# Package 'Haplin'

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Title Analyzing Case-Parent Triad and/or Case-Control Data with SNP Haplotypes

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Type Package

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**Depends** R (>= 3.5.0)

Imports tools, mgcv, MASS, ff, rlang, methods

Suggests knitr, Rmpi, ggplot2, testthat, rmarkdown

**Description** Performs genetic association analyses of case-parent triad (trio) data with multiple markers. It can also incorporate complete or incomplete control triads, for instance independent control children. Estimation is based on haplotypes, for instance SNP haplotypes, even though phase is not known from the genetic data. 'Haplin' estimates relative risk (RR + conf.int.) and p-value associated with each haplotype. It uses maximum likelihood estimation to make optimal use of data from triads with missing genotypic data, for instance if some SNPs has not been typed for some individuals. 'Haplin' also allows estimation of effects of maternal haplotypes and parent-of-origin effects, particularly appropriate in perinatal epidemiology. 'Haplin' allows special models, like X-inactivation, to be fitted on the X-chromosome. A GxE analysis allows testing interactions between environment and all estimated genetic effects. The models were originally described in ``Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396".

License GPL (>= 2)

URL https://haplin.bitbucket.io

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Author Hakon K. Gjessing [aut, cre], Miriam Gjerdevik [ctb] (functions 'lineByLine', 'cbindFiles',

'rbindFiles', 'snpPower', 'snpSampleSize', 'hapSim', 'hapRun',

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'hapPower', 'hapPowerAsymp', and 'hapRelEff'), Julia Romanowska [ctb] (<a href="https://orcid.org/0000-0001-6733-1953">https://orcid.org/0000-0001-6733-1953</a>, new data format, parallelisation, new documentation), Oivind Skare [ctb] (TDT tests)

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# **Description**

Takes a sequence of files and combines them column-wise (side-by-side), i.e. reads each file line by line, pastes corresponding lines, then writes to outfile. Combining files line by line is especially useful when working with large datasets, where the reading of entire files may be time consuming and require a large amount of memory.

#### Usage

```
cbindFiles(infiles, outfile, col.sep, ask = TRUE, verbose = TRUE)
```

#### **Arguments**

infiles	A character vector of names (and paths) of the files to combine.
outfile	A character string giving the name of the resulting file. The name of the file is relative to the current working directory, unless the file name contains a definite path.
col.sep	Specifies the column separator which will be inserted between files.
ask	Logical. Default is "TRUE". If set to "FALSE", an already existing outfile will be overwritten without asking.
verbose	Logical. Default is "TRUE", which means that the line number is displayed for each iteration, i.e. each combined line.

# **Details**

The function cbind combines R objects by columns. However, reading large datafiles may require a large amount of memory and be extremely time consuming. cbindFiles combines the files column-wise, one line at a time, and then writes each line to outfile. This avoids reading the full files into memory.

If infiles contains only one file, this file will be copied directly to outfile, i.e. without any line-by-line modifications.

### Value

There is no useful output; the objective of cbindFiles is to produce outfile.

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#### Author(s)

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#### References

```
Web Site: https://haplin.bitbucket.io
```

#### See Also

```
rbindFiles, lineByLine
```

# **Examples**

```
## Not run:

# Combines the three infiles side-by-side
cbindFiles(infiles = c("myfile1.txt", "myfile2.txt",
    "myfile3.txt"), outfile = "myfile_combined_by_columns.txt",
col.sep = " ", ask = TRUE, verbose = TRUE)

## End(Not run)
```

finishParallelRun

Closing the Rmpi cluster

# **Description**

This function closes all the slaves spawned in initParallelRun and finishes the mpi routines. This function MUST BE called after all the haplinSlide calls and right before exiting the script/R session!

# Usage

```
finishParallelRun()
```

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genDataGetPart

Extracting part of genetic data.

# Description

This function enables to extract (and save for later use) part of genetic data read in with gen-DataRead.

# Usage

```
genDataGetPart(
  data.in = stop("No data given!", call. = FALSE),
  design = stop("Design type must be given!"),
  markers,
  indiv.ids,
  rows,
  cc,
  sex,
  file.out = "my_data_part",
  dir.out = ".",
  overwrite = NULL,
  ...
)
```

#### **Arguments**

data.in The data object (in format as the output of genDataRead).

design The design used in the study - choose from:

• triad - (default), data includes genotypes of mother, father and child;

• *cc* - classical case-control;

• cc.triad - hybrid design: triads with cases and controls;

Any of the following can be given to narrow down the dataset:

markers Vector with numbers or names indicating which markers to choose.

indiv.ids Character vector giving IDs of individuals. CAUTION: in a standard PED file,

individual IDs are not unique, so this will select all individuals with given IDs.

rows Numeric vector giving the positions - this will select only these rows.

cc One or more values to choose based on case-control status ('cc' column).

sex One or more values to choose based on the 'sex' column.

 $\label{thm:continuous} \mbox{ file.out } \mbox{ The base for the output filename (default: "my\_data\_part")}.$ 

dir.out The path to the directory where the output files will be saved.

overwrite Whether to overwrite the output files: if NULL (default), will prompt the user

to give answer; set to TRUE, will automatically overwrite any existing files; and

set to FALSE, will stop if the output files exist.

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... If any additional covariate data are available in data.in, the user can choose based on values of these (see the Examples section).

#### **Details**

The genetic data from GWAS studies can be quite large, and thus the analysis is time-consuming. If a user knows where they want to focus the analysis, they can use this function to extract part of the entire dataset and use only this part in subsequent Haplin analysis.

#### Value

A list object with three elements:

- cov.data a data. frame with covariate data (if available in the input file)
- *gen.data* a list with chunks of the genetic data; the data is divided column-wise, using 10,000 columns per chunk; each element of this list is a ff matrix
- aux a list with meta-data and important parameters.

This now contains only the selected subset of data.

#### Warning

No checks are performed when choosing a subset of the data - it is the user's obligation to check whether the data subset contains correct number of individuals (especially important when using the triad design study) and/or markers!

### **Examples**

```
# The argument 'overwrite' is set to TRUE!
# Read the data:
examples.dir <- system.file( "extdata", package = "Haplin" )</pre>
example.file <- file.path( examples.dir, "HAPLIN.trialdata2.txt" )</pre>
my.gen.data.read <- genDataRead( file.in = example.file, file.out = "trial_data",</pre>
 dir.out = tempdir( check = TRUE ), format = "haplin", allele.sep = "", n.vars = 2,
 cov.header = c( "smoking", "sex" ), overwrite = TRUE )
my.gen.data.read
# Extract part with only men:
men.subset <- genDataGetPart( my.gen.data.read, design = "triad", sex = 1,</pre>
 dir.out = tempdir( check = TRUE ), file.out = "gen_data_men_only", overwrite = TRUE )
men.subset
# Extract the part with only smoking women:
women.smoke.subset <- genDataGetPart( my.gen.data.read, design = "triad",</pre>
  dir.out = tempdir( check = TRUE ), sex = 0, smoking = c( 1,2 ), overwrite = TRUE )
women.smoke.subset
```

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genDataLoad

Loading the data previously read in and saved by "genDataRead"

# **Description**

This function loads the data from the saved .ffData and .RData files, and prepares the data to subsequent analysis.

# Usage

```
genDataLoad(filename = stop("'filename' must be given!"), dir.in = ".")
```

# **Arguments**

```
filename The base of the filenames; i.e. if the data is saved in "my_data_gen.ffData",
    "my_data_gen.RData" and "my_data_cov.RData", then the 'filename' should be
    "my_data".

dir.in The path to the directory where files were saved (defaults to the current directory).
```

#### Value

A list object with three elements:

- cov.data a data. frame with covariate data (if available in the input file)
- *gen.data* a list with chunks of the genetic data; the data is divided column-wise, using 10,000 columns per chunk; each element of this list is a ff matrix
- aux a list with meta-data and important parameters.

genDataPreprocess

Pre-processing of the genetic data

# **Description**

This function prepares the data to be used in Haplin analysis

# Usage

```
genDataPreprocess(
  data.in = stop("You have to give the object to preprocess!"),
  map.file,
  map.header = FALSE,
  design = "triad",
  file.out = "data_preprocessed",
  dir.out = ".",
  ncpu = 1,
  overwrite = NULL
)
```

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#### **Arguments**

data.in	Input data, as loaded by genDataRead or genDataLoad.
map.file	Filename (with path if the file is not in current directory) of the .map file holding the SNP names, if available.
map.header	Logical: does the map.file contain a header in the first row? Default: FALSE.
design	The design used in the study - choose from:
	<ul> <li>triad - (default), data includes genotypes of mother, father and child;</li> <li>cc - classical case-control;</li> </ul>
	• cc.triad - hybrid design: triads with cases and controls
file.out	The core name of the files that will contain the preprocessed data (character string); ready to load next time with genDataLoad function; default: "data_preprocessed".
dir.out	The directory that will contain the saved data; defaults to current working directory.
ncpu	The number of CPU cores to use - this speeds up the process for large datasets significantly. Default is 1 core, maximum is 1 less than the total number of cores available on a current machine (even if the number given by the user is more than that).
overwrite	Whether to overwrite the output files: if NULL (default), will prompt the user to give answer; set to TRUE, will automatically overwrite any existing files; and set to FALSE, will stop if the output files exist.

# Value

A list object with three elements:

- cov.data a data. frame with covariate data (if available in the input file)
- *gen.data* a list with chunks of the genetic data; the data is divided column-wise, using 10,000 columns per chunk; each element of this list is a ff matrix
- aux a list with meta-data and important parameters:
  - variables tabulated information of the covariate data;
  - variables.nas how many NA values per each column of covariate data;
  - alleles all the possible alleles in each marker;
  - alleles.nas how many NA values in each marker;
  - nrows.with.missing how many rows contain any missing allele information;
  - which.rows.with.missing vector of indices of rows with missing data (if any)

# **Details**

The .map file should contain at least two columns, where the second one contains SNP names. Any additional columns should be separated by a whitespace character, but will be ignored. The file should contain a header.

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#### **Examples**

```
# The argument 'overwrite' is set to TRUE!
# First, read the data:
examples.dir <- system.file( "extdata", package = "Haplin" )
example.file <- file.path( examples.dir, "exmpl_data.ped" )
ped.data.read <- genDataRead( example.file, file.out = "exmpl_ped_data",
    dir.out = tempdir( check = TRUE ), format = "ped", overwrite = TRUE )
ped.data.read
# Take only part of the data (if needed)
ped.data.part <- genDataGetPart( ped.data.read, design = "triad", markers = 10:12,
    dir.out = tempdir( check = TRUE ), file.out = "exmpl_ped_data_part", overwrite = TRUE )
# Preprocess as "triad" data:
ped.data.preproc <- genDataPreprocess( ped.data.part, design = "triad",
    dir.out = tempdir( check = TRUE ), file.out = "exmpl_data_preproc", overwrite = TRUE )
ped.data.preproc</pre>
```

genDataRead

Reading the genetic data from a file

# **Description**

This function will read in data from PED or haplin formatted file.

# Usage

```
genDataRead(
  file.in = stop("Filename must be given!", call. = FALSE),
  file.out = NULL,
  dir.out = ".",
  format = stop("Format parameter is required!"),
  header = FALSE,
 n.vars,
  cov.file.in,
  cov.header,
 map.file,
 map.header = FALSE,
  allele.sep = ";",
  na.strings = "NA",
  col.sep = "",
  overwrite = NULL
)
```

#### **Arguments**

file.in The name of the main input file with genotype information.

file.out The base for the output filename (by default, constructed from the input file name).

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dir.out	The path to the directory where the output files will be saved.
format	Format of data (will influence how data is processed) - choose from:
	• haplin - data already in one row per family,
	• ped - data from .ped file, each row represents an individual.
header	Whether the first line of the main input file contains column names; default: FALSE; NB: this is useful only for 'haplin'-formatted files!
n.vars	The number of columns with covariate data (if any) in the main file; NB: if the main file is in PED format, it is assumed that the first 6 columns contain the standard PED-covariates (i.e., family ID, ID of the child, father and mother, sex and case-control status), so in this case setting 'n.vars' is useful only if the PED file contains more than 6 covariate columns.
cov.file.in	Name of the file containing additional covariate data, if any. Caution: unless the 'cov.header' argument is used, it is assumed that the first line of this file contains the header (i.e., the column names of the additional data).
cov.header	The character vector containing the names of covariate columns (in the file with additional covariate data if given by the 'cov.file.in' argument; or in the main file, if it's a "haplin"-formatted file).
map.file	Filename (with path if the file is not in current directory) of the .map file holding the SNP names, if available (see Details).
map.header	Logical: does the map.file contain a header in the first row? Default: FALSE.
allele.sep	Character: separator between two alleles (default: ";").
na.strings	Character or NA: how the missing data is coded (default: "NA").
col.sep	Character: separator between the columns (i.e., markers; default: any whitespace character).
overwrite	Whether to overwrite the output files: if NULL (default), will prompt the user to give answer; set to TRUE, will automatically overwrite any existing files; and set to FALSE, will stop if the output files exist.

# **Details**

The function reads in all the data in the file, creates ff objects to store the genetic information and data.frame to store covariate data (if any). These objects are saved in .RData and .ffData files, which can be later on easily uploaded to R (with genDataLoad) and re-used.

#### Value

A list object with three elements:

- cov.data a data.frame with covariate data (if available in the input file)
- *gen.data* a list with chunks of the genetic data; the data is divided column-wise, using 10,000 columns per chunk; each element of this list is a ff matrix
- aux a list with meta-data and important parameters.

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#### **Details**

The .map file should contain at least two columns, where the second one contains SNP names. Any additional columns should be separated by a whitespace character, but will be ignored. The file should contain a header.

#### Usage note

When reading in a covariate file together with the genotype information, it is advised to include the header in the file, so that there is no doubt to the naming of the data columns.

# **Examples**

```
# The argument 'overwrite' is set to TRUE!
examples.dir <- system.file( "extdata", package = "Haplin" )
# ped format:
example.file2 <- file.path( examples.dir, "exmpl_data.ped" )
ped.data.read <- genDataRead( example.file2, file.out = "exmpl_ped_data",
    dir.out = tempdir( check = TRUE ), format = "ped", overwrite = TRUE )
ped.data.read
# haplin format:
example.file1 <- file.path( examples.dir, "HAPLIN.trialdata2.txt" )
haplin.data.read <- genDataRead( file.in = example.file1,
    file.out = "exmpl_haplin_data", format = "haplin", allele.sep = "", n.vars = 2,
    cov.header = c( "smoking", "sex" ), overwrite = TRUE,
    dir.out = tempdir( check = TRUE ) )
haplin.data.read</pre>
```

getChildren

Getter for all rows with children data

#### **Description**

Wrapper function for genDataGetPart that returns a subset of the data containing only children.

# Usage

```
getChildren(
  data.in = stop("No data given!", call. = FALSE),
  file.out = "my_data_onlyChildren",
  dir.out = ".",
  overwrite = NULL
)
```

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# **Arguments**

data.in	The data object (in format as the output of genDataRead); note that the design of the data is assumed to be triad.
file.out	The base for the output filename (default: "my_data_onlyChildren").
dir.out	The path to the directory where the output files will be saved.
overwrite	Whether to overwrite the output files: if NULL (default), will prompt the user to give answer; set to TRUE, will automatically overwrite any existing files; and set to FALSE, will stop if the output files exist.

