----------------------------

Description

Bold strings in HTML

Usage

```
hbold(strings)
```

Arguments

strings	A vector of strings to bold.
---------	------------------------------

lbold	<i>Bold strings in LaTeX</i>
-------	------------------------------

Description

Bold strings in LaTeX

Usage

```
lbold(strings)
```

Arguments

strings	A vector of strings to bold.
---------	------------------------------

lpvalue	<i>Formats p-values for LaTeX</i>
---------	-----------------------------------

Description

Returns <0.001 if pvalue is <0.001. Else rounds the pvalue to specified significant digits. Will bold the p-value if it is <= 0.05

Usage

```
lpvalue(x, sigdigits = 2)
```

Arguments

x	an integer
sigdigits	number of significant digit to report

mvsum

*Get multivariate summary dataframe***Description**

Returns a dataframe with the model summary and global p-value for multi-level variables.

Usage

```
mvsum(
  model,
  data,
  digits = getOption("reportRmd.digits", 2),
  showN = TRUE,
  showEvent = TRUE,
  markup = TRUE,
  sanitize = TRUE,
  nicenames = TRUE,
  CIwidth = 0.95,
  vif = TRUE
)
```

Arguments

model	fitted model object
data	dataframe containing data
digits	number of digits to round to
showN	boolean indicating sample sizes should be shown for each comparison, can be useful for interactions
showEvent	boolean indicating if number of events should be shown. Only available for logistic.
markup	boolean indicating if you want latex markup
sanitize	boolean indicating if you want to sanitize all strings to not break LaTeX
nicenames	boolean indicating if you want to replace . and _ in strings with a space.
CIwidth	width for confidence intervals, defaults to 0.95
vif	boolean indicating if the variance inflation factor should be included. See details

Details

Global p-values are likelihood ratio tests for lm, glm and polr models. For lme models an attempt is made to re-fit the model using ML and if successful LRT is used to obtain a global p-value. For coxph models the model is re-run without robust variances with and without each variable and a LRT is presented. If unsuccessful a Wald p-value is returned. For GEE and CRR models Wald global p-values are returned.

If the variance inflation factor is requested (VIF=T) then a generalised VIF will be calculated in the same manner as the car package.

VIF for competing risk models is computed by fitting a linear model with a dependent variable comprised of the sum of the model independent variables and then calculating VIF from this linear model.

References

John Fox & Georges Monette (1992) Generalized Collinearity Diagnostics, Journal of the American Statistical Association, 87:417, 178-183, DOI: 10.1080/01621459.1992.10475190

John Fox and Sanford Weisberg (2019). An R Companion to Applied Regression, Third Edition. Thousand Oaks CA: Sage.

nestTable	<i>Combine two table columns into a single column with levels of one nested within levels of the other.</i>
-----------	---

Description

This function accepts a data frame (via the data argument) and combines two columns into a single column with values from the head_col serving as headers and values of the to_col displayed underneath each header. The resulting table is then passed to outTable for printing and output, to use the grouped table as a data frame specify tableOnly=TRUE. By default the headers will be bolded and the remaining values indented.

Usage

```
nestTable(
  data,
  head_col,
  to_col,
  colHeader = "",
  caption = NULL,
  indent = TRUE,
  boldheaders = TRUE,
  hdr_prefix = "",
  hdr_suffix = "",
  digits = getOption("reportRmd.digits", 2),
  tableOnly = FALSE,
  fontsize
)
```

Arguments

data	dataframe
head_col	character value specifying the column name with the headers

to_col	character value specifying the column name to add the headers into
colHeader	character with the desired name of the first column. The default is to leave this empty for output or, for table only output to use the column name 'col1'.
caption	table caption
indent	Boolean should the original values in the to_col be indented
boldheaders	Boolean should the header column values be bolded
hdr_prefix	character value that will prefix headers
hdr_suffix	character value that will suffix headers
digits	number of digits to round numeric columns to, wither a single number or a vector corresponding to the number of numeric columns
tableOnly	boolean indicating if the table should be formatted for printing or returned as a data frame
fontsize	PDF/HTML output only, manually set the table fontsize

Details

Note that it is possible to combine multiple tables (more than two) with this function.

Value

A character vector of the table source code, unless tableOnly=TRUE in which case a data frame is returned

Examples

```
## Investigate models to predict baseline ctDNA and tumour size and display together
## (not clinically useful!)
data(pembroлизумаб)
fit1 <- lm(baseline_ctdna~age+l_size+pd11,data=pembroлизумаб)
m1 <- rm_mvsum(fit1,tableOnly=TRUE)
m1$Response = 'ctDNA'
fit2 <- lm(l_size~age+baseline_ctdna+pd11,data=pembroлизумаб)
m2 <- rm_mvsum(fit2,tableOnly=TRUE)
m2$Response = 'Tumour Size'
nestTable(rbind(m1,m2),head_col='Response',to_col='Covariate')
```

nicename

Lean strings for printing

Description

Returns strings with . and _ replaced by a space. This is nice when printing column names of your dataframe in a report

Usage

```
nicename(strings, check_numbers = TRUE)
```

Arguments

strings vector of strings to give a nice name
 check_numbers boolean indicating if numbers with decimals should be checked for and retained.

niceNum	<i>Round retaining digits</i>
---------	-------------------------------

Description

Round retaining digits

Usage

```
niceNum(x, digits = 2)
```

Arguments

x a vector
 digits numeric

outTable	<i>Print tables to PDF/Latex HTML or Word</i>
----------	---

Description

Output the table nicely to whatever format is appropriate. This is the output function used by the `rm_*` printing functions.

Usage

```
outTable(
  tab,
  row.names = NULL,
  to_indent = numeric(0),
  bold_headers = TRUE,
  rows_bold = numeric(0),
  bold_cells = NULL,
  caption = NULL,
  digits = getOption("reportRmd.digits", 2),
  align,
  applyAttributes = TRUE,
```

```

    keep.rownames = FALSE,
    nicenames = TRUE,
    fontsize,
    chunk_label,
    format = NULL
  )

```

Arguments

tab	a table to format
row.names	a string specifying the column name to assign to the rownames. If NULL (the default) then rownames are removed.
to_indent	numeric vector indicating which rows to indent in the first column.
bold_headers	boolean indicating if the column headers should be bolded
rows_bold	numeric vector indicating which rows to bold
bold_cells	array indices indicating which cells to bold. These will be in addition to rows bolded by rows_bold.
caption	table caption
digits	number of digits to round numeric columns to, wither a single number or a vector corresponding to the number of numeric columns in tab
align	string specifying column alignment, defaults to left alignment of the first column and right alignment of all other columns. The align argument accepts a single string with 'l' for left, 'c' for centre and 'r' for right, with no separations. For example, to set the left column to be centred, the middle column right-aligned and the right column left aligned use: align='crl'
applyAttributes	boolean indicating if the function should use to_indent and bold_cells formatting attributes. This will only work properly if the dimensions of the table output from rm_covsum, rm_uvsum etc haven't changed.
keep.rownames	should the row names be included in the output
nicenames	boolean indicating if you want to replace . and _ in strings with a space
fontsize	PDF/HTML output only, manually set the table fontsize
chunk_label	only used knitting to Word docs to allow cross-referencing
format	if specified ('html','latex') will override the global pandoc setting

Details

Entire rows can be bolded, or specific cells. Currently indentation refers to the first column only. By default, underscores in column names are converted to spaces. To disable this set rm_ to FALSE

Value

A character vector of the table source code, unless tableOnly=TRUE in which case a data frame is returned

Examples

```
# To make custom changes or change the fontsize in PDF/HTML
data("pembrolizumab")
tab <- rm_covsum(data=pembrolizumab,maincov = 'change_ctdna_group',
covs=c('age','sex','pd11','tmb','l_size'),show.tests=TRUE,tableOnly = TRUE)
outTable(tab, fontsize=7)

# To bold columns with the variable names
rows_bold <- c(1,4,7,10,13)
outTable(tab,rows_bold = rows_bold)

# To bold the estimates for male/female
bold_cells <- as.matrix(expand.grid(5:6,1:ncol(tab)))
outTable(tab,bold_cells= bold_cells)

# Output the above table to HTML or LaTeX
#cat(outTable(tab=tab)) #Knits to specified global setting
#cat(outTable(tab, format="html"), file = "tab.html") #HTML output
#cat(outTable(tab, format="latex"), file = "tab.tex") #LaTeX output
```

pembrolizumab	<i>Survival data Survival status and ctDNA levels for patients receiving pembrolizumab</i>
---------------	--

Description

Survival data

Survival status and ctDNA levels for patients receiving pembrolizumab

Usage

```
data('pembrolizumab')
```

Format

A data frame with 94 rows and 15 variables:

id Patient ID

age Age at study entry

sex Patient Sex

cohort Study Cohort

l_size Target lesion size at baseline

pd11 PD L1 percent

tmb log of TMB

baseline_ctdna Baseline ctDNA

change_ctdna_group Did ctDNA increase or decrease from baseline to cycle 3

orr Objective Response
cbr Clinical Beneficial Response
os_status Overall survival status
os_time Overall survival time in months
pfs_status Progression free survival status
pfs_time Progression free survival time in months

Source

<https://www.nature.com/articles/s43018-020-0096-5>

plotuv

Plot multiple bivariate relationships in a single plot

Description

This function is designed to accompany `rm_uvsum` as a means of visualising the results, and uses similar syntax.

Usage

```
plotuv(  
  response,  
  covs,  
  data,  
  showN = FALSE,  
  showPoints = TRUE,  
  na.rm = TRUE,  
  response_title = NULL,  
  return_plotlist = FALSE,  
  ncol = 2,  
  p_margins = c(0, 0.2, 1, 0.2),  
  bpThreshold = 20,  
  mixed = TRUE,  
  violin = FALSE,  
  position = c("dodge", "stack", "fill"),  
  use_labels = TRUE  
)
```

Arguments

<code>response</code>	character vector with names of columns to use for response
<code>covs</code>	character vector with names of columns to use for covariates
<code>data</code>	dataframe containing your data
<code>showN</code>	boolean indicating whether sample sizes should be shown on the plots

showPoints	boolean indicating whether individual data points should be shown when n>20 in a category
na.rm	boolean indicating whether na values should be shown or removed
response_title	character value with title of the plot
return_plotlist	boolean indicating that the list of plots should be returned instead of a plot, useful for applying changes to the plot, see details
ncol	the number of columns of plots to be display in the ggarrange call, defaults to 2
p_margins	sets the TRBL margins of the individual plots, defaults to c(0,0.2,1,.2)
bpThreshold	Default is 20, if there are fewer than 20 observations in a category then dotplots, as opposed to boxplots are shown.
mixed	should a mix of dotplots and boxplots be shown based on sample size? If false then all categories will be shown as either dotplots, or boxplots according the bpThreshold and the smallest category size
violin	Show violin plots instead of boxplots. This will override bpThreshold and mixed.
position	for categorical variables how should barplots be presented. Default is "dodge" IF stack is TRUE then n will not be shown.
use_labels	boolean, default is true if the variables have label attributes this will be shown in the plot instead of the variable names, or if there are no labels then tidy versions of the variable names will be used. If use_labels=FALSE the variable names will be used.

Details

Plots are displayed as follows: If response is continuous For a numeric predictor scatterplot For a categorical predictor: If 20+ observations available boxplot, otherwise dotplot with median line If response is a factor For a numeric predictor: If 20+ observations available boxplot, otherwise dotplot with median line For a categorical predictor barplot Response variables are shown on the ordinate (y-axis) and covariates on the abscissa (x-axis)

Variable names are replaced by their labels if available, or by tidy versions if not. Set use_labels=FALSE to use the variable names.

Value

a list containing plots for each variable in covs

See Also

[ggplot2::ggplot](#) and [ggpubr::ggarrange](#) [replace_plot_labels](#)

Examples

```
## Run multiple univariate analyses on the pembrolizumab dataset to predict cbr and
## then visualise the relationships.
data("pembrolizumab")
```



```
rm_uvsum(data=pembrolizumab,  
response='cbr',covs=c('age','sex','l_size','baseline_ctdna'))  
plotuv(data=pembrolizumab, response='cbr',  
covs=c('age','sex','l_size','baseline_ctdna'),showN=TRUE)
```

psthr

Round and paste with parentheses

Description

Round and paste with parentheses

Usage

```
psthr(x, y = 2)
```

Arguments

x	a numeric vector
y	integer corresponding to the number of digits to round by

pstprn

Paste with parentheses

Description

Paste with parentheses

Usage

```
pstprn(x)
```

Arguments

x	a vector
---	----------

pvalue	<i>Formats p-values</i>
--------	-------------------------

Description

Returns <0.001 if pvalue is <0.001. Else rounds the pvalue to specified significant digits

Usage

```
pvalue(x, digits)
```

Arguments

x	an integer
digits	the number of significant digits to return

replace_plot_labels	<i>Replace variable names with labels in ggplot</i>
---------------------	---

Description

If the data stored in a ggplot object has variable labels then this will replace the variable names with the variable labels. If no labels are set then the variable names will be tidied and a nicer version used.

Usage

```
replace_plot_labels(plot)
```

Arguments

plot	output from a call to ggplot2
------	-------------------------------

See Also

[set_var_labels\(\)](#) for setting individual variable labels, [set_labels\(\)](#) for setting variable labels using a data frame, [extract_labels\(\)](#) for creating a data frame of all variable labels, [clear_labels\(\)](#) for removing variable labels

Examples

```
## Not run:
data("pembrolizumab")
p <- ggplot(pembrolizumab, aes(x=change_ctdna_group, y=baseline_ctdna)) +
  geom_boxplot()
replace_plot_labels(p)
pembrolizumab <- set_var_labels(pembrolizumab,
  change_ctdna_group="Change in ctDNA group")
p <- ggplot(pembrolizumab, aes(x=change_ctdna_group, y=baseline_ctdna)) +
  geom_boxplot()
replace_plot_labels(p)
# Can also be used with a pipe, but expression needs to be wrapped in a brace
(ggplot(pembrolizumab, aes(x=change_ctdna_group, y=baseline_ctdna)) +
  geom_boxplot()) |> replace_plot_labels()

## End(Not run)
```

rmds

Replace dollar signs with html for proper HTML output

Description

Replace dollar signs with html for proper HTML output

Usage

```
rmds(s)
```

Arguments

s a character vector

rm_cifsum

Summarize cumulative incidence by group

Description

Displays event counts and event rates at specified time points for the entire cohort and by group. Gray's test of differences in cumulative incidence is displayed.

Usage

```

rm_cifsum(
  data,
  time,
  status,
  group = NULL,
  eventcode = 1,
  cencode = 0,
  eventtimes,
  eventtimeunit,
  eventtimeLbls = NULL,
  CIwidth = 0.95,
  unformattedp = FALSE,
  na.action = "na.omit",
  showCounts = TRUE,
  showGraystest = TRUE,
  digits = 2,
  caption = NULL,
  tableOnly = FALSE
)

```

Arguments

<code>data</code>	data frame containing survival data
<code>time</code>	string indicating survival time variable
<code>status</code>	string indicating event status variable; must have at least 3 levels, e.g. 0 = censor, 1 = event, 2 = competing risk
<code>group</code>	string or character vector indicating the variable to group observations by
<code>eventcode</code>	numerical variable indicating event, default is 1
<code>cencode</code>	numerical variable indicating censored observation, default is 0
<code>eventtimes</code>	numeric vector specifying when event probabilities should be calculated
<code>eventtimeunit</code>	unit of time to suffix to the time column label if event probabilities are requested, should be plural
<code>eventtimeLbls</code>	if supplied, a vector the same length as <code>eventtimes</code> with descriptions (useful for displaying years with data provided in months)
<code>CIwidth</code>	width of the event probabilities, default is 95%
<code>unformattedp</code>	boolean indicating if you would like the p-value to be returned unformatted (ie not rounded or prefixed with '<'). Should be used in conjunction with the <code>digits</code> argument.
<code>na.action</code>	default is to omit missing values, but can be set to throw an error using <code>na.action='na.fail'</code>
<code>showCounts</code>	boolean indicating if the at risk, events and censored columns should be output, default is TRUE
<code>showGraystest</code>	boolean indicating Gray's test should be included in the final table, default is TRUE

digits	the number of digits to report in the event probabilities, default is 2.
caption	table caption for markdown output
tableOnly	should a dataframe or a formatted object be returned