#### Value

A list object with three elements:

- cov.data a data. frame with covariate data (if available in the input file)
- *gen.data* a list with chunks of the genetic data; the data is divided column-wise, using 10,000 columns per chunk; each element of this list is a ff matrix
- aux a list with meta-data and important parameters.

This now contains only the selected subset of data.

getDyads

Getter only for all dyads (child and one parent)

# **Description**

Wrapper function for genDataGetPart that returns a subset of the data containing only dyads (where the child and only one parent have genetic data), i.e., not triads.

# Usage

```
getDyads(
  data.in = stop("No data given!", call. = FALSE),
  file.out = "my_data_onlyDyads",
  dir.out = ".",
  overwrite = NULL
)
```

# **Arguments**

data.in	The data object (in format as the output of genDataRead); note that the design of the data is assumed to be "triad".
file.out	The base for the output filename (default: "my_data_onlyDyads").
dir.out	The path to the directory where the output files will be saved (default: ".", the current directory).
overwrite	Whether to overwrite the output files: if NULL (default), will prompt the user to give answer; set to TRUE, will automatically overwrite any existing files; and set to FALSE, will stop if the output files exist.

getFathers 13

#### Value

A list object with three elements:

- cov.data a data.frame with covariate data (if available in the input file)
- *gen.data* a list with chunks of the genetic data; the data is divided column-wise, using 10,000 columns per chunk; each element of this list is a ff matrix
- aux a list with meta-data and important parameters.

This now contains only the selected subset of data.

getFathers

Getter for all rows with fathers' data

#### **Description**

Wrapper function for genDataGetPart that returns a subset of the data containing only fathers.

#### Usage

```
getFathers(
  data.in = stop("No data given!", call. = FALSE),
  file.out = "my_data_onlyFathers",
  dir.out = ".",
  overwrite = NULL
)
```

# **Arguments**

data.in	The data object (in format as the output of genDataRead); note that the design of the data is assumed to be triad.
file.out	The base for the output filename (default: "my_data_onlyFathers").
dir.out	The path to the directory where the output files will be saved.
overwrite	Whether to overwrite the output files: if NULL (default), will prompt the user to give answer; set to TRUE, will automatically overwrite any existing files; and set to FALSE, will stop if the output files exist.

# Value

A list object with three elements:

- cov.data a data. frame with covariate data (if available in the input file)
- *gen.data* a list with chunks of the genetic data; the data is divided column-wise, using 10,000 columns per chunk; each element of this list is a ff matrix
- aux a list with meta-data and important parameters.

This now contains only the selected subset of data.

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Getter for all full triads

# Description

Wrapper function for genDataGetPart that returns a subset of the data containing only full triads (where all, the child, the mother and the father have genetic data).

# Usage

```
getFullTriads(
  data.in = stop("No data given!", call. = FALSE),
  file.out = "my_data_onlyTriads",
  dir.out = ".",
  overwrite = NULL
)
```

### **Arguments**

data.in	The data object (in format as the output of genDataRead); note that the design of the data is assumed to be triad.
file.out	The base for the output filename (default: "my_data_onlyTriads").
dir.out	The path to the directory where the output files will be saved.
overwrite	Whether to overwrite the output files: if NULL (default), will prompt the user to give answer; set to TRUE, will automatically overwrite any existing files; and set to FALSE, will stop if the output files exist.

#### Value

A list object with three elements:

- cov.data a data. frame with covariate data (if available in the input file)
- *gen.data* a list with chunks of the genetic data; the data is divided column-wise, using 10,000 columns per chunk; each element of this list is a ff matrix
- aux a list with meta-data and important parameters.

This now contains only the selected subset of data.

getMothers 15

# Description

Wrapper function for genDataGetPart that returns a subset of the data containing only mothers.

# Usage

```
getMothers(
  data.in = stop("No data given!", call. = FALSE),
  file.out = "my_data_onlyMothers",
  dir.out = ".",
  overwrite = NULL
)
```

# **Arguments**

data.in	The data object (in format as the output of genDataRead); note that the design of the data is assumed to be triad.
file.out	The base for the output filename (default: "my_data_onlyMothers").
dir.out	The path to the directory where the output files will be saved.
overwrite	Whether to overwrite the output files: if NULL (default), will prompt the user to give answer; set to TRUE, will automatically overwrite any existing files; and set to FALSE, will stop if the output files exist.

#### Value

A list object with three elements:

- cov.data a data. frame with covariate data (if available in the input file)
- *gen.data* a list with chunks of the genetic data; the data is divided column-wise, using 10,000 columns per chunk; each element of this list is a ff matrix
- *aux* a list with meta-data and important parameters.

This now contains only the selected subset of data.

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gxe

Test for gene-environment interaction

#### **Description**

Performes a gene-environment test to check if haplin estimates of relative risks change over strata of environment. It is typically applied to the output from haplinStrat

#### Usage

```
gxe(object.list)
```

# **Arguments**

object.list

A list of haplin results, almost always the output from haplinStrat. The first element is the result of running haplin on all data; the remaining elements are the results for each stratum separately.

#### **Details**

haplinStrat runs haplin first on the entire input data file, then on each stratum separately. The results from haplinStrat are similar to just manually splitting the file into strata and running haplin on each, with one important difference, however: Since some strata may be small etc., haplin might conceivably choose different haplotypes in different strata, and also choose different reference haplotypes. When first running haplin on the entire file, haplinStrat saves the selected haplotypes and chosen reference category. Then, in the strata-specific runs haplinStrat forces haplin to choose the same haplotypes/reference category in all runs, so that results from different strata are comparable. When applying gxe to the output from haplinStrat, it will test whether there is a statistically significant change in parameter estimates from stratum to stratum, i.e. a gene-environment interaction since strata usually are defined by an environmental exposure. gxe uses Wald tests to test for interactions. It always tests whether there is change in haplotype frequencies from stratum to stratum. More importantly, it separately tests whether any genetic effects, such as fetal genetic effects, maternal effects, or parent-of-origin effects, change significantly over strata. gxe can also be run from within haplinSlide by using the strata argument in haplinSlide.

#### Value

A dataframe with one row for each test that is performed (haplo.freq is the first, the remaining depend on the model that has been estimated). The Wald chi-squared test value, degrees-of-freedom, and resulting p-value are reported.

#### NOTE:

In the future, the structure of the output from gxe will change. In particular, measures of ratios of relative risks will be reported in addition to the p-values

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#### Note

Further information is found on the web page.

# Author(s)

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#### References

Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396.

```
Web Site: https://haplin.bitbucket.io
```

#### See Also

```
haplin, haplinStrat, haplinSlide
```

# **Examples**

```
# All standard haplin runs can be done with haplinStrat.
# Below is an illustration. See the haplin help page for more
# examples.
# Analyzing the effect of fetal genes, including triads with missing data,
# using a multiplicative response model. The first column of the data file
# in this example contains the stratification variable.
result <- haplinStrat("C:/work/data.dat", strata = 1, use.missing = T, response = "mult",</pre>
reference = "ref.cat", winlength = 1)
# Provide summary of separate results:
lapply(result, summary)
# Plot results separately:
par(ask = T)
lapply(result, plot)
# Convert results to table format and stack them over strata:
haptable(result)
# Test for interaction between haplotype risk estimates and the strata variable:
gxe(result)
## End(Not run)
```

haplin

Fitting log-linear models to case-parent triad and/or case-control data

#### **Description**

haplin fits a log-linear model to case-parent triads, case-control data, or combined (hybrid) case-parent control-parent triads or dyads. It estimates marker or haplotype frequencies, and uses the EM algorithm to reconstruct haplotypes and, if requested, impute missing genotypes. haplin prints and plots estimates of relative risks associated with fetal and maternal haplotypes, and in addition allows splitting fetal haplotype effects into maternally and paternally inherited effects. It allows special models, like x-inactivation, to be fitted on the X-chromosome. The result is an object of class haplin, which can be explored with summary, plot, and haptable.

# Usage

```
haplin( data, markers = "ALL",
design = "triad", use.missing = FALSE,
xchrom = FALSE, maternal = FALSE, test.maternal = FALSE,
poo = FALSE, scoretest = "no", ccvar = NULL, strata = NULL,
sex = NULL, comb.sex = "double",
reference = "reciprocal", response = "free",
threshold = 0.01, max.haplos = NULL, haplo.file = NULL,
resampling = "no", max.EM.iter = 50, data.out = "no",
verbose = TRUE, printout = TRUE)
```

#### Arguments

data

An R-object which is the result of using genDataPreprocess. See the web page for a detailed description of how to use this function.

markers

Default is "ALL", which means haplin uses all available markers in the data set in the analysis. For the current version of haplin the number of markers used at a single run should probably not exceed 4 or 5 due to the computational burden. The markers argument can be used to select appropriate markers from the file without creating a new file for the selected markers. The relevant markers can be specified by giving a vector or numbers (e.g., markers = c(1, 3:10)) will use the 10 first markers except marker 2) or characters (e.g., markers = c("m1", "m3", "rs35971")). When running haplin, it may be a good idea to start exploring a few markers at a time, using this argument.

design

The value "triad" is used for the standard case triad design, without independent controls. The value "cc.triad" means a combination of case triads and control triads. This requires the argument ccvar to point to the data column containing the case-control variable. The value "cc" means a simple case-control design, where the parents have not been genotyped (there are no data columns for parental genes). NOTE: design is also set in genDataPreprocess. Almost always, the two arguments should be equal. Occasionally, however, the user might want to override the original argument by switching from 'cc.triad' to 'triad' or vice versa.

. .

use.missing

A logical value used to determine whether triads with missing data should be included in the analysis. When set to TRUE, haplin uses the EM algorithm to obtain risk estimates, also taking into account triads with missing data. The standard errors and p-values are adjusted to correct for this. The default, however, is FALSE. When FALSE, all triads having any sort of missing data are excluded before the analysis is run. Note that haplin only looks at markers actually used in the analysis, so that if the markers argument (see below) is used to select a collection of markers for analysis, haplin only excludes triads with missing data on the included markers.

xchrom

Logical, defaults to "FALSE". If set to "TRUE", haplin assumes the markers are on the x-chromosome. This option should be combined with specifying the sex argument. In addition, comb.sex can be useful. xchrom = T can be combined with poo = T and/or maternal = T.

maternal

If TRUE, maternal effects are estimated as well as the standard fetal effects.

test.maternal

Not yet implemented.

poo

Parent-Of-Origin effects. If TRUE, haplin will split single-dose effects into two separate effect estimates, one for the maternally inherited haplotype, and one for the paternally inherited haplotype. Double dose will be estimated as before.

scoretest

Special interest only. If "no", no score test is computed. If "yes", an overall score p-value is included in the output, and the individual score values are returned in the haplin object. If "only", haplin is only run under the null hypothesis, and a simple score object is returned instead of the full haplin object. Useful if only score testing is needed.

ccvar

Numeric. Should give the column number for the column containing the case-control indicator in the data file. Needed for the "cc" and "cc.triad" designs. The column should contain two numeric values, of which the largest one is always used to denote cases.

strata

Not yet implemented.

sex

To be used with xchrom = TRUE. A numeric value specifying which of the data columns that contains the sex variable. The variable should be coded 1 for males and 2 for females.

comb.sex

To be used with xchrom = TRUE. A character value that specifies how to handle gender differences on the X-chromosome. If set to "males" or "females", analyses are done either for just males or just females, respectively. If set to "single" or "double", males and females are used in a combined analysis. Specifically, when "single", the effect of a (single) allele in males is assumed to equal the effect of a single allele dose in females, and similarly, when "double", a single allele in males is assumed to have the same effect as a double allele dose in females. Default is "double", which corresponds to X-inactivation. See separate description for more details.

reference

Decides how haplin chooses its reference category for the effect estimates. Default value is "reciprocal". With the reciprocal reference the effect of a single or double dose of each haplotype is measured relative to the remaining haplotypes. This means that a new reference category is used for each single haplotype. Other possible values are "population" (which is similar to reciprocal, but where

the reference category is always the total population), and "ref.cat", where a single haplotype is used as reference for all the rest. For ref.cat, the default is to choose the most frequent haplotype as the reference haplotype. The reference haplotype can be set explicitly by giving a numeric value for the reference argument. Note that the numeric value refers to the haplotype's position among the haplotypes selected for analysis by haplin. This means that one should run haplin once first to see what haplotypes are used before giving a numeric value to reference.

response

The default value "free" means that both single- and double dose effects are estimated. Choosing "mult" instead specifies a multiplicative dose-response model.

threshold

Sets the (approximate) lower limit for the haplotype frequencies of those haplotypes that should be retained in the analysis. Hapotypes that are less frequent are removed, and information about this is given in the output. Default is 0.01.

max.haplos
haplo.file

Not yet implemented. Not yet implemented.

resampling

Mostly for testing. Default is "no". When "no", the individual haplotypes reconstructed by the EM algorithm as assumed known when computing CIs and p-values. If set to "jackknife" a jackknife-based resampling procedure is used when computing confidence intervals and p-values for effect estimates. This takes more time, but corrects the CIs and p-values for the uncertainty contained in unphased data. Note: in all recent versions of haplin, the resampling is no longer needed since the confidence intervals and p-values are already corrected in the standard computation.

max.EM.iter

The maximum number of iterations used by the EM algorithm. This value can be increased if necessary, which sometimes is the case with e.g. case-control data which a substantial amount of missing. However, for triad data with little missing information there is usually no need for many iterations.

data.out

Character. Accepts values "no", "prelim", "null" or "full", with "no" as default. For values other than default, haplin returns the data file prepared for analysis rather than the usual haplin estimation results. The data file contains the haplotypes identified for each triad, and a vector of weights giving the probability distribution of different haplotype configurations within a triad. The probabilities are computed from preliminary haplotype frequency estimates, from the null model or from the full likelihood model. The "prelim" option will be much faster but somewhat less precise than the likelihood models.

verbose

Default is T (=TRUE). During the EM algorithm, haplin prints the estimated parameters and deviance for each step. To avoid the output, set this argument to F (=FALSE).

printout

Logical. If TRUE (default), haplin prints a full summary of the results after finishing the estimation. If FALSE, no such printout is given, but the summary function can later be applied to a saved result to get the same summary.

#### **Details**

Input data can be either a haplin format data file, or a PED data. These have to be loaded into R first, using genDataRead or genDataLoad functions, and then pre-processed with the genDataPre-process function. If the PED data file is used, the arguments filename, n.vars, sep, allele.sep,

na.strings, ccvar, and sex need not be specified. The output can be examined by print, summary, plot and haptable.

#### Value

An object of class haplin is returned. (The only exception is when data.out is set different from "no", where haplin will produce a data file with haplotypes identified.)

# Warning

Typically, some of the included haplotypes will be relatively rare, such as a frequency of 1% - 5%. For those haplotypes there may be too little data to estimate the double doses properly, so the estimates may be unreliable. This is seen from the extremely wide confidence intervals. The rare double dose estimates should be disregarded, but the remaining single and double dose estimates are valid. To avoid the problem one can also reduce the model to a purely multiplicative model by setting response = "mult" combined with reference = "ref.cat".

#### Note

Further information is found on the web page.

#### Author(s)

Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health <a href="hakon.gjessing@uib.no">hakon.gjessing@uib.no</a>

#### References

Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396.

```
Web Site: https://haplin.bitbucket.io
```

# See Also

summary.haplin, plot.haplin, pedToHaplin, haptable, haplinSlide, genDataLoad, genDataRead, genDataPreprocess

### **Examples**

```
# setting up the directory with exemplary data
dir.in <- system.file( "extdata", package = "Haplin" )
file.in <- file.path( dir.in, "data.dat" )

# reading data in
data.in <- genDataRead( file.in, file.out = "poo_exmpl_data_read", format = "haplin",
    dir.out = tempdir( check = TRUE ), n.vars = 1, allele.sep = " ", col.sep = " ",
    overwrite = TRUE )</pre>
```

```
# preprocessing the data
data.preproc <- genDataPreprocess( data.in, design = "triad",</pre>
 file.out = "poo_exmpl_data_preproc", dir.out = tempdir( check = TRUE ), overwrite = TRUE )
# running haplin, calculating POO
res.POO <- haplin( data.preproc, markers = 2, poo = TRUE, response = "mult",
  reference = 2, use.missing = TRUE )
res.P00
## Not run:
# 1. Read the data:
my.haplin.data <- genDataRead( file.in = "HAPLIN.trialdata.txt", file.out =</pre>
  "trial_data1", dir.out = ".", format = "haplin", n.vars = 0 )
# 2. Run pre-processing:
haplin.data.prep <- genDataPreprocess( data.in = my.haplin.data, format =</pre>
  "haplin", design = "triad", file.out = "trial_data1_prep", dir.out = ".")
# 3. Analyze:
# Standard run:
haplin( haplin.data.prep )
# Specify path, estimate maternal effects:
haplin( haplin.data.prep, maternal = T )
# Specify path, use haplotype no. 2 as reference:
haplin( haplin.data.prep, reference = 2 )
# Remove more haplotypes from estimation by increasing the threshold
# to 5%:
haplin( haplin.data.prep, threshold = 0.05 )
# Estimate maternal effects, using the most frequent haplotype as reference.
# Use all data, including triads with missing data. Select
# markers 3, 4 and 8 from the supplied data.
haplin( haplin.data.prep, use.missing = T, maternal = T,
reference = "ref.cat", markers = c(3,4,8))
# Note: in this version of haplin, the jackknife is
# no longer necessary since the standard errors are already corrected.
# Some examples showing how to save the haplin result and later
# recall plot and summary results:
# Same analysis as above, saving the result in the object "result.1":
result.1 <- haplin( haplin.data.prep, use.missing = T, maternal = T,
reference = "ref.cat", markers = c(3,4,8))
# Replot the saved result (fetal effects):
plot( result.1 )
# Replot the saved result (maternal effects):
plot( result.1, plot.maternal = T )
```

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```
# Print a very short summary of saved result:
result.1
# A full summary of saved result, with confidence intervals and
# p-values (the same as haplin prints when running):
summary( result.1 )
# Some examples when the data file contains two covariates,
# the second is the case-control variable:
# The following standard triad run is INCORRECT since it disregards
# case status:
haplin("data.dat", use.missing = T, n.vars = 2, design = "triad")
# Combined run on "hybrid" design, correctly using both case-parent
# triads and control-parent triads:
haplin( my.haplin.data, use.missing = T, n.vars = 2, ccvar = 2,
design = "cc.triad" )
# If parent columns are not in the file, a plain case-control
# run can be used:
haplin( my.haplin.data, use.missing = T, n.vars = 2, ccvar = 2,
design = "cc", response = "mult", reference = "ref.cat" )
# An example of how to produce a data file with all possible haplotypes
# identified for each triad, together with their probaility weights:
result.data <- haplin( my.haplin.data, use.missing = T,
markers = c(3,4,8), data.out = "prelim")
# result.data will then contain the data file, with a vector of
# probabilities (freq) computed from the preliminary haplotype
# frequencies.
## End(Not run)
```

haplinSlide

Run haplin analysis in a series of sliding windows over a sequence of markers/SNPs

# Description

Produces a list, each element of which is an object of class haplin, which is the result of fitting the log-linear haplin models to the data one "window" at a time.

# Usage

```
haplinSlide( data, markers = "ALL", winlength = 1,
strata = NULL, table.output = TRUE, cpus = 1, para.env = NULL, slaveOutfile = "",
printout = FALSE, verbose = FALSE, ...)
```

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#### **Arguments**

data R-object of class "haplin.ready", which is e.g., output from genDataPreprocess or genDataLoad, and contains covariate and genetic data. markers Default is "ALL", which means haplinSlide uses all available markers in the data set in the analysis. Alternatively, the relevant markers can be specified by giving a vector or numbers (e.g., markers = c(1, 3:10) will use the 10 first markers except marker 2) or characters (e.g., markers = c("m1", "m3", "rs35971")). haplinSlide will then run haplin on a series of windows selected from the supplied markers. The winlength argument decides the length of the windows. See details. winlength Length of the sliding, overlapping windows to be run along the markers. See details. A single numeric value specifying which data column contains the stratification strata variable. table.output If TRUE, the haptable function will be applied to each result after estimation, greatly reducing the size of the output. If FALSE, each element of the output list is a standard haplin object. To preserve memory, default is set to TRUE. haplinSlide allows parallel processing of its analyses. The cpus argument cpus should preferably be set to the number of available cpu's. If set lower, it will save some capacity for other processes to run. Setting it too high should not cause any serious problems. The user can choose parallel environment to use — "parallel" (default) or "Rmpi" para.env (for use on clusters); this option is used only when cpus argument is larger than slaveOutfile Character. To be used when cpus > 1. If slaveOutfile = "" (default), output from all running cores will be printed in the standard R session window. Alternatively, the output can be saved to a file by specifying the file path and name. Default is FALSE. If TRUE, provides a full summary of each haplin result printout during the run of haplinSlide. verbose Same as for haplin, but defaults to FALSE to reduce output size.

#### **Details**

. . .

haplinSlide runs haplin on a series of overlapping windows of the chosen markers. Except for the markers and winlength arguments, all arguments are used exactly as in haplin itself. For instance, if markers = c(1, 3, 4, 5, 7, 8) and winlength = 4, haplinSlide will run haplin on first the markers c(1, 3, 4, 5), then on c(3, 4, 5, 7), and finally on c(4, 5, 7, 8). The results are returned in a list. The elements are named "1-3-4-5" etc., and can be extracted with, say, summary(res[["1-3-4-5"]]) etc., where res is the saved result. Or the output can be examined by, for instance, using lapply(res, summary) and lapply(res, plot).

Remaining arguments to be used by haplin in each run.

When running haplinSlide on a large number of markers, the output can become prohibitively large. In that case table.output should be set to TRUE, and haplinSlide will return a list of summary "haptables". This list can then be stacked into a single dataframe using toDataFrame. To avoid exessive memory use, the default is table.output = TRUE.

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When multiple cores are available, set the cpus to the number of cores that should be used. This will run haplinSlide in parallel on the chosen number of cores. Note that feedback is provided by each of the cores separately, and some cores may start working on markers far out in the sequence.

#### Value

A list of objects of class haplin is returned.

#### Note

Further information is found on the web page.

# Author(s)

Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health <a href="hakon.gjessing@uib.no">hakon.gjessing@uib.no</a>

#### References

Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396.

```
Web Site: https://haplin.bitbucket.io
```

#### See Also

haplin, summary.haplin, plot.haplin, haptable, toDataFrame

# **Examples**

```
## Not run:
# (Almost) all standard haplin runs can be done with haplinSlide.
# Below is an illustration. See the haplin help page for more
# examples.
#
# 1. Read the data:
my.haplin.data <- genDataRead( file.in = "HAPLIN.trialdata.txt", file.out =
    "trial_data1", dir.out = tempdir( check = TRUE ), format = "haplin", n.vars = 0 )
# 2. Run pre-processing:
haplin.data.prep <- genDataPreprocess( data.in = my.haplin.data,
    format = "haplin", design = "triad", file.out = "trial_data1_prep",
    dir.out = tempdir( check = TRUE ) )
# 3. Analyze:
# Analyzing the effect of fetal genes, including triads with missing data,
# using a multiplicative response model. When winlength = 1, separate</pre>
```

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```
# markers are used. To make longer windows, winlength can be increased
# correspondingly:
result.1 <- haplinSlide( haplin.data.prep, use.missing = T, response = "mult",
reference = "ref.cat", winlength = 1, table.output = F)
# Provide summary of separate results:
lapply(result.1, summary)
# Plot results:
par(ask = T)
lapply(result.1, plot)
## End(Not run)</pre>
```

haplinStrat

Fit haplin to each subset/stratum of data, determined by the argument strata

# **Description**

Produces a list, each element of which is an object of class haplin, which is the result of fitting the log-linear haplin models to each strata stratum independently.

#### **Usage**

```
haplinStrat( data, strata = NULL, ...)
```

#### **Arguments**

data	R-object of class "haplin.ready", which is e.g., output from genDataPreprocess or genDataLoad, and contains covariate and genetic data.
strata	A single integer specifying the number of the column in the covariate data that contains the stratification variable
	Remaining arguments to be used by haplin in each run.

#### **Details**

haplinStrat runs haplin first on the entire input data file, then on each stratum separately. Strata are defined by the strata variable, which can be coded as numerical or character. However, one should use only a moderate number of levels/strata, since haplin will be run independetly on each, and some strata may otherwise have an insufficient amount of data. Running haplinStrat is thus just a simplification of manually splitting the file into strata and running haplin on each; the end result would be the same. The main reason for running haplinStrat is to test for gene-environment interactions. This is achieved by running postTest on the result from haplinStrat. haplinStrat can also be run from within haplinSlide by using the strata argument in haplinSlide.

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#### Value

A list of objects of class haplin is returned. The first element contains the result of running haplin on the entire data file; the remaining elements are the results from each of the strata. The names of the list correspond to the values of the strata variable.

#### Note

Further information is found on the web page.

#### Author(s)

Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health <hakon.gjessing@uib.no>

#### References

Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396.

```
Web Site: https://haplin.bitbucket.io
```

#### See Also

haplin, summary.haplin, plot.haplin, haptable, toDataFrame, haplinSlide, gxe

#### **Examples**

```
# setting up the directory with exemplary data
dir.in <- system.file( "extdata", package = "Haplin" )</pre>
file.in <- paste0( dir.in, "/data.dat" )</pre>
# reading data in
data.in <- genDataRead( file.in, file.out = "poo_exmpl_data_read", format = "haplin",</pre>
  dir.out = tempdir( check = TRUE ), n.vars = 1, allele.sep = " ", col.sep = " ",
  overwrite = TRUE )
# preprocessing the data
data.preproc <- genDataPreprocess( data.in, design = "triad",</pre>
 file.out = "poo_exmpl_data_preproc", dir.out = tempdir( check = TRUE ), overwrite = TRUE )
# running haplinStrat, checking for gene-environment interactions
res.GxE <- haplinStrat( data.preproc, markers = 2, strata = 1, poo = FALSE,
  response = "mult", reference = 2, use.missing = TRUE )
res.GxE
# running haplinStrat, checking for POO-environment interactions
res.PO0xE <- haplinStrat( data.preproc, markers = c(1,2,3), strata = 1, poo = TRUE,
  response = "mult", reference = "ref.cat", use.missing = TRUE )
res.P00xE
```

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```
## Not run:
# All standard haplin runs can be done with haplinStrat.
# Below is an illustration. See the haplin help page for more
# examples.
# Analyzing the effect of fetal genes, including triads with missing data,
# using a multiplicative response model. The first column of the data file
# in this example contains the stratification variable.
result <- haplinStrat("C:/work/data.dat", strata = 1, use.missing = T, response = "mult",
reference = "ref.cat", winlength = 1)
# Provide summary of separate results:
lapply(result, summary)
# Plot results separately:
par(ask = T)
lapply(result, plot)
# Convert results to table format and stack them over strata:
haptable(result)
# Test for interaction between haplotype risk estimates and the strata variable:
postTest(result)
## End(Not run)
```

hapPower

Power simulation for association analyses with Haplin

#### **Description**

Simulates the statistical power of genetic analyses assessing fetal effects, maternal effects and/or parent-of-origin effects. Effects of X-chromosome genes and gene-environment interaction effects are also allowed.

# Usage

```
hapPower(hapRun.result, alpha = 0.05)
```

#### **Arguments**

```
hapRun.result The result of running hapRun alpha alpha is the Type I Error probablity. Equals 0.05 by default.
```

# **Details**

The Haplin framework includes different modules for assessing genetic effects: haplin, haplinStrat and haplinSlide. hapPower simulates the power of these analyses, which enables power calculations of fetal effects, maternal effects and/or parent-of-origin effects. Various family designs, i.e.,

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triads, case-control, the hybrid design, and all intermediate designs, are possible. It also allows power calculation of gene-environment interaction effects and effects on X-chromosome markers.

hapPower calculates statistical power using the result of hapRun, and the target effects must be specified in this function, see Examples below, and details in https://haplin.bitbucket.io/docu/Haplin\_power.pdf.

#### Value

hapPower returns the simulated power.

#### Author(s)

```
Miriam Gjerdevik,
with Hakon K. Gjessing
Professor of Biostatistics
Division of Epidemiology
Norwegian Institute of Public Health
<a href="https://doi.org/10.1007/j.nc">https://doi.org/10.1007/j.nc</a>
```

### References

```
Web Site: https://haplin.bitbucket.io
```

#### See Also

haplin, haplinSlide, hapSim, hapRun, snpPower, snpSampleSize, hapPowerAsymp

#### **Examples**

```
## Not run:
## Simulate power from 100 files using haplin.
## The files consist of fetal effects at two diallelic markers,
## corresponding to haplo.freq = rep(0.25, 4), RR = c(2,1,1,1) and RRstar = c(1,1,1,1).
## The power is simulated for the combination of 100 case triads
## and 100 control triads with no missing data at a 0.05 significance level,
## applying a multiplicative model.
hapRun.res <- hapRun(nall = c(2,2), n.strata = 1, cases = c(mfc=100), controls = c(mfc=100),
haplo.freq = rep(0.25,4), RR = c(2,1,1,1), RRstar = c(1,1,1,1),
hapfunc = "haplin", response = "mult", n.sim = 100, dire = "simfiles", ask = FALSE)
hapPower(hapRun.res)
## Simulate power from 100 files applying haplinStrat.
## The files consist of fetal and maternal effects at two diallelic markers.
## The data is simulated for 500 case triads and 200 control families in the first stratum,
## and 500 case triads and 500 control trids in the second.
## The fetal effects vary across strata,
## whereas the maternal effects are the same.
## One percent of the case triads are missing at random in the second stratum.
hapRun.res <- hapRun(nall = c(2,2), n.strata = 2, cases = c(mfc=500),
controls = list(c(mfc=200), c(mfc=500)), haplo.freq = rep(0.25,4), maternal = TRUE,
```

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```
RR = list(c(1.5,1,1,1),c(1,1,1,1)), RRstar = c(1,1,1,1),
RR.mat = c(1.5,1,1,1), RRstar.mat = c(1,1,1,1), gen.missing.cases = list(NULL,0.01),
use.missing = TRUE, hapfunc = "haplinStrat", n.sim = 100, ask = FALSE)
hapPower(hapRun.res)
## Simulate power at the 0.1 significance level from 1000 files using haplin.
## The files consist of fetal effects at one diallelic locus,
## corresponding to haplo.freq = c(0.1,0.9), RR = c(2,1) and RRstar = c(1,1).
## The data consists of a combination of 100 case triads and 100 control triads.
hapRun.res <- hapRun(nall = c(2), cases = c(mfc=100), controls = c(mfc=100),
haplo.freq = c(0.1,0.9), RR = c(2,1), RRstar = c(1,1),
hapfunc = "haplin", response = "mult", n.sim = 1000, ask = FALSE)
hapPower(hapRun.res, alpha= 0.10)
## The latter example, applying response = "mult", should be comparable to
## the theoretic calculations of snpPower.
snpPower(cases = list(mfc=100), controls = list(mfc=100),
RR = 2, MAF = 0.1, alpha = 0.10)
## End(Not run)
```

hapPowerAsymp

Asymptotic power calculations for genetic association analyses with Haplin

# Description

Computes the asymptotic power for genetic analyses assessing fetal effects, maternal effects and/or parent-of-origin effects. Effects of X-chromosome genes and gene-environment interaction effects are also allowed.