Value

A character vector of the event table source code, unless tableOnly=TRUE in which case a data frame is returned

Examples

```
library(survival)
data(pbc)

# Event probabilities at various time points with replacement time labels
rm_cifsum(data=pbc,time='time',status='status',
eventtimes=c(1825,3650),eventtimeLbls=c(5,10),eventtimeunit='yr')

# Event probabilities by one group
rm_cifsum(data=pbc,time='time',status='status',group='trt',
eventtimes=c(1825,3650),eventtimeunit='day')

# Event probabilities by multiple groups
rm_cifsum(data=pbc,time='time',status='status',group=c('trt','sex'),
eventtimes=c(1825,3650),eventtimeunit='day')
```

rm_compactsum	<i>Output a compact summary table</i>
---------------	---------------------------------------

Description

Outputs a table formatted for pdf, word or html output with summary statistics

Usage

```
rm_compactsum(
  data,
  xvars,
  grp,
  use_mean,
  caption = NULL,
  tableOnly = FALSE,
  covTitle = "",
  digits = 1,
  digits.cat = 0,
  nicenames = TRUE,
```

```

iqr = TRUE,
all.stats = FALSE,
pvalue = TRUE,
effSize = FALSE,
p.adjust = "none",
unformattedp = FALSE,
show.sumstats = FALSE,
show.tests = FALSE,
full = TRUE,
percentage = "col"
)

```

Arguments

data	dataframe containing data
xvars	character vector with the names of covariates to include in table
grp	character with the name of the grouping variable
use_mean	logical indicating whether mean and standard deviation will be returned for continuous variables instead of median. Otherwise, can specify for individual variables using a character vector containing the names of covariates to return mean and sd for (if use_mean is not supplied, all covariates will have median summaries). See examples.
caption	character containing table caption (default is no caption)
tableOnly	logical, if TRUE then a dataframe is returned, otherwise a formatted printed object is returned (default is FALSE)
covTitle	character with the name of the covariate (predictor) column. The default is to leave this empty for output or, for table only output to use the column name 'Covariate'
digits	numeric specifying the number of digits for summarizing mean data. Digits can be specified for individual variables using a named vector in the format digits=c("var1"=2,"var2"=3). If a variable is not in the vector the default will be used for it (default is 1). See examples
digits.cat	numeric specifying the number of digits for the proportions when summarizing categorical data (default is 0)
nicenames	logical indicating if you want to replace . and _ in strings . with a space
iqr	logical indicating if you want to display the interquartile range (Q1-Q3) as opposed to (min-max) in the summary for continuous variables
all.stats	logical indicating if all summary statistics (Q1, Q3 + min, max on a separate line) should be displayed. Overrides iqr
pvalue	logical indicating if you want p-values included in the table
effSize	logical indicating if you want effect sizes and their 95% confidence intervals included in the table. Effect sizes calculated include Cramer's V for categorical variables, and Cohen's d, Wilcoxon r, Epsilon-squared, or Omega-squared for numeric/continuous variables
p.adjust	p-adjustments to be performed

unformattedp	logical indicating if you would like the p-value to be returned unformatted (ie. not rounded or prefixed with '<'). Best used with tableOnly = T and outTable function. See examples
show.sumstats	logical indicating if the type of statistical summary (mean, median, etc) used should be shown.
show.tests	logical indicating if the type of statistical test and effect size (if effSize = TRUE) used should be shown in a column beside the p-values.
full	logical indicating if you want the full sample included in the table, ignored if grp is not specified
percentage	choice of how percentages are presented, either column (default) or row

Details

Comparisons for categorical variables default to chi-square tests, but if there are counts of <5 then the Fisher Exact test will be used. For grouping variables with two levels, either t-tests (mean) or wilcoxon tests (median) will be used for numerical variables. Otherwise, ANOVA (mean) or Kruskal- Wallis tests will be used. The statistical test used can be displayed by specifying show.tests = TRUE. Statistical tests and effect sizes for grp and/ or xvars with less than 2 counts in any level will not be shown.

Effect sizes are calculated as Cohen d for between group differences if the variable is summarised with the mean, otherwise Wilcoxon R if summarised with a median. Cramer's V is used for categorical variables, omega is used for differences in means among more than two groups and epsilon for differences in medians among more than two groups. Confidence intervals are calculated using bootstrapping.

tidyselect can only be used for xvars and grp arguments. Additional arguments (digits, use_mean) must be passed in using characters if variable names are used.

Value

A character vector of the table source code, unless tableOnly = TRUE in which case a data frame is returned. The output has the following attribute:

- "description", which describes what is included in the output table and the type of statistical summary for each covariate. When applicable, the types of statistical tests used will be included. If effSize = TRUE, the effect sizes for each covariate will also be mentioned.

References

- Smithson, M. (2002). Noncentral Confidence Intervals for Standardized Effect Sizes. (07/140 ed., Vol. 140). SAGE Publications. doi:10.4135/9781412983761.n4
- Steiger, J. H. (2004). Beyond the F Test: Effect Size Confidence Intervals and Tests of Close Fit in the Analysis of Variance and Contrast Analysis. Psychological Methods, 9(2), 164–182. doi:10.1037/1082989X.9.2.164
- Kelley, T. L. (1935). An Unbiased Correlation Ratio Measure. Proceedings of the National Academy of Sciences - PNAS, 21(9), 554–559. doi:10.1073/pnas.21.9.554
- Okada, K. (2013). Is Omega Squared Less Biased? A Comparison of Three Major Effect Size Indices in One-Way ANOVA. Behavior Research Methods, 40(2), 129-147.

Breslow, N. (1970). A generalized Kruskal-Wallis test for comparing K samples subject to unequal patterns of censorship. *Biometrika*, 57(3), 579-594.

FRITZ, C. O., MORRIS, P. E., & RICHLER, J. J. (2012). Effect Size Estimates: Current Use, Calculations, and Interpretation. *Journal of Experimental Psychology. General*, 141(1), 2–18. doi:10.1037/a0024338

Examples

```
data("pembrolizumab")
rm_compactsum(data = pembrolizumab, xvars = c("age",
"change_ctdna_group", "l_size", "pd11"), grp = "sex", use_mean = "age",
digits = c("age" = 2, "l_size" = 3), digits.cat = 1, iqr = TRUE,
show.tests = TRUE)

# Other Examples (not run)
## Include the summary statistic in the variable column
#rm_compactsum(data = pembrolizumab, xvars = c("age",
#"change_ctdna_group"), grp = "sex", use_mean = "age", show.sumstats=TRUE)

## To show effect sizes
#rm_compactsum(data = pembrolizumab, xvars = c("age",
#"change_ctdna_group"), grp = "sex", use_mean = "age", digits = 2,
#effSize = TRUE, show.tests = TRUE)

## To return unformatted p-values
#rm_compactsum(data = pembrolizumab, xvars = c("l_size",
#"change_ctdna_group"), grp = "cohort", effSize = TRUE, unformattedp = TRUE)

## Using tidyselect
#pembrolizumab |> rm_compactsum(xvars = c(age, sex, pd11), grp = cohort,
#effSize = TRUE)
```

rm_covsum

Outputs a descriptive covariate table

Description

Returns a data frame corresponding to a descriptive table.

Usage

```
rm_covsum(
  data,
  covs,
  maincov = NULL,
  caption = NULL,
  tableOnly = FALSE,
  covTitle = "",
```



```

  digits = 1,
  digits.cat = 0,
  nicenames = TRUE,
  IQR = FALSE,
  all.stats = FALSE,
  pvalue = TRUE,
  effSize = FALSE,
  p.adjust = "none",
  unformattedp = FALSE,
  show.tests = FALSE,
  testcont = c("rank-sum test", "ANOVA"),
  testcat = c("Chi-squared", "Fisher"),
  full = TRUE,
  include_missing = FALSE,
  percentage = c("column", "row"),
  dropLevels = TRUE,
  excludeLevels = NULL,
  numobs = NULL,
  fontsize,
  chunk_label
)

```

Arguments

<code>data</code>	dataframe containing data
<code>covs</code>	character vector with the names of columns to include in table
<code>maincov</code>	covariate to stratify table by
<code>caption</code>	character containing table caption (default is no caption)
<code>tableOnly</code>	Logical, if TRUE then a dataframe is returned, otherwise a formatted printed object is returned (default).
<code>covTitle</code>	character with the names of the covariate (predictor) column. The default is to leave this empty for output or, for table only output to use the column name 'Covariate'.
<code>digits</code>	number of digits for summarizing mean data
<code>digits.cat</code>	number of digits for the proportions when summarizing categorical data (default: 0)
<code>nicenames</code>	boolean indicating if you want to replace . and _ in strings with a space
<code>IQR</code>	boolean indicating if you want to display the inter quantile range (Q1,Q3) as opposed to (min,max) in the summary for continuous variables
<code>all.stats</code>	boolean indicating if all summary statistics (Q1,Q3 + min,max on a separate line) should be displayed. Overrides IQR.
<code>pvalue</code>	boolean indicating if you want p-values included in the table
<code>effSize</code>	boolean indicating if you want effect sizes included in the table. Can only be obtained if pvalue is also requested. Effect sizes calculated include Cramer's V for categorical variables, Cohen's d, Wilcoxon r, or Eta-squared for numeric/continuous variables.

<code>p.adjust</code>	p-adjustments to be performed. Uses the p.adjust function from base R
<code>unformattedp</code>	boolean indicating if you would like the p-value to be returned unformatted (ie not rounded or prefixed with '<'). Best used with <code>tableOnly = T</code> and <code>outTable</code> function. See examples.
<code>show.tests</code>	boolean indicating if the type of statistical test and effect size used should be shown in a column beside the pvalues. Ignored if <code>pvalue=FALSE</code> .
<code>testcont</code>	test of choice for continuous variables, one of <i>rank-sum</i> (default) or <i>ANOVA</i>
<code>testcat</code>	test of choice for categorical variables, one of <i>Chi-squared</i> (default) or <i>Fisher</i>
<code>full</code>	boolean indicating if you want the full sample included in the table, ignored if <code>maincov</code> is <code>NULL</code>
<code>include_missing</code>	Option to include NA values of <code>maincov</code> . NAs will not be included in statistical tests
<code>percentage</code>	choice of how percentages are presented, one of <i>column</i> (default) or <i>row</i>
<code>dropLevels</code>	logical, indicating if empty factor levels be dropped from the output, default is <code>TRUE</code> .
<code>excludeLevels</code>	a named list of covariate levels to exclude from statistical tests in the form <code>list(varname =c('level1','level2'))</code> . These levels will be excluded from association tests, but not the table. This can be useful for levels where there is a logical skip (ie not missing, but not presented). Ignored if <code>pvalue=FALSE</code> .
<code>numobs</code>	named list overriding the number of people you expect to have the covariate
<code>fontsize</code>	PDF/HTML output only, manually set the table fontsize
<code>chunk_label</code>	only used if output is to Word to allow cross-referencing

Details

Comparisons for categorical variables default to chi-square tests, but if there are counts of <5 then the Fisher Exact test will be used and if this is unsuccessful then a second attempt will be made computing p-values using MC simulation. If `testcont='ANOVA'` then the t-test with unequal variance will be used for two groups and an ANOVA will be used for three or more. The statistical test used can be displayed by specifying `show.tests=TRUE`.

Effect size can be obtained when p-value is requested.

Further formatting options are available using `tableOnly=TRUE` and outputting the table with a call to `outTable`.

A newer version of this function is [rm_compactsum](#) which is more flexible and displays few rows of output.

Value

A character vector of the table source code, unless `tableOnly=TRUE` in which case a data frame is returned

References

Ellis, P.D. (2010) The essential guide to effect sizes: statistical power, meta-analysis, and the interpretation of research results. Cambridge: Cambridge University Press. doi:10.1017/CBO9780511761676

Lakens, D. (2013) Calculating and reporting effect sizes to facilitate cumulative science: a practical primer for t-tests and ANOVAs. *Frontiers in Psychology*, 4; 863:1-12. doi:10.3389/fpsyg.2013.00863

See Also

[covsum](#), [fisher.test](#), [chisq.test](#), [wilcox.test](#), [kruskal.test](#), [anova](#), [rstatix::cramer_v](#), [rstatix::eta_squared](#), and [outTable](#)

Examples

```
data("pembrolizumab")
rm_covsum(data=pembrolizumab, maincov = 'orr',
covs=c('age', 'sex', 'pd11', 'tmb', 'l_size', 'change_ctdna_group'),
show.tests=TRUE)

# To Show Effect Sizes
rm_covsum(data=pembrolizumab, maincov = 'orr',
covs=c('age', 'sex'),
effSize=TRUE)

# To make custom changes or change the fontsize in PDF/HTML
tab <- rm_covsum(data=pembrolizumab,maincov = 'change_ctdna_group',
covs=c('age', 'sex', 'pd11', 'tmb', 'l_size'),show.tests=TRUE,tableOnly = TRUE)
outTable(tab, fontsize=7)

# To return unformatted p-values
tab <- rm_covsum(data=pembrolizumab, maincov = 'orr',
covs=c('age', 'sex', 'pd11', 'tmb', 'l_size', 'change_ctdna_group'),
show.tests=TRUE,unformattedp=TRUE,tableOnly=TRUE)
outTable(tab,digits=5)
outTable(tab,digits=5, applyAttributes=FALSE) # remove bold/indent
```

Description

Multivariable (or univariate) regression models are re-formatted for reporting and a global p-value is added for the evaluation of factor variables.

Multivariable (or univariate) regression models are re-formatted for reporting and a global p-value is added for the evaluation of factor variables.

Usage

```
rm_mvsum(
  model,
  data,
  digits = getOption("reportRmd.digits", 2),
  covTitle = "",
  showN = TRUE,
  showEvent = TRUE,
  CIwidth = 0.95,
  vif = TRUE,
  whichp = c("levels", "global", "both"),
  caption = NULL,
  tableOnly = FALSE,
  p.adjust = "none",
  unformattedp = FALSE,
  nicenames = TRUE,
  chunk_label,
  fontsize
)
```

```
rm_mvsum(
  model,
  data,
  digits = getOption("reportRmd.digits", 2),
  covTitle = "",
  showN = TRUE,
  showEvent = TRUE,
  CIwidth = 0.95,
  vif = TRUE,
  whichp = c("levels", "global", "both"),
  caption = NULL,
  tableOnly = FALSE,
  p.adjust = "none",
  unformattedp = FALSE,
  nicenames = TRUE,
  chunk_label,
  fontsize
)
```

Arguments

model	model fit
data	data that model was fit on (an attempt will be made to extract this from the model)
digits	number of digits to round estimates to, does not affect p-values
covTitle	character with the names of the covariate (predictor) column. The default is to leave this empty for output or, for table only output to use the column name 'Covariate'.

showN	boolean indicating sample sizes should be shown for each comparison, can be useful for interactions
showEvent	boolean indicating if number of events should be shown. Only available for logistic.
CIwidth	width for confidence intervals, defaults to 0.95
vif	boolean indicating if the variance inflation factor should be included. See details
whichp	string indicating whether you want to display p-values for levels within categorical data ("levels"), global p values ("global"), or both ("both"). Irrelevant for continuous predictors.
caption	table caption
tableOnly	boolean indicating if unformatted table should be returned
p.adjust	p-adjustments to be performed. Uses the p.adjust function from base R
unformattedp	boolean indicating if you would like the p-value to be returned unformatted (ie not rounded or prefixed with '<'). Should be used in conjunction with the digits argument.
nicenames	boolean indicating if you want to replace . and _ in strings with a space
chunk_label	only used if output is to Word to allow cross-referencing
fontsize	PDF/HTML output only, manually set the table fontsize

Details

Global p-values are likelihood ratio tests for lm, glm and polr models. For lme models an attempt is made to re-fit the model using ML and if successful LRT is used to obtain a global p-value. For coxph models the model is re-run without robust variances with and without each variable and a LRT is presented. If unsuccessful a Wald p-value is returned. For GEE and CRR models Wald global p-values are returned. For negative binomial models a deviance test is used.

If the variance inflation factor is requested (VIF=T, default) then a generalised VIF will be calculated in the same manner as the car package.

As of version 0.1.1 if global p-values are requested they will be included in the p-value column.

As of R 4.4.0 profile likelihood confidence intervals will be calculated automatically and there is no longer an option to force Wald tests.