# Usage

```
hapPowerAsymp(nall = 2, n.strata = 1, cases, controls, haplo.freq,
RR, RRcm, RRcf, RRstar, RR.mat, RRstar.mat,
xchrom = F, sim.comb.sex = "double", BR.girls,
response = "mult", alpha = 0.05, ...)
```

# **Arguments**

cases

nall A vector of the number of alleles at each locus. By default a diallellic SNP.

n.strata The number of strata.

A list of the number of case families. Each element is a vector of the number of families of the specified family design in the corresponding stratum. The possible family designs, i.e., the possible names of the elements, are "mfc" (full triad), "mc" (mother-child dyad), "fc" (father-child dyad) or "c" (a single case child).

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controls A list of the number of control families. Each element is a vector of the number of families of the specified family design in the corresponding stratum. The

possible family designs are "mfc" (full triad), "mc" (mother-child-dyad), "fc" (father-child dyad), "mf" (mother-father dyad), "c" (a single control child), "m"

(a single control mother) or "f" (a single control father).

haplo.freq A list of which each element is a numeric vector of the haplotype frequencies

in each stratum. The frequencies are normalized and sum to one. The Details section shows how to implement this argument in agreement with the possible

haplotypes.

RR A list of which each element is a numeric vector of the relative risks in each stra-

tum. The Details section shows how to implement this argument in agreement

with the possible haplotypes.

RRcm A list of numeric vectors. Each vector contains the relative risks associated with

the haplotypes transmitted from the mother for this stratum. See Details for description of how to implement this argument in agreement with the possible

haplotypes.

RRcf A list of numeric vectors. Each vector contains the relative risks associated

with the haplotypes transmitted from the father for this stratum. See Details for description of how to implement this argument in agreement with the possible

haplotypes.

RRstar A list of numeric vectors. Estimates how much double-dose children would

deviate from the risk expected in a multiplicative dose-response relationship.

RR.mat The interpretation is similar to RR but for maternal genetic effects.

RRstar.mat The interpretation is similar to RRstar but for maternal genetic effects.

xchrom Logical. Equals FALSE by default, which indicates analyses of autosomal mark-

ers. If TRUE, hapPowerAsymp performs power analyses of X-linked markers.

sim.comb.sex To be used with xchrom = TRUE. A character value that specifies how to handle

gender differences on the X-chromosome. If "single", the effect of a (single) allele in males is equal to the effect of a single allele dose in females, and similarly, if "double", a single allele in males has the same effect as a double allele

dose in females. Default is "double", which corresponds to X-inactivation.

BR.girls To be used with xchrom = TRUE. Gives the ratio of baseline risk for females

relative to the baseline risk for males.

response The default value "mult" specifies a multiplicative dose-response model. response

= "free" is not yet implemented.

alpha alpha is the Type I Error probablity. Equals 0.05 by default.

.. Could include argument reference. By default, the most frequent allele or

haplotype is chosen as reference. The reference haplotype can be set explicitly

by giving a numeric value for the reference argument.

#### **Details**

The Haplin framework includes different modules for assessing genetic effects: haplin, haplinStrat and haplinSlide. hapPowerAsymp computes the asymptotic power for these analyses, which enables power calculations of fetal effects, maternal effects and/or parent-of-origin effects. Various

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family designs, i.e., triads, case-control, the hybrid design, and all intermediate designs, are possible. It also allows power calculation of gene-environment interaction effects and effects on X-chromosome markers.

hapPower computes power through "brute force" simulations using hapRun. This is a robust way of checking software implementations, asymptotic approximations and attained significance level. However, both power and the corresponding sample size calculations can be performed much more efficiently using asymptotic approximations. The asymptotic power is calculated applying the noncentrality parameter of the Wald tests, which use the asymptotic normal distribution of the log-scale parameter. The function hapCovar (used by hapPowerAsymp) computes the variance-covariance matrix by applying the log-linear model combined with the EM algorithm.

#### Specifying haplotype risks:

The number of haplotypes used in the simulations is determined by the nall argument, since prod(nall) different haplotypes can be made from the specified number of markers, length(nall). The arguments haplo.freq, RR, RRcm, RRcf, RRstar, RR.mat, and RRstar.mat are all lists where each element represents a stratum. Within each stratum, the arguments are vectors of length equal to the number of haplotypes, specifying the relative risk etc. associated with each haplotype. The stratum specific arguments may be simplified if the number of strata is one, or if the arguments are equal across all strata. The haplotypes are determined by creating all possible haplotypes from the given markers, in a sequence where the first marker varies mostly quickly. For instance, if nall = c(3,2), the first marker has 3 alleles, the second has 2, and 6 haplotypes are possible. Taken in order, the haplotypes are 1-1, 1-2, 1-2, 1-2, and 1-2. When specifying, say, 1-2, 1-2, 1-2, 1-2, and 1-2. When specifying, say, 1-2, and second with 1-2 has a double risk compared to the rest. With, for instance, two strata, the specification 1-2 has a double risk compared to the rest. With, for instance, two strata, the specification 1-2 has a double risk compared to the rest. With, for instance, two strata, the specification 1-2 has a double risk stratum, not the second. The simplest example would be with nall 1-2 and 1-2 has a double simulate a single SNP where the second allele has a double risk.

# Specifying genetic effects:

Standard fetal effects are specified by the arguments RR and RRstar, whereas parent-of-origin effects are addressed by the arguments RRcm, RRcf and RRstar. Maternal effects are included by the additional arguments RRmat and RRstar.mat.

#### Value

hapPowerAsymp returns the asymptotic power for the relevant genetic effects. The first element of the list depicts the power for each haplotype analyzed separately. If there are more than two possible haplotypes, the second element displays the overall power for all haplotypes combined.

#### Author(s)

Miriam Gjerdevik, with Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health

<hakon.gjessing@uib.no>

#### References

```
Web Site: https://haplin.bitbucket.io
```

#### See Also

haplin, haplinSlide, haplinStrat, hapSim, hapRun, snpPower, hapPower

#### **Examples**

```
## Calculate the asymptotic power for a triad design
## when the minor allele increases the fetal risk by twofold.
## Assumes a multiplicative dose-response relationship.
hapPowerAsymp(nall = c(2), n.strata = 1, cases = list(c(mfc=120)),
haplo.freq = c(0.1,0.9), RR = c(2,1), RRstar = c(1,1))
## Calculate the asymptotic power for the hybrid design when
## the minor allele increases the fetal risk by twofold
## in the first stratum and no effect is seen in the second
## i.e., gene-environment interaction (GxE) effects.
hapPowerAsymp(nall = c(2), n.strata = 2, cases = list(c(mfc=100)),
controls = list(c(mfc=100)), haplo.freq = c(0.1,0.9),
RR = list(c(2,1), c(1,1)), RRstar = c(1,1))
## Calculate the asymptotic GxE power assessing maternal- and
## parent-of-origin effects at two diallelic loci.
hapPowerAsymp(nall = c(2,2), n.strata = 2, cases = c(mfc=500),
haplo.freq = c(0.1, 0.2, 0.3, 0.4),
RRcm = list(c(3,1,1,1), c(1,1,1,1)), RRcf = c(1,1,1,1), RRstar = c(1,1,1,1),
RR.mat = list(c(1.5,1,1,1),c(1,1,1,1)), RRstar.mat = c(1,1,1,1))
```

hapRelEff

Relative efficiency comparing different study designs in genetic association analysis with Haplin

# **Description**

Computes the relative efficiency for different study designs in genetic association analysis.

#### **Usage**

```
hapRelEff(nall = 2, cases.comp, controls.comp,
cases.ref, controls.ref, haplo.freq,
RR, RRcm, RRcf, RRstar, RR.mat, RRstar.mat,
xchrom = F, sim.comb.sex = "double", BR.girls,
response = "mult", ...)
```

#### **Arguments**

nall A vector of the number of alleles at each locus. By default a diallellic SNP.

cases.comp A list of the number of case families in the comparison design. Its element is

a vector of the number of families of the specified family design. The possible family designs, i.e., the possible names of the elements, are "mfc" (full triad), "mc" (mother-child dyad), "fc" (father-child dyad) or "c" (a single case child).

controls.comp A list of the number of control families in the comparison design. Its element is

a vector of the number of families of the specified family design. The possible family designs are "mfc" (full triad), "mc" (mother-child-dyad), "fc" (father-child dyad), "mf" (mother-father dyad), "c" (a single control child), "m" (a sin-

gle control mother) or "f" (a single control father).

cases.ref A list of the number of case families in the reference design. The options are the

same as for cases.comp.

controls.ref A list of the number of control families in the reference design. The options are

the same as for controls.comp.

haplo.freq A list of which its element is a numeric vector of the haplotype frequencies. The

frequencies are normalized and sum to one. The Details section shows how to

implement this argument in agreement with the possible haplotypes.

RR A list of which its element is a numeric vector of the relative risks. The Details

section shows how to implement this argument in agreement with the possible

haplotypes.

RRcm A numeric vector in list format containing the relative risks associated with the

haplotypes transmitted from the mother. See Details for description of how to

implement this argument in agreement with the possible haplotypes.

RRcf A numeric vector in list format containing the relative risks associated with the

haplotypes transmitted from the father. See Details for description of how to

implement this argument in agreement with the possible haplotypes.

RRstar A numeric vector in list format. Estimates how much double-dose children

would deviate from the risk expected in a multiplicative dose-response relation-

ship.

RR.mat The interpretation is similar to RR but for maternal genetic effects.

RRstar.mat The interpretation is similar to RRstar but for maternal genetic effects.

xchrom Logical. Equals FALSE by default, which indicates analyses of autosomal mark-

ers. If TRUE, analyses are perforned on X-linked markers.

sim.comb.sex To be used with xchrom = TRUE. A character value that specifies how to handle

gender differences on the X-chromosome. If "single", the effect of a (single) allele in males is equal to the effect of a single allele dose in females, and similarly, if "double", a single allele in males has the same effect as a double allele dose in females. Default is "double", which corresponds to X-inactivation.

BR.girls To be used with xchrom = TRUE. Gives the ratio of baseline risk for females

relative to the baseline risk for males.

response The default value "mult" specifies a multiplicative dose-response model. response

= "free" is not yet implemented.

... Could include argument reference. By default, the most frequent allele or haplotype is chosen as reference. The reference haplotype can be set explicitly by giving a numeric value for the reference argument.

#### **Details**

hapRelEff compares two study designs for genetic association analysis, using a term called relative efficiency. The relative efficiency is defined as the ratio of variances of estimators for the same parameter, computed from two different designs, or equivalently, the ratio of the sample sizes needed for each of the two designs to achieve the same level and power. The number of genotyped individuals within each design is accounted for. The relative efficiency estimated under the null hypothesis, i.e., when all relative risks are equal to one, is known as the Pitman efficiency. To compute the variance-covariance estimates for each design, hapRelEff calls the function hapCovar, which calculates the asymptotic variance-covariance matrix by applying the log-linear model combined with the EM algorithm.

The relative efficiency can be computed for fetal effects, maternal effects and parent-of-origin effect, as well as effects on X-chromosome markers. Various study designs, i.e., case-parent triads, the standard case-control design, the hybrid design, and all intermediate designs, can be compared.

Note that the exact number of case families and control families in the reference or comparison design is irrelevant, as this will be accounted for in the relative efficiency estimate. However, the ratio of control families to case families within the reference or comparison design must be specified correctly. See the Examples section.

#### Specifying haplotype risks:

The number of haplotypes used in the simulations is determined by the nall argument, since prod(nall) different haplotypes can be made from the specified number of markers, length(nall). The arguments haplo.freq, RR, RRcm, RRcf, RRstar, RR.mat, and RRstar.mat are all lists, containing vectors of length equal to the number of haplotypes, specifying the relative risk etc. associated with each haplotype. However, the function will work without using the list format. The haplotypes are determined by creating all possible haplotypes from the given markers, in a sequence where the first marker varies mostly quickly. For instance, if nall = c(3,2), the first marker has 3 alleles, the second has 2, and 6 haplotypes are possible. Taken in order, the haplotypes are 1-1, 2-1, 3-1, 1-2, 2-2, and 3-2. When specifying, say, RR = c(1,2,1,1,1,1) the haplotype 2-1 has a double risk compared to the rest. The simplest example would be with nall = c(2) and RR = c(1,2), which would simulate a single SNP where the second allele has a double risk.

# Specifying genetic effects:

Standard fetal effects are specified by the arguments RR and RRstar, whereas parent-of-origin effects are addressed by the arguments RRcm, RRcf and RRstar. Maternal effects are included by the additional arguments RRmat and RRstar.mat.

#### Value

hapRelEff returns the relative efficiency estimate, comparing two study designs. The first element of the list depicts the relative efficiency for each haplotype analyzed separately. If there are more than two possible haplotypes, the second element displays the overall relative efficiency for all haplotypes combined.

# Author(s)

Miriam Gjerdevik, with Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health

<hakon.gjessing@uib.no>

#### References

Web Site: https://haplin.bitbucket.io

#### See Also

haplin, hapPowerAsymp, hapRun, snpPower, hapPower

#### **Examples**

```
## Child effects: Calculate the efficiency of the standard case-control design
## (with an equal number of case and control children)
## relative to the case-parent triad design
## under the null hypothesis when the minor allele frequency is 0.1.
hapRelEff(nall = c(2), cases.comp = c(c=1),
controls.comp = c(c=1), cases.ref = c(mfc=1),
haplo.freq = c(0.1,0.9), RR = c(1,1))
## Child effects: Calculate the efficiency of the standard case-control design,
## with twice as many cases as controls, relative to the case-parent triad design
\#\# under the null hypothesis when the minor allele frequency is 0.2.
hapRelEff(nall = c(2), cases.comp = c(c=2),
controls.comp = c(c=1), cases.ref = c(mfc=1),
haplo.freq = c(0.2,0.8), RR = c(1,1))
## Child and maternal effects: Calculate the efficiency of the case-mother dyad design
## relative to the case-parent triad design
## under the null hypothesis when the minor allele frequency is 0.1.
hapRelEff(nall = c(2), cases.comp = c(mc=1), cases.ref = c(mfc=1),
haplo.freq = c(0.1,0.9), RR = c(1,1), RR.mat=c(1,1))
## PoO effects: Calculate the efficiency of the full hybrid design,
## with twice as many control families as case families,
## relative to the case-parent triad design
## under the null hypothesis when the minor allele frequency is 0.1.
hapRelEff(nall = c(2), cases.comp = c(mfc=1),
controls.comp = c(mfc=2), cases.ref = c(mfc=1),
haplo.freq = c(0.1,0.9), RRcm = c(1,1), RRcf = c(1,1))
```

hapRun

Simulates genetic data and runs Haplin for each simulation

## **Description**

Calculates Haplin results by first simulating genetic data, allowing a various number of family designs, and then running Haplin on the simulations. The simulated data may contain of fetal effects, maternal effects and/or parent-of-origin effects. The function allows for simulations and calculations on both autosomal and X-chromosome markers, assuming Hardy-Weinberg equilibrium. It enables simulation and calculation of gene-environment interaction effects, i.e, the input (relative risks, number of cases etc.) may vary across strata. hapRun calls haplin, haplinStrat or haplinSlide to run on the simulated data files.

#### Usage

```
hapRun(nall, n.strata= 1, cases, controls, haplo.freq,
RR, RRcm, RRcf, RRstar, RR.mat, RRstar.mat, hapfunc = "haplin",
gen.missing.cases = NULL, gen.missing.controls = NULL,
n.sim = 1000, xchrom = FALSE, sim.comb.sex = "double", BR.girls, dire,
ask = TRUE, cpus = 1, slaveOutfile = "", ...)
```

## **Arguments**

cases

nall A vector of the number of alleles at each locus.

n. strata The number of strata.

A list of the number of case families. Each element is a vector of the number of families of the specified family design(s) in the corresponding stratum. The possible family designs, i.e., the possible names of the elements, are "mfc" (full triad), "mc" (mother-child dyad), "fc" (father-child dyad) or "c" (a single case

child). See Details for a thorough description.

A list of the number of control families. Each element is a vector of the number of families of the specified family design(s) in the corresponding stratum. The possible family designs are "mfc" (full triad), "mc" (mother-child-dyad), "fc" (father-child dyad), "mf" (mother-father dyad), "c" (a single control child), "m" (a single control mother) or "f" (a single control father). See Details for a thorough description.

A list of which each element is a numeric vector of the haplotype frequencies in each stratum. The frequencies are normalized and sum to one. The Details section shows how to implement this argument in agreement with the possible haplotypes.

A list of which each element is a numeric vector of the relative risks in each stratum. The Details section shows how to implement this argument in agreement with the possible haplotypes.

controls

haplo.freq

RR

RRcm A list of numeric vectors. Each vector contains the relative risks associated with the haplotypes transmitted from the mother for this stratum. See Details for description of how to implement this argument in agreement with the possible haplotypes.

RRcf A list of numeric vectors. Each vector contains the relative risks associated

with the haplotypes transmitted from the father for this stratum. See Details for description of how to implement this argument in agreement with the possible haplotypes.

A list of numeric vectors. Estimates how much double-dose children would deviate from the risk expected in a multiplicative dose-response relationship. The interpretation is similar to RR when simulating genetic data with maternal

RRstar.mat The interpretation is similar to RRstar when simulating genetic data with maternal effects.

hapfunc Defines which haplin function to run, the options being "haplin", "haplinSlide" or "haplinStrat". "haplinSlide" is however only partially implemented.

gen.missing.cases

Generates missing values at random for the case families. Set to NULL by default, i.e., no missing values generated. See Details for description of how to

fault, i.e., no missing values generated. See Details for description of how to implement this argument.
gen.missing.controls

Generates missing values at random for the control families. Set to NULL by default, i.e., no missing values generated. See Details for description of how to implement this argument.

The number of simulations, i.e., the number of simulated data files.

xchrom Logical. Equals FALSE by default, which indicates simulation of autosomal markers. If TRUE, hapSim simulates X-linked genes.

To be used with xchrom = TRUE. A character value that specifies how to handle gender differences on the X-chromosome. If "single" the effect of a (single)

gender differences on the X-chromosome. If "single", the effect of a (single) allele in males is equal to the effect of a single allele dose in females, and similarly, if "double", a single allele in males has the same effect as a double allele dose in females. Default is "double", which corresponds to X-inactivation.

BR.girls To be used with xchrom = TRUE. Gives the ratio of baseline risk for females to the baseline risk for males.

Gives the directory of the simulated data files. Missing by default, which means that none of the files are saved to files.

Logical. If TRUE, hapSim will ask before overwriting the files in an already existing directory.

Allows parallel processing of its analyses. The cpus argument should preferably be set to the number of available CPUs. If set lower, it will save some capacity for other processes to run. Setting it too high should not cause any serious problems.

Character. If slaveOutfile = "" (default), output from all running cores will be printed in the standard R session window. Alternatively, the output can be saved to a file by specifying the file path and name.

Arguments to be used by haplin, haplinSlide or haplinStrat.

Gene

effects.

sim.comb.sex

n.sim

RRstar

RR.mat

dire

ask

cpus

slaveOutfile

. . .

#### **Details**

hapRun applies haplin, haplinSlide or haplinStrat on each data file simulated by hapSim. It provides simulations on various family designs, i.e., triads, case-control, the hybrid design, and all intermediate designs. The simulated files may accommodate fetal effects, maternal effects and/or parent-of-origin effects. hapRun allows simulation of both autosomal and X-chromosome markers, assuming Hardy-Weinberg equilibrium. It also enables simulation and calculation of geneenvironment interaction effects.

Details on how to implement the arguments listed above are provided by hapSim and the Examples section below. The stratum specific arguments may be simplified if the number of strata is one, or if the arguments are equal across all strata.

haplin, haplinStrat and haplinSlide will run with default values unless otherwise specified by hapRun. For example, if hapfunc = "haplin", haplin will use response = "free" unless response = "mult" is explicitly given as an argument in hapRun. Moreover, triads with missing data are only included in the haplin analysis if the argument use.missing equals TRUE (default in hapRun). Please confer https://haplin.bitbucket.io/docu/Haplin\_power.pdf for further details and examples.

For information on the arguments to be passed on to haplin, haplinStrat and haplinSlide, please consult their help pages.

Note that RR.mat and RRstar.mat and RRcm and RRcf are required for hapSim to simulate maternal and parent-of-origin effects, respectively. To calculate these effects, however, arguments maternal = TRUE and/or poo = TRUE must be specified.

gen.missing.cases and gen.missing.controls are flexible arguments. By default, both equal NULL, which means that no missing data are generated at random. If the arguments are single numbers, missing data are generated at random with this proportion for all cases and/or controls. If the arguments are vectors of length equal to the number of loci, missing data are generated with the corresponding proportion for each locus. The arguments can also be matrices with the number of rows equal to the number of loci and three columns. Each row corresponds to a locus, and the columns correspond to mothers, fathers and children, respectively.

#### Value

If hapfunc = "haplin", hapRun returns a dataframe consisting of results from running haplin on each simulated file. The first two columns are:

sim.no The name of the directory from which the results are calculated, i.e., the simu-

lation number

row.no The row number within each simulation

haptable gives detailed information of the full dataframe.

If hapfunc = "haplinSlide", hapRun returns a list of which each element contains the results from a single run of haplinSlide. Consult suest for a thorough description of the output. Note, however, that hapfunc = "haplinSlide" is currently only implemented for diallelic markers, and the reference category is always chosen to be the first haplotype (see hapSim for a description of the haplotype grid).

If hapfunc = "haplinStrat", haplinStrat is used to estimate gene-effects in each stratum of the exposure covariate, and the results from all strata are compared using gxe. hapRun returns a list, where each element is the result of a single run of gxe.

Additionaly, if dire is not missing by default, the simulated files from which the Haplin results are calculated, are stored in the given directory.

#### Author(s)

```
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Professor of Biostatistics
Division of Epidemiology
Norwegian Institute of Public Health
<a href="https://doi.org/10.1007/j.com/">https://doi.org/10.1007/j.com/</a>
```

#### References

```
Web Site: https://haplin.bitbucket.io
```

#### See Also

haplin, haplinSlide, hapSim, haptable, suest, hapPower, hapPowerAsymp

# **Examples**

```
## Not run:
## Simulate Haplin results from 100 files using the multiplicative model in haplin.
## The files consist of fetal effects at two diallelic markers,
## corresponding to haplo.freq = rep(0.25, 4), RR = c(2,1,1,1)
## and RRstar = c(1,1,1,1). That is, the first allele has a doubled risk
## relative to the rest. The data consists of a combination of
## 100 case triads and 100 control triads with no missing data.
## No environmental factors are considered, i.e. the number of strata is one.
hapRun(nall = c(2,2), n.strata = 1, cases = c(mfc=100), controls = c(mfc=100),
haplo.freq = rep(0.25,4), RR = c(2,1,1,1), RRstar = c(1,1,1,1),
hapfunc = "haplin", response = "mult", n.sim = 100, dire = "simfiles", ask = FALSE)
## Simulate power from 100 files applying haplinStrat.
## The files consist of fetal and maternal effects at two diallelic markers.
## The data is simulated for 500 case triads and 200 control families in the first stratum,
## and 500 case triads and 500 control trids in the second.
## The fetal effects vary across strata,
## whereas the maternal effects are the same.
## One percent of the case triads are missing at random in the second stratum.
hapRun(nall = c(2,2), n.strata = 2, cases = c(mfc=500),
controls = list(c(mfc=200), c(mfc=500)), haplo.freq = rep(0.25, 4), maternal = TRUE,
RR = list(c(1.5,1,1,1),c(1,1,1,1)), RRstar = c(1,1,1,1),
RR.mat = c(1.5,1,1,1), RRstar.mat = c(1,1,1,1),
gen.missing.cases = list(NULL,0.01), use.missing = TRUE, hapfunc = "haplinStrat",
n.sim = 100, ask = FALSE)
## Simulate Haplin results from 100 files using haplin.
## The files consist of fetal effects at one diallelic locus,
```

```
## corresponding to haplo.freq = rep(0.5,2), RR = c(1.5,1) and RRstar = c(1,1).
## We have a combination of 100 case triads and
## 100 control triads with no missing data.
## No environmental effects are considered.
hapRun(nall = c(2), n.strata = 1, cases = c(mfc=100), controls = c(mfc=100),
haplo.freq = rep(0.5,2), RR = c(1.5,1), RRstar = c(1,1),
hapfunc = "haplin", n.sim = 100, dire = "simfiles", ask = FALSE)
## End(Not run)
```

hapSim

Simulation of genetic data in Haplin format

# **Description**

Simulates genetic data in Haplin format, consisting of fetal effects, maternal effects and/or parentof-origin effects. Allows for simulation of both autosomal and X-linked markers, assuming Hardy-Weinberg equilibrium. Enables stratified simulations for gene-environment interaction analyses, i.e the input (relative risks, number of cases etc) may vary across different strata.

## Usage

```
hapSim(nall, n.strata = 1, cases, controls, haplo.freq,
RR, RRcm, RRcf, RRstar, RR.mat, RRstar.mat,
gen.missing.cases = NULL, gen.missing.controls = NULL,
n.sim = 1000, xchrom = F, sim.comb.sex = "double", BR.girls, dire = "simfiles",
ask = TRUE, verbose = TRUE, cpus = 1)
```

#### **Arguments**

nall A vector of the number of alleles at each locus.

The number of strata. n.strata

> A list of the number of case families. Each element is a vector of the number of families of the specified family design(s) in the corresponding stratum. The possible family designs, i.e., the possible names of the elements, are "mfc" (full triad), "mc" (mother-child dyad), "fc" (father-child dyad) or "c" (a single case

child). See Details for a thorough description.

A list of the number of control families. Each element is a vector of the number of families of the specified family design(s) in the corresponding stratum. The possible family designs are "mfc" (full triad), "mc" (mother-child-dyad), "fc" (father-child dyad), "mf" (mother-father dyad), "c" (a single control child), "m" (a single control mother) or "f" (a single control father). See Details for a

thorough description.

haplo.freq A list of which each element is a numeric vector of the haplotype frequencies in

each stratum. The frequencies will be normalized so that they sum to one. The Details section shows how to implement this argument in agreement with the

possible haplotypes.

cases

controls

RR A list of which each element is a numeric vector of the relative risks in each stra-

tum. The Details section shows how to implement this argument in agreement

with the possible haplotypes.

**RRcm** A list of numeric vectors. Each vector contains the relative risks associated with

> the haplotypes transmitted from the mother for this stratum. See Details for description of how to implement this argument in agreement with the possible

haplotypes.

RRcf A list of numeric vectors. Each vector contains the relative risks associated

> with the haplotypes transmitted from the father for this stratum. See Details for description of how to implement this argument in agreement with the possible

haplotypes.

A list of numeric vectors. Estimates how much double-dose children would RRstar

deviate from the risk expected in a multiplicative dose-response relationship.

RR.mat The interpretation is similar to RR when simulating genetic data with maternal

effects.

The interpretation is parallel to RRstar when simulating genetic data with ma-RRstar.mat

ternal effects.

gen.missing.cases

Generates missing values at random for the case families. Set to NULL by default, i.e., no missing values generated. See Details for description of how to

implement this argument.

gen.missing.controls

Generates missing values at random for the control families. Set to NULL by default, i.e., no missing values generated. See Details for description of how to

implement this argument.

n.sim The number of simulations, i.e., the number of simulated data files.

xchrom Logical. Equals FALSE by default, which indicates simulation of autosomal

markers. If TRUE, hapSim simulates X-linked genes.

To be used with xchrom = TRUE. A character value which specifies how to handle sim.comb.sex

> gender differences on the X-chromosome. If "single", the effect of a (single) allele in males is equal to the effect of a single allele dose in females, and similarly, if "double", a single allele in males has the same effect as a double allele

dose in females. Default is "double", which corresponds to X-inactivation.

To be used with xchrom = TRUE. Gives the ratio of baseline risk for females to BR.girls

the baseline risk for males.

dire Gives the directory of the simulated data files.

Logical. If TRUE, hapSim will ask before overwriting the files in an already ask

existing directory.

Logical. Default is TRUE, which means that the file name is displayed for each verbose

iteration. Works only when cpus = 1.

cpus Allows simulations to be performed in parallel. The cpus argument should

preferably be set to the number of available cores. If set lower, it will save some capacity for other processes to run. Setting it too high should not cause

any serious problems.

#### **Details**

hapSim simulates allele values for case and control families at multiple markers (typically in LD) simultaneously. The number of markers/SNPs involved will typically be in the range 1 to 6. Data are simulated to produce relative risks of disease as specified by the user input. Simulations can be performed separately over a number of strata so as to simulate gene-environment interactions.

## Specifying haplotype risks:

The number of haplotypes used in the simulations is determined by the nall argument, since prod(nall) different haplotypes can be made from the specified number of markers, length(nall). The arguments haplo.freq, RR, RRcm, RRcf, RRstar, RR.mat, and RRstar.mat are all lists where each element represents a stratum. Within each stratum, the arguments are vectors of length equal to the number of haplotypes, specifying the relative risk etc. associated with each haplotype. The stratum specific arguments may be simplified if the number of strata is one, or if the arguments are equal across all strata.

The haplotypes are determined by creating all possible haplotypes from the given markers, in a sequence where the first marker varies mostly quickly. For instance, if nall = c(3,2), the first marker has 3 alleles, the second has 2, and 6 haplotypes are possible. Taken in order, the haplotypes are 1-1, 2-1, 3-1, 1-2, 2-2, and 3-2. When specifying, say, RR = c(1,2,1,1,1,1) the haplotype 2-1 has a double risk compared to the rest. With, for instance, two strata, the specification RR = list(c(1,2,1,1,1,1), c(1,1,1,1,1,1)) would mean that the risk associated with 2-1 is elevated only in the first stratum, not the second.