The number of decimal places to display the statistics can be changed with digits, but this will not change the display of p-values. If more significant digits are required for p-values then use tableOnly=TRUE and format as desired.

Global p-values are likelihood ratio tests for lm, glm and polr models. For lme models an attempt is made to re-fit the model using ML and if successful LRT is used to obtain a global p-value. For coxph models the model is re-run without robust variances with and without each variable and a LRT is presented. If unsuccessful a Wald p-value is returned. For GEE and CRR models Wald global p-values are returned. For negative binomial models a deviance test is used.

If the variance inflation factor is requested (VIF=T) then a generalised VIF will be calculated in the same manner as the car package.

The number of decimal places to display the statistics can be changed with digits, but this will not change the display of p-values. If more significant digits are required for p-values then use tableOnly=TRUE and format as desired.

Value

A character vector of the table source code, unless tableOnly=TRUE in which case a data frame is returned

A character vector of the table source code, unless tableOnly=TRUE in which case a data frame is returned

References

John Fox & Georges Monette (1992) Generalized Collinearity Diagnostics, Journal of the American Statistical Association, 87:417, 178-183, doi:[10.1080/01621459.1992.10475190](https://doi.org/10.1080/01621459.1992.10475190)

John Fox and Sanford Weisberg (2019). An R Companion to Applied Regression, Third Edition. Thousand Oaks CA: Sage.

John Fox & Georges Monette (1992) Generalized Collinearity Diagnostics, Journal of the American Statistical Association, 87:417, 178-183, doi:[10.1080/01621459.1992.10475190](https://doi.org/10.1080/01621459.1992.10475190)

John Fox and Sanford Weisberg (2019). An R Companion to Applied Regression, Third Edition. Thousand Oaks CA: Sage.

Examples

```
data("pembrolizumab")
glm_fit = glm(change_ctdna_group~sex:age+baseline_ctdna+l_size,
data=pembrolizumab,family = 'binomial')
rm_mvsum(glm_fit)

#linear model with p-value adjustment
lm_fit=lm(baseline_ctdna~age+sex+l_size+tmb,data=pembrolizumab)
rm_mvsum(lm_fit,p.adjust = "bonferroni")
#Coxph
require(survival)
res.cox <- coxph(Surv(os_time, os_status) ~ sex+age+l_size+tmb, data = pembrolizumab)
rm_mvsum(res.cox, vif=TRUE)
data("pembrolizumab")
glm_fit = glm(change_ctdna_group~sex:age+baseline_ctdna+l_size,
data=pembrolizumab,family = 'binomial')
rm_mvsum(glm_fit)

#linear model with p-value adjustment
lm_fit=lm(baseline_ctdna~age+sex+l_size+tmb,data=pembrolizumab)
rm_mvsum(lm_fit,p.adjust = "bonferroni")
#Coxph
require(survival)
res.cox <- coxph(Surv(os_time, os_status) ~ sex+age+l_size+tmb, data = pembrolizumab)
rm_mvsum(res.cox, vif=TRUE)
```

rm_survdiff	<i>Display event counts, expected event counts and logrank test of differences</i>
-------------	--

Description

This is a wrapper function around the `survdiff` function to display overall event rates and group-specific rates along with the log-rank test of a difference in survival between groups in a single table suitable for markdown output. Median survival times are included by default but can be removed setting `median=FALSE`

Usage

```
rm_survdiff(
  data,
  time,
  status,
  covs,
  strata,
  includeVarNames = FALSE,
  digits = 1,
  showCols = c("N", "Observed", "Expected"),
  CIwidth = 0.95,
  conf.type = "log",
  caption = NULL,
  tableOnly = FALSE,
  fontsize
)
```

Arguments

<code>data</code>	data frame containing survival data
<code>time</code>	string indicating survival time variable
<code>status</code>	string indicating event status variable
<code>covs</code>	character vector indicating variables to group observations by
<code>strata</code>	string indicating the variable to stratify observations by
<code>includeVarNames</code>	boolean indicating if the variable names should be included in the output table, default is <code>FALSE</code>
<code>digits</code>	the number of digits in the survival rate
<code>showCols</code>	character vector indicating which of the optional columns to display, defaults to <code>c('N','Observed','Expected')</code>
<code>CIwidth</code>	width of the median survival estimates, default is <code>95%</code>
<code>conf.type</code>	type of confidence interval see survival::survfit for details. Default is 'log'.

caption	table caption
tableOnly	should a dataframe or a formatted object be returned
fontsize	PDF/HTML output only, manually set the table fontsize

Value

A character vector of the survival table source code, unless tableOnly=TRUE in which case a data frame is returned

See Also

[survival::survdiff](#)

Examples

```
#' # Differences between sex
data("pembrolizumab")
rm_survdiff(data=pembrolizumab,time='os_time',status='os_status',
covs='sex',digits=1)

# Differences between sex, stratified by cohort
rm_survdiff(data=pembrolizumab,time='os_time',status='os_status',
covs='sex',strata='cohort',digits=1)
# Differences between sex/cohort groups
rm_survdiff(data=pembrolizumab,time='os_time',status='os_status',
covs=c('sex','cohort'),digits=1)
```

rm_survsum

Summarise survival data by group

Description

Displays event counts, median survival time and survival rates at specified times points for the entire cohort and by group. The logrank test of differences in survival curves is displayed.

Usage

```
rm_survsum(
  data,
  time,
  status,
  group = NULL,
  survtimes = NULL,
  survtimeunit,
  survtimesLbls = NULL,
  CIwidth = 0.95,
  unformattedp = FALSE,
  conf.type = "log",
```



```

na.action = "na.omit",
showCounts = TRUE,
showLogrank = TRUE,
eventProb = FALSE,
digits = getOption("reportRmd.digits", 2),
caption = NULL,
tableOnly = FALSE,
fontsize
)

```

Arguments

<code>data</code>	data frame containing survival data
<code>time</code>	string indicating survival time variable
<code>status</code>	string indicating event status variable
<code>group</code>	string or character vector indicating the variable(s) to group observations by. If this is left as NULL (the default) then summaries are provided for the entire cohort.
<code>survtimes</code>	numeric vector specifying when survival probabilities should be calculated.
<code>survtimeunit</code>	unit of time to suffix to the time column label if survival probabilities are requested, should be plural
<code>survtimesLbels</code>	if supplied, a vector the same length as <code>survtimes</code> with descriptions (useful for displaying years with data provided in months)
<code>CIwidth</code>	width of the survival probabilities, default is 95%
<code>unformattedp</code>	boolean indicating if you would like the p-value to be returned unformatted (ie not rounded or prefixed with '<'). Should be used in conjunction with the <code>digits</code> argument.
<code>conf.type</code>	type of confidence interval see survival::survfit for details. Default is 'log'.
<code>na.action</code>	default is to omit missing values, but can be set to throw an error using <code>na.action='na.fail'</code>
<code>showCounts</code>	boolean indicating if the at risk, events and censored columns should be output; default is TRUE
<code>showLogrank</code>	boolean indicating if the log-rank test statistic and p-value should be output; default is TRUE
<code>eventProb</code>	boolean indicating if event probabilities, rather than survival probabilities, should be displayed; default is FALSE
<code>digits</code>	the number of digits in the survival rate, default is 2, unless the <code>reportRmd.digits</code> option is set
<code>caption</code>	table caption for markdown output
<code>tableOnly</code>	should a dataframe or a formatted object be returned
<code>fontsize</code>	PDF/HTML output only, manually set the table fontsize

Details

This summary table is supplied for simple group comparisons only. To examine differences in groups with stratification see [rm_survdiff](#). To summarise differences in survival rates controlling for covariates see [rm_survtime](#).