The simplest example would be with nall = c(2) and RR = c(1,2), which would simulate a single SNP where the second allele has a double risk.

#### Output file format:

The format of the simulated files is relatively flexible and allows multi-allelic markers and various designs. If both case and control families are present, the simulated files contain a leading column of the case/control status (1/0). If xchrom=TRUE, the neighboring column to the left of the genetic data contains the sex information (1 = male, 2 = female). Each line represents genotypes for a case or control triad.

There are six columns for each locus, two for the mother (M), two for the father (F) and two for the child (C). The columns are placed in the following sequence: M11 M12 F11 F12 C11 C12 M21 M22 F21 F22 C21 C22... etc, where the first number indicates marker, and the second number indicates the first or second allele at this locus. Columns are separated by a single white space, and missing data are coded as NA.

Intermediate designs, for instance mother-child dyads, are represented as full triads with columns of absent family members set to missing. In the case of a pure case-control design, however, each line represents a single individual, and there are no columns representing mothers and fathers.

There are no row or column names in the files.

Some examples are given below. See <a href="https://haplin.bitbucket.io/docu/haplin\_data\_format">httml</a> for a thorough description of the Haplin format. Note that this description separates the two alleles for an individual within a locus by a semi-colon, such as 1;2. This is, however, not necessary. Confer the document <a href="https://haplin.bitbucket.io/docu/Haplin\_power.pdf">https://haplin.bitbucket.io/docu/Haplin\_power.pdf</a> for details and examples on how to perform the simulations.

gen.missing.cases and gen.missing.controls are flexible arguments. By default, both equal NULL, which means that no missing data are generated at random. If the arguments are single

numbers, missing data are generated at random with this proportion for all cases and/or controls. If the arguments are vectors of length equal to the number of loci, missing data are generated with the corresponding proportion for each locus. The arguments can also be matrices with the number of rows equal to the number of loci and three columns. Each row corresponds to a locus, and the columns correspond to mothers, fathers and children, respectively.

## Author(s)

Miriam Gjerdevik, with Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health

<hakon.gjessing@uib.no>

#### References

Web Site: https://haplin.bitbucket.io

#### See Also

haplin, hapRun, hapPower

## **Examples**

```
## Simulate genetic data (100 files) at two diallelic markers, consisting of fetal effects
## corresponding to haplo.freq = rep(0.25, 4), RR = c(2,1,1,1) and RRstar = c(1,1,1,1),
## for the combination of 1000 case triads and 1000 control triads with no missing data.
## Only one stratum.
hapSim(nall = c(2,2), n.strata = 1, cases = c(mfc=1000),
controls = c(mfc=1000), haplo.freq = rep(0.25,4),
RR = c(2,1,1,1), RRstar = c(1,1,1,1), n.sim = 100, dire = "simfiles")
## Simulate genetic data (100 files) at two diallelic markers,
## consisting of fetal and maternal effects corresponding to
## haplo.freq = rep(0.25, 4), RR = c(2,1,1,1), RRstar = c(1,1,1,1),
## RR.mat = c(2,1,1,1) and RRstar.mat = c(1,1,1,1),
## for 1000 case triads and zero control families.
## One percent of the case triads are missing at random. One stratum only.
hapSim(nall = c(2,2), n.strata=1, cases = c(mfc=1000),
controls = c(mfc=0), haplo.freq = rep(0.25,4), RR = c(2,1,1,1),
RRstar = c(1,1,1,1), RR.mat = c(2,1,1,1), RRstar.mat = c(1,1,1,1),
gen.missing.cases = 0.01, n.sim = 100, dire = "simfiles")
## Simulate genetic data (100 files) at two diallelic markers. In the first stratum,
## we have a combination of 500 case triads and 500 control triads with
## haplo.freq = rep(0.25, 4), RR = c(2,1,1,1) and RRstar = c(1,1,1,1).
## In the second stratum, we have 300 case triads and 500 control triads with
## haplo.freq = rep(0.25, 4), RR = c(1,1,1,1) and RRstar = c(1,1,1,1).
```

haptable 45

```
## One percent of the control triads are missing at random in the first stratum.  \begin{array}{l} \text{hapSim}(\text{nall} = \text{c(2,2)}, \text{ n.strata= 2, cases} = \text{list}(\text{c(mfc=500)}, \text{c(mfc=300)}), \\ \text{controls} = \text{c(mfc=500)}, \text{haplo.freq} = \text{rep(0.25,4)}, \\ \text{RR} = \text{list}(\text{c(2,1,1,1)}, \text{c(1,1,1,1)}), \\ \text{RRstar} = \text{c(1,1,1,1)}, \\ \text{gen.missing.controls} = \text{list(0.01}, \text{NULL)}, \\ \text{n.sim} = \text{100}, \\ \text{dire} = \text{"simfiles"}) \\ \text{## End(Not run)}
```

haptable

Create haplin table

# **Description**

Create a comprehensive table of haplin output

## Usage

```
haptable(object)
```

#### **Arguments**

object

A haplin object, i.e. the result of running haplin.

#### **Details**

haptable extracts the most important information from a haplin object to produce a summary table. The table can then be saved with, for instance, write.table, making the results easily accessible to other applications. You can also use output to produce and save the same results.

#### Value

— A dataframe is returned, with the following columns: —-

Name(s) of marker(s) investigated marker alleles A listing of the alleles found at each marker counts Frequency counts of alleles at each marker HWE.pv P-value from Hardy-Weinberg equilibrium test at each marker **Original** Number of triads before removal Number of triads after removal of missing After.rem.NA After.rem.Mend.inc. Number of triads after removal of Mendelian inconsistencies After.rem.unused.haplos Number of triads after removal of unused (rare) haplotypes pv.overall Overall likelihood ratio p-value (test of all genetic effects combined) haplos Haplotypes (or single-marker alleles) found during estimation haplofreq Estimated haplotype frequencies

46 haptable

haplofreq.lower

Lower 95% CI for estimated haplotype frequencies

haplofreq.upper

Upper 95% CI for estimated haplotype frequencies

reference Reference method. If ref.cat is used, the reference category is labeled "ref"

RR.est. Estimated single dose relative risk

RR. lower

Lower 95% CI for single dose relative risk

Upper 95% CI for single dose relative risk

RR. p. value

P-values for individual single dose effect

RRdd.est. Estimated double dose relative risk

RRdd.lower Lower 95% CI for double dose relative risk
Upper 95% CI for double dose relative risk
RRdd.p.value P-values for individual double dose effect

NOTE1 When maternal = TRUE, there will be additional columns:

RRm.est. Estimated single dose relative risk for maternal haplotype

RRm.lower Lower 95% CI for single dose relative risk for maternal haplotype
RRm.upper Upper 95% CI for single dose relative risk for maternal haplotype
RRm.p.value P-values for individual single dose effect of maternal haplotype
RRmdd.est. Estimated double dose relative risk for maternal haplotype

RRmdd.lower Lower 95% CI for double dose relative risk for maternal haplotype

RRmdd.upper Upper 95% CI for double dose relative risk for maternal haplotype

RRmdd.p.value P-values for individual double dose effect of maternal haplotype

NOTE2 When poo = TRUE, the RR.est., RR.lower, and RR.upper columns will be re-

placed by the following columns:

RRcm.est. Estimated single dose relative risk, when inherited from the mother

RRcm.lower Lower 95% CI for single dose relative risk, when inherited from the mother Upper 95% CI for single dose relative risk, when inherited from the mother RRcm.p.value P-values for individual single dose effects, when inherited from the mother

RRcf.est. Estimated single dose relative risk, when inherited from the father

RRcf.lower

Lower 95% CI for single dose relative risk, when inherited from the father

Upper 95% CI for single dose relative risk, when inherited from the father

RRcf.p.value

P-values for individual single dose effects, when inherited from the father

RRcm\_RRcf.est. An estimate of parent-of-origin effect, i.e. the ratio RRcm/RRcf

RRcm\_RRcf.lower

Lower 95% CI for ratio RRcm/RRcf

RRcm\_RRcf.upper

Upper 95% CI for ratio RRcm/RRcf

RRcm\_RRcf.p.value

P-value for parent-of-origin effect RRcm/RRcf at that marker

initParallelRun 47

# Note

Further information is found on the web page

# Author(s)

Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health <hakon.gjessing@uib.no>

#### References

```
Web Site: https://haplin.bitbucket.io
```

# See Also

```
haplin, output
```

## **Examples**

```
## Not run:

# Produce a table containing the most important output from haplin:
res <- haplin("data.dat", use.missing = T, maternal = T)
haptable(res)

## End(Not run)</pre>
```

initParallelRun

Initialization of the Rmpi cluster

# Description

This function prepares a cluster using Rmpi package. The initialization is paired with closing of the cluster using the finishParallelRun function.

## Usage

```
initParallelRun(cpus)
```

# **Arguments**

cpus

Number of cores to use (optional). If given, only that number of CPUs will be used. By default (if not set), the Rmpi will check how many CPUs are available in the system and take the maximum number.

48 lineByLine

# Description

Modifies a data file line by line, i.e. reads a file line by line, converts each line, then writes to the modified file. This method is especially useful when modifying large datasets, where the reading of entire files may be time consuming and require a large amount of memory.

# Usage

```
lineByLine(infile, outfile, linefunc = identity, choose.lines = NULL,
choose.columns = NULL, col.sep = " ", ask = TRUE,
blank.lines.skip = TRUE, verbose = TRUE, ...)
```

# Arg

Ę	rguments			
	infile	A character string giving the name and path of the file to be modified.		
	outfile	A character string giving the name of the modified file. The name of the file is relative to the current working directory, unless the file name contains a definite path.		
	linefunc	lineByLine modifies each line using linefunc. Default is the identity function. The user may define his or her own line-modifying functions, see Details for a thorough description.		
	choose.lines	A numeric vector of lines to be selected or dropped from infile. Positive values refer to lines to be chosen, whereas negative values refer to lines to be skipped. The vector cannot include both positive and negative values at the same time. If "NULL" (default), all lines are selected.		
	choose.columns	A numeric vector of columns to be selected (positive values) or skipped (negative values) from infile. The vector cannot include both positive and negative values at the same time. By default, all columns are selected without reordering among the columns. Duplication and reordering among the selected columns will occur in the modified file corresponding to the order in which the columns are listed.		
	col.sep	Specifies the separator that splits the columns in infile. By default, col.sep = "" (space). To split at all types of spaces or blank characters, set col.sep = "[[:space:]]" or col.sep = "[[:blank:]]".		
	ask	Logical. Default is "TRUE". If set to "FALSE", an already existing outfile will be overwritten without asking.		
	blank lines ski	n		

blank.lines.skip

Logical. If "TRUE" (default), lineByLine ignores blank lines in the input.

Logical. Default is "TRUE", which means that the line number is displayed verbose for each iteration, in addition to output from linefunc. If choose.columns

contains invalid column numbers, this will also be displayed.

Further arguments to be passed to linefunc.

lineByLine 49

#### **Details**

When reading large datafiles, functions such as read.table can use a large amount of memory and be extremely time consuming. Instead of reading the entire file at once, lineByLine reads one line at a time, modifies the line using linefunc, and then writes the line to outfile.

The user may specify his or her own line-converting function. This function must take the argument x, a character vector representing a single line of the file, split at spaces. However, additional arguments may be included. If verbose equals "TRUE", output should be displayed. The modified vector is returned.

The framework of the line-modifying function may look something like this:

```
lineModify <- function(x){
    xnew <- x

## Define any modifications, for instance recoding missing values in a dataset from NA to 0:
    xnew[is.na(.xnew)] <- 0

## Just to monitor progress, display, for instance, 10 first elements, without newline:
    cat(paste(.xnew[1:min(10, length(.xnew))], collapse = " "))

## Return converted vector
    return(.xnew)
}</pre>
```

See Haplin:::lineConvert for an additional example of a line-modifying function.

## Value

lineByLine returns the number of lines read, although invisible. The main objective is the modified file.

# Author(s)

```
Miriam Gjerdevik,
with Hakon K. Gjessing
Professor of Biostatistics
Division of Epidemiology
Norwegian Institute of Public Health
```

```
<hakon.gjessing@uib.no>
```

#### References

```
Web Site: https://haplin.bitbucket.io
```

#### See Also

convertPed

50 nindiv

## **Examples**

```
## Not run:
## Extract the first ten columns from "myfile.txt",
## without reordering
lineByLine(infile = "myfile.txt", outfile = "myfile_modified.txt",
choose.columns = c(1:10))
## End(Not run)
```

nfam

Count the number of families in the data

## **Description**

This is a help function to count the number of families in an object read in with genDataRead (or loaded with genDataLoad). Note: it is assumed that the study design is either 'triad' or 'cc.triad'.

## Usage

```
nfam(data.in)
```

# **Arguments**

data.in

The data read in by genDataRead.

## Value

How many families (integer).

nindiv

Count the number of individuals in the data

# **Description**

This is a help function to count the number of individuals in an object read in with genDataRead (or loaded with genDataLoad).

# Usage

```
nindiv(data.in, design = "triad")
```

nsnps 51

# **Arguments**

data.in The data read in by genDataRead.

design The design used in the study - choose from:

- triad (default) data includes genotypes of mother, father and child;
- *cc* classical case-control;
- cc.triad hybrid design: triads with cases and controls

# Value

How many individuals (integer).

nsnps

Count the number of markers in the data

# Description

This is a help function to count the number of markers in an object read in with genDataRead (or loaded with genDataLoad).

# Usage

```
nsnps(data.in, design = "triad")
```

## **Arguments**

data.in The data read in by genDataRead.

design The design used in the study - choose from:

- triad (default) data includes genotypes of mother, father and child;
- cc classical case-control;
- cc.triad hybrid design: triads with cases and controls

## Value

How many markers (integer).

52 output

output

Save files with summary, table, and plot from a haplin object.

# Description

Create summary tables and figure from a haplin object. Save results as separate files in a specified directory.

## Usage

```
output(object, dirname, ask = T)
```

# **Arguments**

object A haplin object, i.e. the result of running haplin.

dirname Text string, for instance "c:/work/haplinresults". Name of directory where re-

sults should be saved. Default is to save results in the current working directory.

ask Logical. If TRUE (default), you will be asked before overwriting any files with

the same name. If FALSE, output will overwrite without warning.

#### **Details**

After having run haplin and saved the result (in the R workspace), the output function will extract summary results, a summary table, and a plot of the results and save them to the specified directory. The filenames will be haplin\_summary.txt, haplin\_table.txt and haplin\_plot.jpg, respectively. output simply uses the available functions summary, haptable, and plot to produce the files, but is a quick way of saving all the relevant results.

#### Note

Further information is found on the web page.

## Author(s)

Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health <a href="https://doi.org/10.2016/j.nc">https://doi.org/10.2016/j.j.nc</a>

## References

Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396.