Value

A character vector of the survival table source code, unless `tableOnly=TRUE` in which case a data frame is returned

See Also

[survival::survfit](#)

Examples

```
# Simple median survival table
data("pembrolizumab")
rm_survsum(data=pembrolizumab,time='os_time',status='os_status')

# Survival table with yearly survival rates
rm_survsum(data=pembrolizumab,time='os_time',status='os_status',
survtimes=c(12,24),survtimesLbbs=1:2, survtimeunit='yr')

#Median survival by group
rm_survsum(data=pembrolizumab,time='os_time',status='os_status',group='sex')

# Survival Summary by cohort, displayed in years
rm_survsum(data=pembrolizumab,time='os_time',status='os_status',
group="cohort",survtimes=seq(12,72,12),
survtimesLbbs=seq(1,6,1),
survtimeunit='years')

# Survival Summary by Sex and ctDNA group
rm_survsum(data=pembrolizumab,time='os_time',status='os_status',
group=c('sex','change_ctdna_group'),survtimes=c(12,24),survtimeunit='mo')
```

rm_survtime

Display survival rates and events for specified times

Description

This is a wrapper for the `survfit` function to output a tidy display for reporting. Either Kaplan Meier or Cox Proportional Hazards models may be used to estimate the survival probabilities.

Usage

```
rm_survtime(
  data,
  time,
  status,
  covs = NULL,
  strata = NULL,
  type = "KM",
```

```

    survtimes,
    survtimeunit,
    strata.prefix = NULL,
    survtimesLb1s = NULL,
    showCols = c("At Risk", "Events", "Censored"),
    CIwidth = 0.95,
    conf.type = "log",
    na.action = "na.omit",
    showCounts = TRUE,
    digits = getOption("reportRmd.digits", 2),
    caption = NULL,
    tableOnly = FALSE,
    fontsize
)

```

Arguments

data	data frame containing survival data
time	string indicating survival time variable
status	string indicating event status variable
covs	character vector with the names of variables to adjust for in coxph fit
strata	string indicating the variable to group observations by. If this is left as NULL (the default) then event counts and survival rates are provided for the entire cohort.
type	survival function, if no covs are specified defaults to Kaplan-Meier, otherwise the Cox PH model is fit. Use type='PH' to fit a Cox PH model with no covariates.
survtimes	numeric vector specifying when survival probabilities should be calculated.
survtimeunit	unit of time to suffix to the time column label if survival probabilities are requested, should be plural
strata.prefix	character value describing the grouping variable
survtimesLb1s	if supplied, a vector the same length as survtimes with descriptions (useful for displaying years with data provided in months)
showCols	character vector specifying which of the optional columns to display, defaults to c('At Risk','Events','Censored')
CIwidth	width of the survival probabilities, default is 95%
conf.type	type of confidence interval see survival::survfit for details. Default is 'log'.
na.action	default is to omit missing values, but can be set to throw an error using na.action='na.fail'
showCounts	boolean indicating if the at risk, events and censored columns should be output, default is TRUE
digits	the number of digits in the survival rate, default is 2.
caption	table caption for markdown output
tableOnly	should a dataframe or a formatted object be returned
fontsize	PDF/HTML output only, manually set the table fontsize

Details

If covariates are supplied then a Cox proportional hazards model is fit for the entire cohort and each strata. Otherwise the default is for Kaplan-Meier estimates. Setting `type = 'PH'` will force a proportional hazards model.

Value

A character vector of the survival table source code, unless `tableOnly=TRUE` in which case a data frame is returned

See Also

[survival::survfit](#)

Examples

```
# Kaplan-Meier survival probabilities with time displayed in years
data("pembrolizumab")
rm_survtime(data=pembrolizumab,time='os_time',status='os_status',
strata="cohort",type='KM',survtimes=seq(12,72,12),
survtimesLbIs=seq(1,6,1),
survtimeunit='years')

# Cox Proportional Hazards survival probabilities
rm_survtime(data=pembrolizumab,time='os_time',status='os_status',
strata="cohort",type='PH',survtimes=seq(12,72,12),survtimeunit='months')

# Cox Proportional Hazards survival probabilities controlling for age
rm_survtime(data=pembrolizumab,time='os_time',status='os_status',
covs='age',strata="cohort",survtimes=seq(12,72,12),survtimeunit='months')
```

rm_uvsum

Output several univariate models nicely in a single table

Description

#'A table with the model parameters from running separate univariate models on each covariate. For factors with more than two levels a Global p-value is returned.

A table with the model parameters from running separate univariate models on each covariate. For factors with more than two levels a Global p-value is returned.

Usage

```
rm_uvsum(
  response,
  covs,
  data,
```

```
digits = getOption("reportRmd.digits", 2),
covTitle = "",
caption = NULL,
tableOnly = FALSE,
removeInf = FALSE,
p.adjust = "none",
unformattedp = FALSE,
whichp = c("levels", "global", "both"),
chunk_label,
gee = FALSE,
id = NULL,
corstr = NULL,
family = NULL,
type = NULL,
offset = NULL,
strata = 1,
nicenames = TRUE,
showN = TRUE,
showEvent = TRUE,
CIwidth = 0.95,
reflevel = NULL,
returnModels = FALSE,
fontsize,
forceWald = FALSE
)

rm_uvsum(
  response,
  covs,
  data,
  digits = getOption("reportRmd.digits", 2),
  covTitle = "",
  caption = NULL,
  tableOnly = FALSE,
  removeInf = FALSE,
  p.adjust = "none",
  unformattedp = FALSE,
  whichp = c("levels", "global", "both"),
  chunk_label,
  gee = FALSE,
  id = NULL,
  corstr = NULL,
  family = NULL,
  type = NULL,
  offset = NULL,
  strata = 1,
  nicenames = TRUE,
  showN = TRUE,
```

```

    showEvent = TRUE,
    CIwidth = 0.95,
    refllevel = NULL,
    returnModels = FALSE,
    fontsize,
    forceWald = FALSE
  )

```

Arguments

response	string vector with name of response
covs	character vector with the names of columns to fit univariate models to
data	dataframe containing data
digits	number of digits to round estimates and CI to. Does not affect p-values.
covTitle	character with the names of the covariate (predictor) column. The default is to leave this empty for output or, for table only output to use the column name 'Covariate'.
caption	character containing table caption (default is no caption)
tableOnly	boolean indicating if unformatted table should be returned
removeInf	boolean indicating if infinite estimates should be removed from the table
p.adjust	p-adjustments to be performed. Uses the p.adjust function from base R
unformattedp	boolean indicating if you would like the p-value to be returned unformatted (ie not rounded or prefixed with '<'). Should be used in conjunction with the digits argument.
whichp	string indicating whether you want to display p-values for levels within categorical data ("levels"), global p values ("global"), or both ("both"). Irrelevant for continuous predictors.
chunk_label	only used if output is to Word to allow cross-referencing
gee	boolean indicating if gee models should be fit to account for correlated observations. If TRUE then the id argument must specify the column in the data which indicates the correlated clusters.
id	character vector which identifies clusters. Only used for geeglm
corstr	character string specifying the correlation structure. Only used for geeglm. The following are permitted: "independence", "exchangeable", "ar1", "unstructured" and "userdefined"
family	description of the error distribution and link function to be used in the model. Only used for geeglm
type	string indicating the type of univariate model to fit. The function will try and guess what type you want based on your response. If you want to override this you can manually specify the type. Options include "linear", "logistic", "poisson", "coxph", "crr", "boxcox", "ordinal", "geeglm"
offset	string specifying the offset term to be used for Poisson or negative binomial regression. Example: offset="log(follow_up)"

strata	character vector of covariates to stratify by. Only used for coxph and crr
nicenames	boolean indicating if you want to replace . and _ in strings with a space
showN	boolean indicating if you want to show sample sizes
showEvent	boolean indicating if you want to show number of events. Only available for logistic.
CIwidth	width of confidence interval, default is 0.95
reflevel	manual specification of the reference level. Only used for ordinal regression This will allow you to see which model is not fitting if the function throws an error
returnModels	boolean indicating if a list of fitted models should be returned. If this is TRUE then the models will be returned, but the output will be suppressed. In addition to the model elements a data element will be appended to each model so that the fitted data can be examined, if necessary. See Details
fontsize	PDF/HTML output only, manually set the table fontsize
forceWald	[Deprecated] forceWald = TRUE is no longer supported; this function will always use profile likelihoods as per the inclusion of the MASS confidence intervals into base from from R 4.4.0

Details

Global p-values are likelihood ratio tests for lm, glm and polr models. For lme models an attempt is made to re-fit the model using ML and if,successful LRT is used to obtain a global p-value. For coxph models the model is re-run without robust variances with and without each variable and a LRT is presented. If unsuccessful a Wald p-value is returned. For GEE and CRR models Wald global p-values are returned.

As of version 0.1.1 if global p-values are requested they will be included in the p-value column.