Web Site: https://haplin.bitbucket.io

plot.haplin 53

## See Also

```
haplin
```

## **Examples**

```
## Not run:

# Run haplin and save results in separate files
# in the c:\work\haplinresults directory:
res <- haplin("data.dat", use.missing = T, maternal = T)
output(res, dirname = "c:/work/haplinresults")

## End(Not run)</pre>
```

plot.haplin

Plot a haplin object

#### **Description**

Plot a haplin object and (optionally) produce picture files

#### Usage

```
## S3 method for class 'haplin'
plot(x, reference, separate.plots = F, filename,
filetype = "png", use.dd, ...)
```

## **Arguments**

x A haplin object, i.e. the result of running haplin. This is the only required

argument.

reference Same as reference argument in haplin. Note that when plotting, you can

only choose "reciprocal", "population" or "ref.cat". You cannot use a numeric value to change the reference category, to do that haplin must be run

over again. (See the reference argument of haplin.)

separate.plots Logical. If you estimate effects of both fetal and maternal genes you can de-

cide whether or not to plot them in the same plot. The default is the same plot (TRUE), the alternative (FALSE) means in separate plots. If you choose separate plots you may have to set the graphics window to "recording" to make sure

you can scroll back to the first plot.

filename If you want a file containing the plot to be produced, give a character string for

the filename.

filetype The default filetype is "png", alternatively you can choose "jpeg".

use.dd Numeric vector indicating which double dose estimates should be plotted. For

instance, if set to c(1,3) only the first and third haplotypes will be drawn with double dose estimates. This is useful if some haplotypes are rare and you want

to exclude the uncertain estimates from the plot.

Further arguments to be passed on to the plot function

54 plot.haplinSlide

# Note

Further information is found on the web page.

## Author(s)

```
Hakon K. Gjessing
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```

## References

Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396.

```
Web Site: https://haplin.bitbucket.io
```

## See Also

haplin

## **Examples**

```
## Not run:

# Produce separate plots for child and mother, dump plots to files:
res <- haplin("data.dat", use.missing = T, maternal = T)
plot(res, separate.plots = T, filename = "Haplinres.png")

## End(Not run)</pre>
```

plot.haplinSlide

Plotter function for haplinSlide.

# **Description**

This will plot any haplinSlide object in one figure.

# Usage

```
## S3 method for class 'haplinSlide'
plot(
    x,
    filename,
    title,
    windows,
```

plot.haplinSlide 55

```
plot.signif.only = FALSE,
  signif.thresh = 0.05,
  y.lim,
  x.title,
  y.title,
  file.width,
  file.height,
  ...
)
```

# Arguments

X	The haplinSlide object (NB: only the output produced by running haplinSlide with the table.output argument set to TRUE!)	
filename	If the plot should be saved to the disk, give the name of the output file including the file extension.	
title	If the user wishes to override the default title of the plot, give it here.	
windows	Numerical vector. If given, this will only plot the chosen windows.	
plot.signif.on	ly	
	Logical: whether to filter out the "non-significant" markers from the plot. Default: FALSE, i.e., plot everything.	
signif.thresh	The threshold defining the significant p-values: if plot.signif.only == TRUE, then only the markers with relative risk p-values lower than the threshold will be kept for plotting. Default: 0.05.	
y.lim	Vector with two numbers setting the Y limits of the plotted graph.	
x.title	Title for the X axis (default: "marker").	
y.title	Title for the Y axis (default: "RR (log scale)").	
file.width	Width (in inches) for the output plot, if a filename was given.	
file.height	Height (in inches) for the output plot, if a filename was given.	
	other arguments (ignored).	

# **Details**

The haplinSlide object is a list of haplin results - by default in haptable form. This is used to plot the relative risk estimates for all the markers in one plot, to enable easy comparison. NB: those estimates that have infinite confidence interval will not be plotted.

# Value

```
ggplot object.
```

56 plot.haplinStrat

plot.haplinStrat

Plotter function for haplinStrat results.

# Description

This will automatically plot all haplinStrat results on one figure.

# Usage

```
## S3 method for class 'haplinStrat'
plot(
    x,
    filename,
    title,
    windows,
    plot.signif.only = FALSE,
    signif.thresh = 0.05,
    y.lim,
    x.title,
    y.title,
    file.width,
    file.height,
    ...
)
```

# Arguments

	X	The haplinSlide object (NB: only the output produced by running haplinSlid with the table.output argument set to TRUE!)	
filename If the plot should be saved to the disk, give the name of the output file in the file extension.		If the plot should be saved to the disk, give the name of the output file including the file extension.	
	title	If the user wishes to override the default title of the plot, give it here.	
	windows	Numerical vector. If given, this will only plot the chosen windows.	
	plot.signif.onl	у	
		Logical: whether to filter out the "non-significant" markers from the plot. Default: FALSE, i.e., plot everything.	
	signif.thresh	The threshold defining the significant p-values: if plot.signif.only == TRUE, then only the markers with relative risk p-values lower than the threshold will be kept for plotting. Default: 0.05.	
	y.lim	Vector with two numbers setting the Y limits of the plotted graph.	
	x.title	Title for the X axis (default: "marker").	
	y.title	Title for the Y axis (default: "RR (log scale)").	
	file.width	Width (in inches) for the output plot, if a filename was given.	
	file.height	Height (in inches) for the output plot, if a filename was given.	
		other arguments (ignored).	

plot.haptable 57

#### **Details**

This function uses the same style as plot.haplinSlide and plots all of the haplinStrat results on one figure, for easy comparison. NB: those estimates that have infinite confidence interval will not be plotted.

#### Value

```
ggplot object.
```

plot.haptable

Plot a haptable object

## Description

Plot a haptable object, which is the result of running haptable on a haplin result, and (optionally) produce picture files.

# Usage

```
## S3 method for class 'haptable'
plot(x, separate.plots = F, filename,
filetype = "png", use.dd, verbose = T, ...)
```

#### **Arguments**

x A haptable object, i.e. the result of running haptable on a result from haplin.

This is the only required argument.

separate.plots Logical. If you estimate effects of both fetal and maternal genes you can de-

cide whether or not to plot them in the same plot. The default is the same plot (TRUE), the alternative (FALSE) means in separate plots. If you choose separate plots you may have to set the graphics window to "recording" to make sure

you can scroll back to the first plot.

filename If you want a file containing the plot to be produced, give a character string for

the filename.

filetype The default filetype is "png", alternatively you can choose "jpeg".

use.dd Numeric vector indicating which double dose estimates should be plotted. For

instance, if set to c(1,3) only the first and third haplotypes will be drawn with double dose estimates. This is useful if some haplotypes are rare and you want

to exclude the uncertain estimates from the plot.

verbose Turns on or off some minor comments when plotting
... Further arguments to be passed on to the plot function

## Note

Further information is found on the web page.

58 plotPValues

## Author(s)

Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health <hakon.gjessing@uib.no>

#### References

Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396.

```
Web Site: https://haplin.bitbucket.io
```

# See Also

haplin

## **Examples**

```
## Not run:

# Directly plotting the haplin result. Produce separate plots for child and mother,
# dump plots to files:
res <- haplin("data.dat", use.missing = T, maternal = T)
plot(res, separate.plots = T, filename = "Haplinres.png")

# Create haptable from the haplin result and plot the results in the table:
res <- haplin("data.dat", use.missing = T, maternal = T)
tab <- haptable(res)
plot(tab)

# Create haptables for the 10 first markers of a data file using haplinSlide.
# Create plots for each result:
res <- haplinSlide("data.dat", markers = 1:10, use.missing = T, maternal = T, table.output = T)
lapply(res, plot)

## End(Not run)</pre>
```

plotPValues

Plotting p-values for relative risks

## **Description**

This function plots p-values for the relative risks calculated by haplinSlide.

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## Usage

```
plotPValues(
  object,
  windows,
  which.p.val = "overall",
  plot.signif.only = FALSE,
  signif.thresh = 0.05,
  title,
  filename
)
```

# **Arguments**

object The haplinSlide results: list of haptable objects.

windows Numerical vector; if given, the plot will be restricted to only those.

which.p.val Character string specifying which p-values to choose for plotting: "overall" (de-

fault), "child", "child.double", "maternal", "maternal.double", "paternal". The last three options can be chosen only if haplinSlide was run with maternal =

TRUE or poo = TRUE.

plot.signif.only

Logical: whether to filter out the "non-significant" markers from the plot. De-

fault: FALSE, i.e., plot everything.

signif.thresh The threshold defining the significant p-values: if plot.signif.only == TRUE,

then only the markers with relative risk p-values lower than the threshold will

be kept for plotting. Default: 0.05.

title Optional character string for the title of the figure.

filename If the plot should be saved to the disk, give the name of the output file including

the file extension.

## **Details**

The output of haplinSlide can be very lengthy and not suitable for an overall plot of all the relative risks (RR) on one figure. Therefore, it's advised to first plot only the p-values for each window (user can choose which p-values to plot - see parameter which.p.val), and only then plot the RRs for specific windows, for which the p-values are significant.

#### Value

Invisibly returns the table with only the plotted p-values.

pQQ

pQQ

QQ-plot with confidence intervals for a vector of p-values

# **Description**

Produces a QQ-plot of p-values. The x-axis is -log10 of the expected p-values (under a null hypothesis of no effects), the y-axis is -log10 values of the actual p-values. A (pointwise) confidence interval can be drawn, and names of snps/genes corresponding to the most significant ones can be added.

# Usage

```
pQQ(pvals, nlabs = 6, conf = 0.95, lim, mark = 0.05, ...)
```

# **Arguments**

pvals	A vector of p-values.
nlabs	The number of (most significant) p-values to be labeled using names(pvals).
conf	The confidence level of the pointwise confidence band. The default is 0.95. The confidence intervals are computed under the assumption of the p-values being drawn independently from a uniform [0,1] distribution. To leave out the confidence interval, set this to FALSE.
lim	A vector of length 2 giving the plot limits (on a $log10$ scal, for instance $c(0,4)$ ). Plot limits are computed automatically. However, if other plot limits are desireable, they can be set using this argument.
mark	By default, the $0.05$ significance level is marked by lines. Can be changed to a different value, or set to FALSE.
• • •	Other arguments passed on to the plotting function.

# **Details**

The pvals argument should be a vector of p-values to be plotted. If the vector has names corresponding to marker (snp) names, the plot will label some of the most significant points with the marker names.

## Value

No value is returned.

# Author(s)

Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health <hakon.gjessing@uib.no> print.haplin 61

## References

Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396.

Web Site: https://haplin.bitbucket.io

print.haplin

Print a haplin object

## **Description**

Print basic information about a haplin object

## Usage

```
## S3 method for class 'haplin' print(x, ...)
```

# Arguments

x A haplin object, i.e. the result of running haplin.

... Other arguments, passed on to print.

## Note

Further information is found on the web page

## Author(s)

Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health <hakon.gjessing@uib.no>

## References

Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396.

```
Web Site: https://haplin.bitbucket.io
```

## See Also

haplin

```
print.summary.haplin Print the summary of a haplin object
```

# Description

Print the result of applying summary to a haplin object

# Usage

```
## S3 method for class 'summary.haplin'
print(x, digits, ...)
```

# **Arguments**

x A haplin object, i.e. the result of running haplin.

digits The number of digits to be used in the printout. Defaults to 3.

... Other arguments (ignored).

## Note

Further information is found on the web page

# Author(s)

Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health <a href="hakon.gjessing@uib.no">hakon.gjessing@uib.no</a>

## References

Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396.

```
Web Site: https://haplin.bitbucket.io
```

# See Also

haplin

rbindFiles 63

## **Examples**

```
## Not run:

# Standard summary:
res <- haplin("data.dat", use.missing = T, maternal = T)
summary(res)

# Increase number of digits in printout
print(summary(res), digits = 8)

## End(Not run)</pre>
```

rbindFiles

Combine a sequence of files by rows

# Description

Takes a sequence of files and combines them by rows, without reading the full files into memory. This is especially useful when dealing with large datasets, where the reading of entire files may be time consuming and require a large amount of memory.

# Usage

```
rbindFiles(infiles, outfile, col.sep, header = FALSE, ask = TRUE,
verbose = FALSE, add.file.number = FALSE, blank.lines.skip = FALSE)
```

# Arguments

infiles	A character vector of names (and paths) of the files to combine.
outfile	A character string giving the name of the modified file. The name of the file is relative to the current working directory, unless the file name contains a definite path.
col.sep	Specifies the separator used to split the columns in the files. To split at all types of spaces or blank characters, set col.sep = "[[:space:]]" or col.sep = "[[:blank:]]".
header	A logical variable which indicates if the first line in each file contains the names of the variables. If "TRUE", outfile will display this header in its first row, assuming the headers for each file are identical. Equals FALSE by default, i.e. no headers assumed.
ask	Logical. Default is "TRUE". If set to "FALSE", an already existing outfile will be overwritten without asking.
verbose	Logical. Default is "TRUE", which means that the line number is displayed for each iteration, i.e. each combined line.

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```
add.file.number
```

A logical variable which equals "FALSE" by deafult. If "TRUE", an extra first column will be added to the outfile, consisting of the file numbers for each line.

blank.lines.skip

Logical. If "TRUE" (default), lineByLine ignores blank lines in the input.

## **Details**

The function rbind combines R objects by rows. However, reading large data files may require a large amount of memory and be extremely time consuming. rbindFiles avoids reading the full files into memory. It reads the files line by line, possibly modifies each line, then writes to outfile. If however, header, verbose, add.file.number and blank.lines.skip are all set to "FALSE" (their default values), the files are appended directly, thus evading line-by-line modifications. In the case where infiles contains only one file and no output or modifications are requested (verbose, add.file.number and blank.lines.skip equal "FALSE"), an identical copy of this file is made.

#### Value

There is no useful output; the objective of rbindFiles is to produce outfile.

## Note

Combining the files by reading each file line by line is less time efficient than appending the files directly. For this reason, if header = FALSE, changing the values of the logical variables verbose, add.file.number and blank.lines.skip from "FALSE" to "TRUE" should not be done unless absolutely necessary.

## Author(s)

Miriam Gjerdevik, with Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health <hakon.gjessing@uib.no>

#### References

```
Web Site: https://haplin.bitbucket.io
```

#### See Also

```
cbindFiles, lineByLine
```

# Examples

```
## Not run:
# Combines the three infiles, by rows
rbindFiles(file.names = c("myfile1.txt", "myfile2.txt", "myfile3.txt"),
```

showGen 65

```
outfile = "myfile_combined_by_rows.txt", col.sep = " ", header = TRUE, verbose = TRUE)
## End(Not run)
```

showGen

Display chosen genotypes

# Description

This is a help function to extract genotypes from an object read in with genDataRead (or loaded with genDataLoad).

# Usage

```
showGen(data.in, design = "triad", n = 5, from, to, sex, markers = 1:5)
```

# Arguments

data.in	The data read in by genDataRead.	
design	The design used in the study - choose from:	
	<ul> <li><i>triad</i> - data includes genotypes of mother, father and child;</li> <li><i>cc</i> - classical case-control;</li> </ul>	
	• <i>cc.triad</i> - hybrid design: triads with cases and controls;	
	•	
n	Number of rows to display or "all" (default: 5).	
from	From which row to display (optional, default: from the first).	
to	To which row to display (optional).	
sex If the sex column is part of the phenotypic information, the user car based on one of the categories used in this column (optional); NB: this combine with the 'to' and 'from' arguments.		
markers	A vector specifying which markers to display or "all" (default: first 5); NB: the user can specify the markers by numbers or by their names.	

# Value

A table with genotypes extracted from 'data.in'.

66 showSNPnames

showPheno	Display phenotype part of data	

## **Description**

This is a help function to extract phenotypic (and covariate) data from an object read in with gen-DataRead (or loaded with genDataLoad).

# Usage

```
showPheno(data.in, n = 5, from, to, sex)
```

# Arguments

data.in	The data read in by genDataRead).
n	Number of rows to display or "all" (default: 5).
from	From which row to display (optional, default: from the first).
to	To which row to display (optional).
sex If the sex column is part of the phenotypic information, the user can cobased on one of the categories used in this column (optional); NB: this do combine with the 'to' and 'from' arguments.	

## Value

A table with phenotypic and covariate data (if any) extracted from 'data.in'.

# Description

This is a help function to extract marker names from an object generated by genDataRead, genDataLoad, genDataGetPart or genDataPreprocess.

# Usage

```
showSNPnames(data.in, n = 5, from, to)
```

# **Arguments**

data.in	The data as outputted by genDataRead, genDataLoad, genDataGetPart or gen-	
	DataPreprocess.	
n	Number of names to display or "all" (default: 5).	
from	From which marker to display (optional, default: from the first).	
to	o To which marker to display (optional).	

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## Value

A vector with marker names, as read in from map.file or generated dummy names.

snpPos

Find the column numbers of SNP identifiers/SNP numbers in a ped file

# **Description**

Gives the column numbers of SNP identifiers or SNP numbers in a standard ped file, calculated from the SNP's positions in the corresponding map file. The column numbers are sorted in the same order as snp.select. These positions may be useful when extracting a selection of SNPs from a ped file.

# Usage

```
snpPos(snp.select, map.file, blank.lines.skip = TRUE)
```

# **Arguments**

snp. select A character vector of the SNP identifiers (RS codes) or a numeric vector of the

SNP numbers.

map.file A character string giving the name and path of the standard map file to be used.

See Details for a description of the standard map format.

blank.lines.skip

Logical. If "TRUE" (default), snpPos ignores blank lines in map.file.

#### **Details**

To extract certain SNPs from a standard ped file, one has to know their positions in the ped file. This can be obtained from the corresponding map file.

The map file should look something like this:

Chromosome	SNP-identifier	Base-pair-position
1	RS9629043	554636
1	RS12565286	711153
1	RS12138618	740098

Alternatively, the map file could contain four columns. The column values should then be: Chromosome, SNP-identifier, Genetic-distance, Base-pair-position.

A header must be added to the map file if this does not already exist.

The format of the corresponding ped file should be something like this:

68 snpPos

```
1104 1104-1 1104-2 1104-3 1
1104 1104-2
                0
                        0
                          1
                                        2
1104 1104-3
                 0
                        0
                         2
                                        0
                                        2
1105 1105-1 1105-2 1105-3 2 2
                                1
                                   1
                                     2
1105 1105-2
                 0
                        0
                          1
                             1
                                1
                                   1
                                        2
1105 1105-3
                 0
                        0 2 1 1 1 3 2
```

The column values are: Family id, Individual id, Father's id, Mother's id, Sex (1 = male, 2 = female, alternatively: 1 = male, 0 = female), and Case-control status (1 = controls, 2 = cases, alternatively: 0 = controls, 1 = cases).

Column 7 and onwards contain the genotype data, with alleles in separate columns. A "0" is used to denote missing data.

#### Value

A vector of the column numbers of the SNP identifiers/SNP numbers in the ped file, sorted in the same order as given in snp.select.

## Note

The function does not check if the map file is formatted correctly or if the map and ped file have the same number of SNPs. The corresponding positions of the SNPs in the ped file may not be correct if the ped file has a different format from the given example.

## Author(s)

Miriam Gjerdevik, with Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health <hakon.gjessing@uib.no>

## References

```
Web Site: https://haplin.bitbucket.io
```

#### See Also

```
convertPed, lineByLine
```

#### **Examples**

```
## Not run:
# Find the column numbers of the SNP identifiers "RS9629043" and "RS12565286" in
# a standard ped file
snpPos(snp.select = c("RS9629043", "RS12565286"), map.file = "mygwas.map")
```

snpPower 69

## End(Not run)

snpPower Power calculations for a single SNP	Power calculations for a single SNP	

# Description

Calculates power for a single SNP. Allows for power computations of several combinations simultaneously.

# Usage

```
snpPower(cases, controls, RR, MAF, alpha = 0.05)
```

# **Arguments**

_	
cases	A list of the number of case families. Each element contains the number of families of a specified family design. The possible family designs, i.e. the possible names of the elements, are "mfc" (full triad), "mc" (mother-child dyad), "fc" (father-child dyad) or "c" (a single case child).
controls	A list of the number of control families. Each element contains the number of families of a specified family design. The possible family designs are "mfc" (full triad), "mc" (mother-child-dyad), "fc" (father-child dyad), "mf" (mother-father dyad), "c" (a single control child), "m" (a single control mother) or "f" (a single control father).
RR	A numeric vector of the relative risks (the effect sizes of interest).
MAF	A numeric vector of the minor allele frequencies.

## **Details**

alpha

snpPower computes power for a single SNP by counting the number of "real" case alleles, "real" control alleles and pseudo-control alleles. The pseudo-control alleles are the non-transmitted alleles, possibly from both case families and control families. It assumes a multiplicative dose-response model. snpPower uses the asymptotic normal approximation for the natural logarithm of the odds ratio for calculating power (the relative risks and odds ratios are used interchangeably due to the "rare disease assumption").

A numeric vector of the Type I Errors. Equals 0.05 by default.

snpPower allows for power calculations for mixtures of the possible case family designs and control family designs. The argument cases could, for example, consist of a combination of 1000 full case triads (family design "mfc") and 500 single case children (family design "c"). It is also feasible to compute power for several combinations of the input variables simultaneously. See Examples for further details.

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#### Value

snpPower returns a data frame containing the combinations of input variables and the corresponding power calculations.

## Author(s)

```
Miriam Gjerdevik,
with Hakon K. Gjessing
Professor of Biostatistics
Division of Epidemiology
Norwegian Institute of Public Health
```

<hakon.gjessing@uib.no>

#### References

Skare O, Jugessur A, Lie RT, Wilcox AJ, Murray JC, Lunde A, Nguyen TT, Gjessing HK. Application of a novel hybrid study design to explore gene-environment interactions in orofacial clefts. Annals of Human Genetics (2012) 76, pp. 221-236.

```
Web Site: https://haplin.bitbucket.io
```

#### See Also

haplin, snpSampleSize, hapRun, hapPower, hapPowerAsymp

#### **Examples**

```
## Compute power for a single SNP,
## for the combination of 1000 case triads, RR = 1.2, MAF = 0.1 and alpha = 0.05
snpPower(cases = list(mfc = 1000), controls = list(mfc = 0), RR = 1.2, MAF = 0.1)

## Compute power for a single SNP,
## for the combination of 1000 case triads and 500 single case children (altogether),
## 5000 control triads, RR = 1.1, MAF = 0.1 and alpha = 0.05
snpPower(cases = list(mfc = 1000, c = 500), controls = list(mfc=5000),
RR = 1.1, MAF = 0.1, alpha = 0.05)

## Compute power for a single SNP,
## for the combination of 500 case triads, 10000 control triads,
## relative risk of 1.2 and minor allele frequency of 0.1,
## and also for the combination of 1000 case triads, 10000 control triads,
## relative risk of 1.1 and minor allele frequency of 0.1
snpPower(cases = list(mfc = c(500, 1000)), controls = list(mfc = 10000),
RR = c(1.2, 1.1), MAF = 0.1)
```

snpSampleSize 71

	snpSampleSize	Sample size calculations for a single SNP	
--	---------------	---	--

#### **Description**

Sample size calculations for a single SNP. Allows for sample size calculations of several combinations simultaneously.

# Usage

```
snpSampleSize(fam.cases, fam.controls, fraction = 0.5,
RR, MAF, alpha = 0.05, power = 0.80)
```

## **Arguments**

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fam.cases	A character vector of the case family design. The possible family designs are "mfc" (full triad), "mc" (mother-child dyad), "fc" (father-child dyad) and "c" (a single case child).
fam.controls	A character vector of the control family design. The possible family designs are "mfc" (full triad), "mc" (mother-child-dyad), "fc" (father-child dyad), "mf" (mother-father dyad), "c" (a single control child), "m" (a single control mother), "f" (a single control father) or "no_controls" (no control families).
fraction	A numeric vector of the proportion of case families. Equals 0.5 by default, i.e. there are as many case families as control families. If fam.controls equals "no_controls", fraction is automatically set to 1.
RR	A numeric vector of the relative risks (the effect sizes of interest).
MAF	A numeric vector of the minor allele frequencies.
alpha	A numeric vector of the Type I Errors. Equals 0.05 by default.
power	A numeric vector of the desired probability of identifying a difference in the relative risks. Default is 0.80.

#### **Details**

snpSampleSize computes the number of case and control families required for a single SNP to attain the desired power. It assumes a multiplicative dose-response model. snpSampleSize calculates the fraction of case alleles corresponding to the given family designs and then uses the asymptotic normal approximation for the natural logarithm of the odds ratio for calculating the sample sizes (the relative risks and odds ratios are used interchangeably due to the "rare disease assumption").

snpSampleSize allows for sample size calculations of several combinations of the input variables at once. The Examples section provides further details.

#### Value

snpSampleSize returns a data frame containing the combinations of input variables and the corresponding sample size calculations.

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#### Author(s)

Miriam Gjerdevik, with Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health

<hakon.gjessing@uib.no>

## References

Skare O, Jugessur A, Lie RT, Wilcox AJ, Murray JC, Lunde A, Nguyen TT, Gjessing HK. Application of a novel hybrid study design to explore gene-environment interactions in orofacial clefts. Annals of Human Genetics (2012) 76, pp. 221-236.

Web Site: https://haplin.bitbucket.io

#### See Also

haplin, snpPower

#### **Examples**

```
## Compute sample sizes for a single SNP,
## when the specified case family design is a full triad, there are no control families,
## RR = 1.1, MAF = 0.1, alpha = 0.05 and power = 0.9
snpSampleSize(fam.cases = "mfc", fam.controls = "no_controls",
RR = 1.1, MAF = 0.1, alpha = 0.05, power = 0.9)

## Compute sample sizes for a single SNP,
## for the combination of case triads, control triads, fraction = 0.5, RR = 1.2,
## MAF = 0.1, alpha = 0.05 and power = 0.8, and also for the combination of case triads,
## control children, fraction = 0.5, RR = 1.2, MAF = 0.1, alpha = 0.05 and power = 0.9
snpSampleSize(fam.cases = "mfc", fam.controls = c("mfc", "c"),
RR = 1.2, MAF = 0.1, alpha = 0.05, power = c(0.8, 0.9))
```

suest

Compute a joint p-value for a list of haplin fits (usually from a sliding window approach), correcting for multiple testing.

# Description

The first argument to suest should be a list of haplin estimation results (from the same data file), usually the output from haplinSlide. suest produces as a result a joint overall p-value based on aggregating the individual p-values and then correcting for multiple testing. The correction is achieved by using the principle of "seemingly unrelated" estimation, taking into account the correlation between the individual estimation results.

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#### Usage

suest(reslist)

#### **Arguments**

reslist A list whose elements are different haplin runs on the same data file, typically

the output of haplinSlide.

#### **Details**

haplinSlide runs haplin on a series of overlapping windows of markers from the same data file, typically within the same gene. Since each run produces a separate overall p-value, suest computes a joint overall p-value for the gene (or region) that has been scanned. It corrects the overall p-value for multiple testing, also taking into account the fact that the sequence of estimates produced by haplinSlide will be dependent, both because they are computed on the same data set and also since the windows are overlapping (if the window length is larger than 1). If the suest estimation fails (which doesn't happen very often), a standard Bonferroni correction is used instead. Important: haplinSlide must be run with the option table.output = FALSE to provide suest with enough information.

#### Value

A list is returned, the most important elements of which are:

pval.obs The overall score p-values from each haplin run pval.obs.corr The joint p-value, corrected for multiple testing

bonferroni A logical, usually FALSE, which means the suest estimation went well. If

TRUE, it means that the suest estimation failed for some reason, and a standard

Bonferroni correction was used instead.

#### Note

Further information is found on the web page.

# Author(s)

Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health <hakon.gjessing@uib.no>

# References

Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396.

Web Site: https://haplin.bitbucket.io

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## See Also

```
haplin, haplinSlide
```

#### **Examples**

```
## Not run:
# (Almost) all standard haplin runs can be done with haplinSlide.
# Below is an illustration. See the haplin help page for more
# Analyzing the effect of fetal genes, including triads with missing data,
# using a multiplicative response model. When winlength = 1, separate
# markers are used. To make longer windows, winlength can be increased
# correspondingly:
result.1 <- haplinSlide("C:/work/data.dat", use.missing = T, response = "mult",
reference = "ref.cat", winlength = 1, table.output = F)
# Provide summary of separate results:
lapply(result.1, summary)
# Plot results:
par(ask = T)
lapply(result.1, plot)
# Compute an overall p-value for the scan, corrected for multiple testing
# and dependencies between windows:
suest(result.1)
## End(Not run)
```

summary.haplin

Summary of a haplin object

# Description

Provides detailed information about estimation results from a haplin object.

#### Usage

```
## S3 method for class 'haplin'
summary(object, reference, ...)
```

## **Arguments**

object A haplin object, i.e. the result of running haplin.

reference Same as reference argument in haplin. Note that when producing the sum-

mary, you can only choose "reciprocal", "population" or "ref.cat". You cannot use a numeric value to change the reference category, to do that haplin

must be run over again. (See the reference argument of haplin.)

. . . Other arguments (ignored).

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## Note

Further information is found on the web page

# Author(s)

Hakon K. Gjessing Professor of Biostatistics Division of Epidemiology Norwegian Institute of Public Health <hakon.gjessing@uib.no>

#### References

Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396.

```
Web Site: https://haplin.bitbucket.io
```

## See Also

haplin

# **Examples**

```
## Not run:

# Produce separate plots for child and mother, dump plots to files:
res <- haplin("data.dat", use.missing = T, maternal = T)
summary(res)

## End(Not run)</pre>
```

toDataFrame

Stack dataframes from haplinSlide into a single dataframe

# **Description**

When haplinSlide is run with the option table.output = T, the result is a list of haptables, i.e. tables with summary haplin results for each window haplinSlide is run on. toDataFrame stacks the separate dataframes into one large dataframe containing all results.

# Usage

```
toDataFrame(x, reduce = F)
```

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## Arguments

x The output from haplinSlide run with option table.output = TRUE.

reduce Reduce output to one line per marker

## **Details**

When haplinSlide is run with winlength = 1 on SNP markers, each table in the output has only two rows, and can be condensed to a single row. By setting the argument reduce to TRUE, toDataFrame reduces each table to one line and returns a dataframe with one line for each SNP. In more general situations, with multi-allelic markers or, more commonly, winlength set to 2 or more, each output table will typically have more than two rows and cannot be reduced, so reduce should be set to FALSE.

#### Value

The output is a dataframe. First column contains the marker names. Second column are row numbers, counted within each output table. The remaining columns are identical to the individual output columns, which are described in more detail in the help file for haptable.

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#### References

Gjessing HK and Lie RT. Case-parent triads: Estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Annals of Human Genetics (2006) 70, pp. 382-396.

Web Site: https://haplin.bitbucket.io

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