The number of decimals places to display the statistics can be changed with digits, but this will not change the display of p-values. If more significant digits are required for p-values then use tableOnly=TRUE and format as desired.

tidyselect can only be used for response and covs variables. Additional arguments must be passed in using characters

Global p-values are likelihood ratio tests for lm, glm and polr models. For lme models an attempt is made to re-fit the model using ML and if,successful LRT is used to obtain a global p-value. For coxph models the model is re-run without robust variances with and without each variable and a LRT is presented. If unsuccessful a Wald p-value is returned. For GEE and CRR models Wald global p-values are returned.

The number of decimals places to display the statistics can be changed with digits, but this will not change the display of p-values. If more significant digits are required for p-values then use tableOnly=TRUE and format as desired.

Value

A character vector of the table source code, unless tableOnly=TRUE in which case a data frame is returned

A character vector of the table source code, unless tableOnly=TRUE in which case a data frame is returned

See Also

[uvsum](#), [lm.glm](#), [cmprsk::crr](#), [survival::coxph](#), [nlme::lme](#), [geepack::geeglm](#), [MASS::glm.nb](#), [covsum](#), [fisher.test](#), [chisq.test](#), [wilcox.test](#), [kruskal.test](#), [anova](#), [rstatix::cramer_v](#), [rstatix:eta_squared](#), and [outTable](#)

Examples

```
# Examples are for demonstration and are not meaningful
# Coxph model with 90% CI
data("pembrolizumab")
rm_uvsum(response = c('os_time', 'os_status'),
covs=c('age', 'sex', 'baseline_ctdna', 'l_size', 'change_ctdna_group'),
data=pembrolizumab, CIwidth=.9)

# Linear model with default 95% CI
rm_uvsum(response = 'baseline_ctdna',
covs=c('age', 'sex', 'l_size', 'pd11', 'tmb'),
data=pembrolizumab)

# Logistic model with default 95% CI
rm_uvsum(response = 'os_status',
covs=c('age', 'sex', 'l_size', 'pd11', 'tmb'),
data=pembrolizumab, family = binomial)
# Poisson models returned as model list
mList <- rm_uvsum(response = 'baseline_ctdna',
covs=c('age', 'sex', 'l_size', 'pd11', 'tmb'),
data=pembrolizumab, returnModels=TRUE)
#'
# GEE on correlated outcomes
data("ctDNA")
rm_uvsum(response = 'size_change',
covs=c('time', 'ctdna_status'),
gee=TRUE,
id='id', corstr="exchangeable",
family=gaussian("identity"),
data=ctDNA, showN=TRUE)

# Using tidyselect
pembrolizumab |> rm_uvsum(response = sex,
covs = c(age, cohort))
# Examples are for demonstration and are not meaningful
# Coxph model with 90% CI
data("pembrolizumab")
rm_uvsum(response = c('os_time', 'os_status'),
covs=c('age', 'sex', 'baseline_ctdna', 'l_size', 'change_ctdna_group'),
data=pembrolizumab, CIwidth=.9)

# Linear model with default 95% CI
rm_uvsum(response = 'baseline_ctdna',
covs=c('age', 'sex', 'l_size', 'pd11', 'tmb'),
data=pembrolizumab)
```



```

# Logistic model with default 95% CI
rm_uvsum(response = 'os_status',
covs=c('age','sex','l_size','pd11','tmb'),
data=pembrolizumab,family = binomial)
# Poisson models returned as model list
mList <- rm_uvsum(response = 'baseline_ctdna',
covs=c('age','sex','l_size','pd11','tmb'),
data=pembrolizumab, returnModels=TRUE)
#'
# GEE on correlated outcomes
data("ctDNA")
rm_uvsum(response = 'size_change',
covs=c('time','ctdna_status'),
gee=TRUE,
id='id', corstr="exchangeable",
family=gaussian("identity"),
data=ctDNA,showN=TRUE)

```

rm_uv_mv

Combine univariate and multivariable regression tables

Description

This function will combine `rm_uvsum` and `rm_mvsum` outputs into a single table. The `tableOnly` argument must be set to `TRUE` when tables to be combined are created. The resulting table will be in the same order as the `uvsum` table and will contain the same columns as the `uvsum` and `mvsum` tables, but the p-values will be combined into a single column. There must be a variable overlapping between the `uvsum` and `mvsum` tables and all variables in the `mvsum` table must also appear in the `uvsum` table.

Usage

```

rm_uv_mv(
  uvsumTable,
  mvsumTable,
  covTitle = "",
  vif = FALSE,
  showN = FALSE,
  showEvent = FALSE,
  caption = NULL,
  tableOnly = FALSE,
  chunk_label,
  fontsize
)

```

Arguments

`uvsumTable` Output from `rm_uvsum`, with `tableOnly=TRUE`

mvsumTable	Output from rm_mvsum, with tableOnly=TRUE
covTitle	character with the names of the covariate (predictor) column. The default is to leave this empty for output or, for table only output to use the column name 'Covariate'.
vif	boolean indicating if the variance inflation factor should be shown if present in the mvsumTable. Default is FALSE.
showN	boolean indicating if sample sizes should be displayed.
showEvent	boolean indicating if number of events (dichotomous outcomes) should be displayed.
caption	table caption
tableOnly	boolean indicating if unformatted table should be returned
chunk_label	only used if output is to Word to allow cross-referencing
fontsize	PDF/HTML output only, manually set the table fontsize

Value

A character vector of the table source code, unless tableOnly=TRUE in which case a data frame is returned

See Also

[rm_uvsum](#), [rm_mvsum](#)

Examples

```
require(survival)
data("pembrolizumab")
uvTab <- rm_uvsum(response = c('os_time', 'os_status'),
  covs=c('age', 'sex', 'baseline_ctdna', 'l_size', 'change_ctdna_group'),
  data=pembrolizumab, tableOnly=TRUE)
mv_surv_fit <- coxph(Surv(os_time, os_status)~age+sex+
  baseline_ctdna+l_size+change_ctdna_group, data=pembrolizumab)
uvTab <- rm_mvsum(mv_surv_fit)

#linear model
uvtab<-rm_uvsum(response = 'baseline_ctdna',
  covs=c('age', 'sex', 'l_size', 'pd11', 'tmb'),
  data=pembrolizumab, tableOnly=TRUE)
lm_fit=lm(baseline_ctdna~age+sex+l_size+tmb, data=pembrolizumab)
mvtab<-rm_mvsum(lm_fit, tableOnly = TRUE)
rm_uv_mv(uvtab, mvtab, tableOnly=TRUE)

#logistic model
uvtab<-rm_uvsum(response = 'os_status',
  covs=c('age', 'sex', 'l_size', 'pd11', 'tmb'),
  data=pembrolizumab, family = binomial, tableOnly=TRUE)
logis_fit<-glm(os_status~age+sex+l_size+pd11+tmb, data = pembrolizumab, family = 'binomial')
mvtab<-rm_mvsum(logis_fit, tableOnly = TRUE)
rm_uv_mv(uvtab, mvtab, tableOnly=TRUE)
```

sanitizestr	<i>Sanitizes strings to not break LaTeX</i>
-------------	---

Description

Strings with special characters will break LaTeX if returned 'asis' by knitr. This happens every time we use one of the main reportRmd functions. We first sanitize our strings with this function to stop LaTeX from breaking.

Usage

```
sanitizestr(str)
```

Arguments

str	a vector of strings to sanitize
-----	---------------------------------

scrolling_table	<i>Output a scrollable table</i>
-----------------	----------------------------------

Description

This function accepts the output of a call to knitr::kable or reportRmd::outTable and, if the output format is html, will produce a scrollable table. Otherwise a regular table will be output for pandoc/latex

Usage

```
scrolling_table(knitrTable, pixelHeight = 500)
```

Arguments

knitrTable	output from a call to knitr::kable or outTable
pixelHeight	the height of the scroll box in pixels, default is 500

Examples

```
data("pembrolizumab")
tab <- rm_covsum(data=pembrolizumab,maincov = 'change_ctdna_group',
covs=c('age','cohort','sex','pd11','tmb','l_size'),full=F)
scrolling_table(tab,pixelHeight=300)
```

set_labels	<i>Set variable labels</i>
------------	----------------------------

Description

Assign variable labels to a data.frame from a lookup table.

Usage

```
set_labels(data, names_labels)
```

Arguments

data	data frame to be labelled
names_labels	data frame with column 1 containing variable names from data and column 2 containing variable labels. Other columns will be ignored.

Details

Useful if variable labels have been imported from a data dictionary. The first column in names_labels must contain the variable name and the second column the variable label. The column names are not used.

If no label is provided then the existing label will not be changed. To remove a label set the label to NA.

See Also

[set_var_labels\(\)](#) for setting individual variable labels, [extract_labels\(\)](#) for creating a data frame of all variable labels, [clear_labels\(\)](#) for removing variable labels

Examples

```
data("ctDNA")
# create data frame with labels
lbls <- data.frame(c1=c('cohort', 'size_change'),
c2=c('Cancer cohort', 'Change in tumour size'))
# set labels and return labelled data frame
set_labels(ctDNA, lbls)
```

set_var_labels	<i>Set variable labels</i>
----------------	----------------------------

Description

Set variable labels for a data frame using name-label pairs.

Usage

```
set_var_labels(data, ...)
```

Arguments

data	data frame containing variables to be labelled
...	Name-label pairs the name gives the name of the column in the output and the label is a character vector of length one.

Details

If no label is provided for a variable then the existing label will not be changed. To remove a label set the label to NA.

See Also

[set_labels\(\)](#) for setting variable labels using a data frame, [extract_labels\(\)](#) for creating a data frame of all variable labels, [clear_labels\(\)](#) for removing variable labels

Examples

```
# set labels using name-label pairs
# and return labelled data frame
data("ctDNA")
ctDNA |> set_var_labels(
  ctdna_status="detectable ctDNA",
  cohort="A cohort label")
```

uvmodels	<i>Aligning models and parameters</i>
----------	---------------------------------------

Description

Aligning models and parameters

Usage

```
uvmodels
```

Format

A data frame with 16 rows and 5 variables:

type model type

family model linking family

gee boolean indicating if gee are used

autoreg_class class of model fit

beta description of output parameter

Source

internal

uvsum

Get univariate summary dataframe

Description

Returns a dataframe corresponding to a univariate regression table

Usage

```
uvsum(  
  response,  
  covs,  
  data,  
  digits = getOption("reportRmd.digits", 2),  
  id = NULL,  
  corstr = NULL,  
  family = NULL,  
  type = NULL,  
  offset = NULL,  
  gee = FALSE,  
  strata = 1,  
  markup = TRUE,  
  sanitize = TRUE,  
  nicenames = TRUE,  
  showN = TRUE,  
  showEvent = TRUE,  
  CIwidth = 0.95,  
  refllevel = NULL,  
  returnModels = FALSE,  
  forceWald  
)
```

Arguments

response	string vector with name of response
covs	character vector with the names of columns to fit univariate models to
data	dataframe containing data
digits	number of digits to round to
id	character vector which identifies clusters. Used for GEE and coxph models.
corstr	character string specifying the correlation structure. Only used for geeglm. The following are permitted: "independence", "exchangeable", "ar1", "unstructured" and "userdefined"
family	specify details of the model used. This argument does not need to be specified and should be used with caution. By default, gaussian errors are used for linear models, the binomial family with logit link is used for logistic regression and poisson with log link is used for poisson regression. This can be specified with the type argument, or will be inferred from the data type. See family . Ignored for ordinal and survival regression and if the type argument is not explicitly specified.
type	string indicating the type of univariate model to fit. The function will try and guess what type you want based on your response. If you want to override this you can manually specify the type. Options include "linear", "logistic", "poisson", "coxph", "crr", "boxcox", "ordinal" and "negbin"
offset	string specifying the offset term to be used for Poisson or negative binomial regression. Example: offset="log(follow_up)"
gee	boolean indicating if gee models should be fit to account for correlated observations. If TRUE then the id argument must specify the column in the data which indicates the correlated clusters.
strata	character vector of covariates to stratify by. Only used for coxph and crr
markup	boolean indicating if you want latex markup
sanitize	boolean indicating if you want to sanitize all strings to not break LaTeX
nicenames	boolean indicating if you want to replace . and _ in strings with a space
showN	boolean indicating if you want to show sample sizes
showEvent	boolean indicating if you want to show number of events. Only available for logistic.
CIwidth	width of confidence interval, default is 0.95
reflevel	manual specification of the reference level. Only used for ordinal. This may allow you to debug if the function throws an error.
returnModels	boolean indicating if a list of fitted models should be returned.
forceWald	boolean indicating if Wald confidence intervals should be used instead of profile likelihood. This is not recommended, but can speed up computations. To use throughout a document use options(reportRmd.forceWald=TRUE)

Details

Univariate summaries for a number of covariates, the type of model can be specified. If unspecified the function will guess the appropriate model based on the response variable.

Confidence intervals are extracted using `confint` where possible. Otherwise Student t distribution is used for linear models and the Normal distribution is used for proportions.

`returnModels` can be used to return a list of the univariate models, which will be the same length as `covs`. The data used to run each model will include all cases with observations on the response and covariate. For gee models the data are re-ordered so that the ids appear sequentially and proper estimates are given.

See Also

[lm,glm,cmprsk::crr,survival::coxph,nlme::lme,geepack::geeglm,MASS::polr,MASS::glm.nb](#)

Index

- * **dataframe**
 - covsum, 5
 - mvsum, 25
 - rm_covsum, 40
 - uvsum, 62
- * **datasets**
 - ctDNA, 8
 - pembrolizumab, 30
 - uvmodels, 61
- * **helper**
 - addspace, 3
 - cap, 4
 - formatp, 16
 - hbld, 24
 - lbld, 24
 - lpvalue, 24
 - nicename, 27
 - niceNum, 28
 - psthr, 33
 - pstprn, 33
 - pvalue, 34
 - rmds, 35
 - sanitizestr, 59
- * **model**
 - boxcoxfitRx, 3
 - crrRx, 7
- * **plot**
 - forestplot2, 10
 - forestplotMV, 11
 - forestplotUV, 13
 - forestplotUVMV, 14
 - plotuv, 31
- addspace, 3
- anova, 7, 43, 56
- boxcoxfitRx, 3
- cap, 4
- chisq.test, 7, 43, 56
- clear_labels, 4
- clear_labels(), 34, 60, 61
- cmprsk::crr, 7, 56, 64
- covsum, 5, 43, 56
- crrRx, 7
- ctDNA, 8
- excelCol, 8
- excelColLetters, 9
- extract_labels, 10
- extract_labels(), 34, 60, 61
- family, 63
- fisher.test, 7, 43, 56
- forestplot2, 10
- forestplotMV, 11
- forestplotUV, 13
- forestplotUVMV, 14
- formatp, 16
- geepack::geeglm, 56, 64
- geoR_boxcoxfit, 16
- ggkmcif, 17
- ggkmcif2, 21
- ggkmcif2Parameters, 22
- ggkmcif_paste, 23
- ggplot2::ggplot, 32
- ggpubr::ggarrange, 32
- glm, 56, 64
- hbld, 24
- kruskal.test, 7, 43, 56
- lbld, 24
- lm, 56, 64
- lpvalue, 24
- MASS::glm.nb, 56, 64
- MASS::polr, 64
- mvsum, 25

nestTable, 26
nicename, 27
niceNum, 28
nlme::lme, 56, 64

outTable, 28, 43, 56

p.adjust, 42, 45, 54
pembrolizumab, 30
plotuv, 31
psthr, 33
pstprn, 33
pvalue, 34

replace_plot_labels, 32, 34
rm_cifsum, 35
rm_compactsum, 37, 42
rm_covsum, 40
rm_mvsum, 43, 58
rm_survdiff, 47, 49
rm_survsum, 48
rm_survtime, 49, 50
rm_uv_mv, 57
rm_uvsum, 31, 52, 58
rmds, 35
rstatix::cramer_v, 43, 56
rstatix::eta_squared, 43, 56

sanitizestr, 59
scrolling_table, 59
set_labels, 60
set_labels(), 34, 61
set_var_labels, 61
set_var_labels(), 34, 60
survival::coxph, 56, 64
survival::survdiff, 48
survival::survfit, 47, 49–52

uvmodels, 61
uvsum, 56, 62

wilcox.test, 7, 43, 56